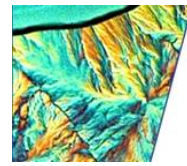
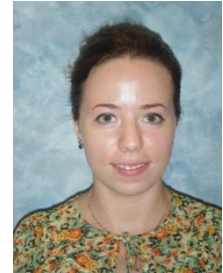


# Executive Committee



OSA

Therapeutic  
Laser Applications  
Technical Group



Chair: Elina A. Vitol

Advisor: David R. Busch, UT Southwestern Medical Center at Dallas

Committee Member: Yannis M. Paulus, Assistant Professor, University of Michigan

Committee Member: Guido Perrone, Associate Professor, Politecnico di Torino

Committee Member: Felix Fanjul-Velez, Associate Professor, University of Cantabria

Industry Liaison: Robbie Thomas, Photon Force, Ltd.

Social Media Liaison: Mariia Shutova, Texas A&M University

# Where to find information about the group



Browser address bar: [https://www.osa.org/en-us/get\\_involved/technical\\_groups/bmo/therapeutic](https://www.osa.org/en-us/get_involved/technical_groups/bmo/therapeutic)

Navigation: < Navigate OSA | Other OSA Sites | Not a Member? Join OSA | Login | Search OSA

Utility links: About OSA | Awards | Career | Directories | Video | Newsroom | Help

OSA The Optical Society | 100 Since 1916

Menu: Journals & Proceedings | Meetings & Exhibits | Explore Membership | Industry Programs | **Get Involved** | Foundation & Giving | Living History


Breadcrumbs: Home / Get Involved / Technical Groups / Bio-Medical Optics

## Therapeutic Laser Applications (BA)

### Get Involved

- Diversity & Inclusion
- Public Policy
- Chapters and Sections Map
- Technical Groups
  - Bio-Medical Optics
    - Microscopy and Optical Coherence Tomography (BM)
    - Molecular Probes and Nanobio-Optics (BP)
    - Optical Biosensors (BB)
    - Optical Trapping and Manipulation in Molecular and Cellular Biology (BT)
    - Therapeutic Laser Applications (BA)**
    - Tissue Imaging and Spectroscopy (Bs)
    - Photobiomodulation (BL)
  - Fabrication, Design & Instrumentation
  - Information Acquisition, Processing & Display

### Therapeutic Laser Applications



This group focuses on the use of lasers in surgery or in other treatments of disease. This includes the use of lasers as surgical tools for tissue cutting, welding, and coagulation, as well as the use of optics to initiate cell-damaging photochemical reactions for the treatment of diseases such as cancer. In addition, optics, spectroscopy, and imaging provide unique tools that may allow real-time diagnostics of the efficacy of clinical procedures. For many of these applications, the development of optical tools for appropriate light delivery, especially for fiber-based or endoscopic delivery to tissues that are not directly accessible, is critical. In addition, this group emphasizes basic science studies of the mechanisms by which light can affect tissue in adverse or therapeutic ways.

### Upcoming Technical Group Webinars

#### Photoacoustic Imaging of the Eye

**Hosted By:** Therapeutic Laser Applications Technical Group

24 October 2019, 10:00 AM - 11:00 AM

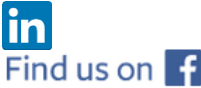
[Register Now](#)


### Announcements

If you are a member of the Therapeutic Laser Applications Technical Group and have ideas for activities and initiatives to help engage this community, please [share them with the chair, Elina Vitol](#).

View [OSA Technical Group webinars](#) on-demand at any time or register for any of our upcoming webinars [online](#). Each webinar is an hour long and features a technical presentation on a topic selected by your OSA Technical Groups.

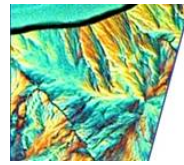
### Join our Online Community



Find us on 

Work in Optics



# We want you to join us!



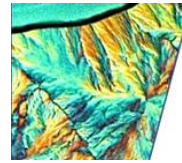
# OSA

Therapeutic  
Laser Applications  
Technical Group



- Select **Therapeutic Laser Applications** as one of 5 technical groups of interest at your OSA membership account page
- Attend our networking events, webinars and poster sessions
- Join us on LinkedIn and Facebook to keep in touch  
- Look out for emails from the committee about group activities
- Interested in presenting your research? Have ideas for technical group events?  
Want to reach out to your fellow group members?
  - Contact us at [elina.vitol@gmail.com](mailto:elina.vitol@gmail.com) or [TGactivities@osa.org](mailto:TGactivities@osa.org)

# Upcoming webinars



OSA

Therapeutic  
Laser Applications  
Technical Group



1 November 2019, 12 pm EST

## ***Recent advances in tissue biomechanics using Dynamic Optical Coherence Elastography***

Kirill Larin, PhD, University of Houston

Registration is  
now open!

21 January 2020, 11am EST\*

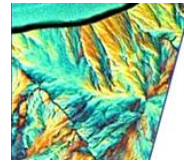
## ***Thermomechanical effect of infrared laser for cartilage regeneration***

Yulia M. Alexandrovskaya, PhD

Institute of Photon Technologies, Federal Scientific Research  
Centre “Crystallography and Photonics” of the Russian  
Academy of Sciences (RAS)

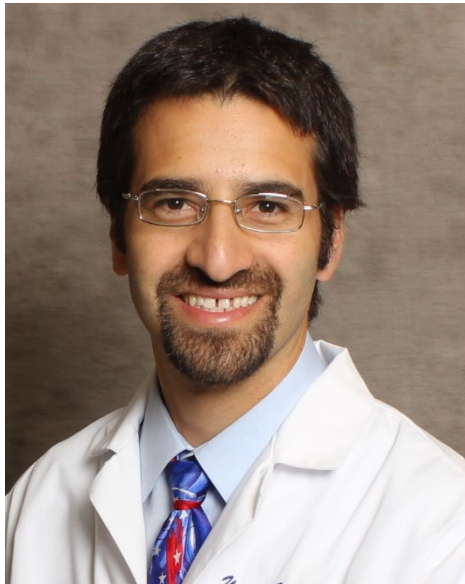
Registration will  
open in December

# Welcome to today's webinar!



# OSA

Therapeutic  
Laser Applications  
Technical Group



## PHOTOACOUSTIC IMAGING OF THE EYE

Yannis M. Paulus, M.D., F.A.C.S.

Assistant Professor, University of Michigan  
Department of Ophthalmology and Visual Sciences, and  
Department of Biomedical Engineering

OCTOBER 24, 2019

# Photoacoustic Imaging of the Eye

**Yannis M. Paulus, M.D., F.A.C.S.**

Assistant Professor

Department of Ophthalmology & Visual Sciences

Department of Biomedical Engineering

University of Michigan Kellogg Eye Center

OSA Therapeutic Laser Applications Technical Group

Webinar

October 24, 2019



# Disclosures

- Inventor University of Michigan patents
  - Method and Apparatus for Removing Microvessels
  - RetinaScope Apparatus
  - Photomediated Ultrasound Therapy Method and Apparatus
  - Purely Organic Phosphorescent Nanoparticles for In Vivo Oxygen Sensing
  - Laser Ultrasound Body Sculpting
  - Multi-modal imaging for cell tracking
- Co-Founder companies PhotoSonoX LLC, OcuBell.
- CEO of PhotoSonoX LLC
- Consultant for Oraya Therapeutics, Quattro Consulting, Sonify Biosciences, Allergan Regional Advisory Board, Putnam Associated Consulting, Roda Consulting, ENDRA Life Sciences, MediBeacon Inc
- Will discuss several preclinical systems not approved by the FDA

# Imaging is critical

- We can understand and diagnose what we can see
- Early disease detection
- Improved diagnosis
- Improved disease monitoring
- Better patient outcomes
- Precision medicine tailored to each patient's molecular profile
- Improved understanding of pathophysiology
  - Change name: Central Serous Retinopathy to Central Serous Chorioretinopathy
  - Acute Posterior Multifocal Placoid Pigment Epitheliopathy (APMPPE) to Acute Multifocal Placoid Choroidopathy (AMP-C)<sup>1,2</sup>
- **Retina is very unique.** The eye is optically transparent, so we can directly visualize neurons and microvasculature with high resolution optical imaging.

<sup>1</sup>Zhang AY, Han IC, Goldberg MF. Renaming of Acute Posterior Multifocal Placoid Pigment Epitheliopathy (APMPPE) to Acute Multifocal Placoid Choroidopathy (AMP-C). JAMA Ophthalmol. 2017 Mar 1;135(3):185

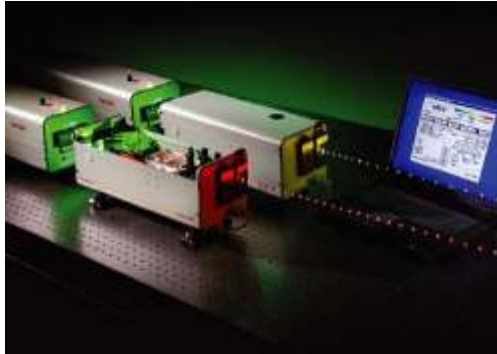
<sup>2</sup>Jampol LM, Goldstein DA, Fawzi AA. Keeping the Name of Acute Posterior Multifocal Placoid Pigment Epitheliopathy. JAMA Ophthalmol. 2017 Mar 1;135(3):186.



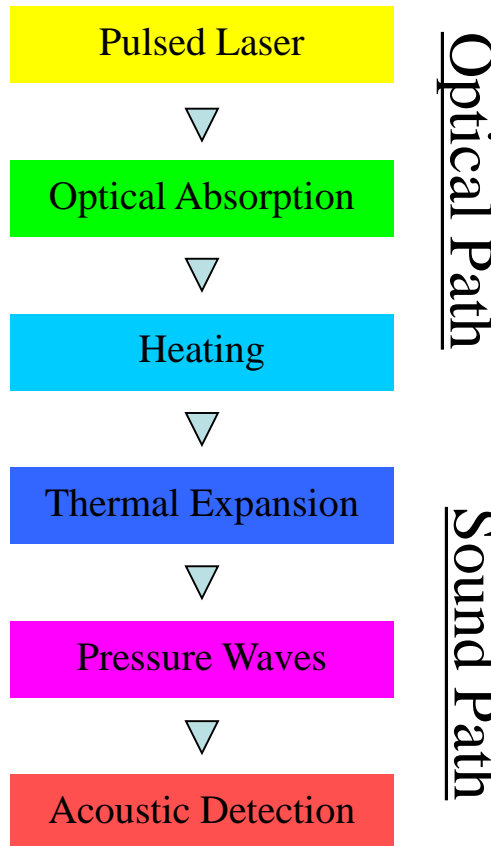
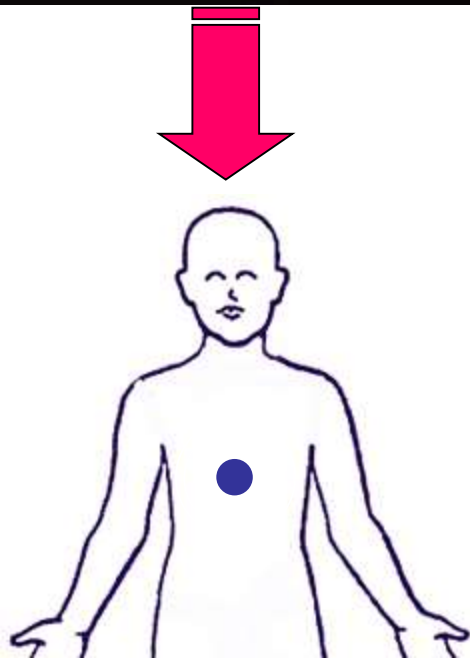
# Retinal Imaging today

- Fundus photography 1853
- Ultrasonography (A & B) 1956
- Fluorescein angiography 1961
- Indocyanine green angiography 1972
- Scanning laser ophthalmoscopy 1981
- Optical Coherence Tomography 1991
  - Time domain
  - Spectral domain
  - OCTA
  - Swept Source
  - Intra-operative
  - Hand-held/peds
  - Different wavelength
  - Doppler OCT
- Ultra-wide field imaging
- Fundus autofluorescence (FAF)
- Retinal oximetry
- Fluorescent lifetime imaging ophthalmoscopy (FLIO)
- Real-time image-guidance of laser photocoagulation
- Adaptive Optics
- Handheld/Smartphone-based fundus imaging
- Automated interpretation/ deep learning
- **Photoacoustic Imaging**
- **Multimodal Imaging**
- **Molecular Imaging**

# Photoacoustic Imaging



Photoacoustic effect: conversion light to sound.  
Optoacoustic = thermoacoustic



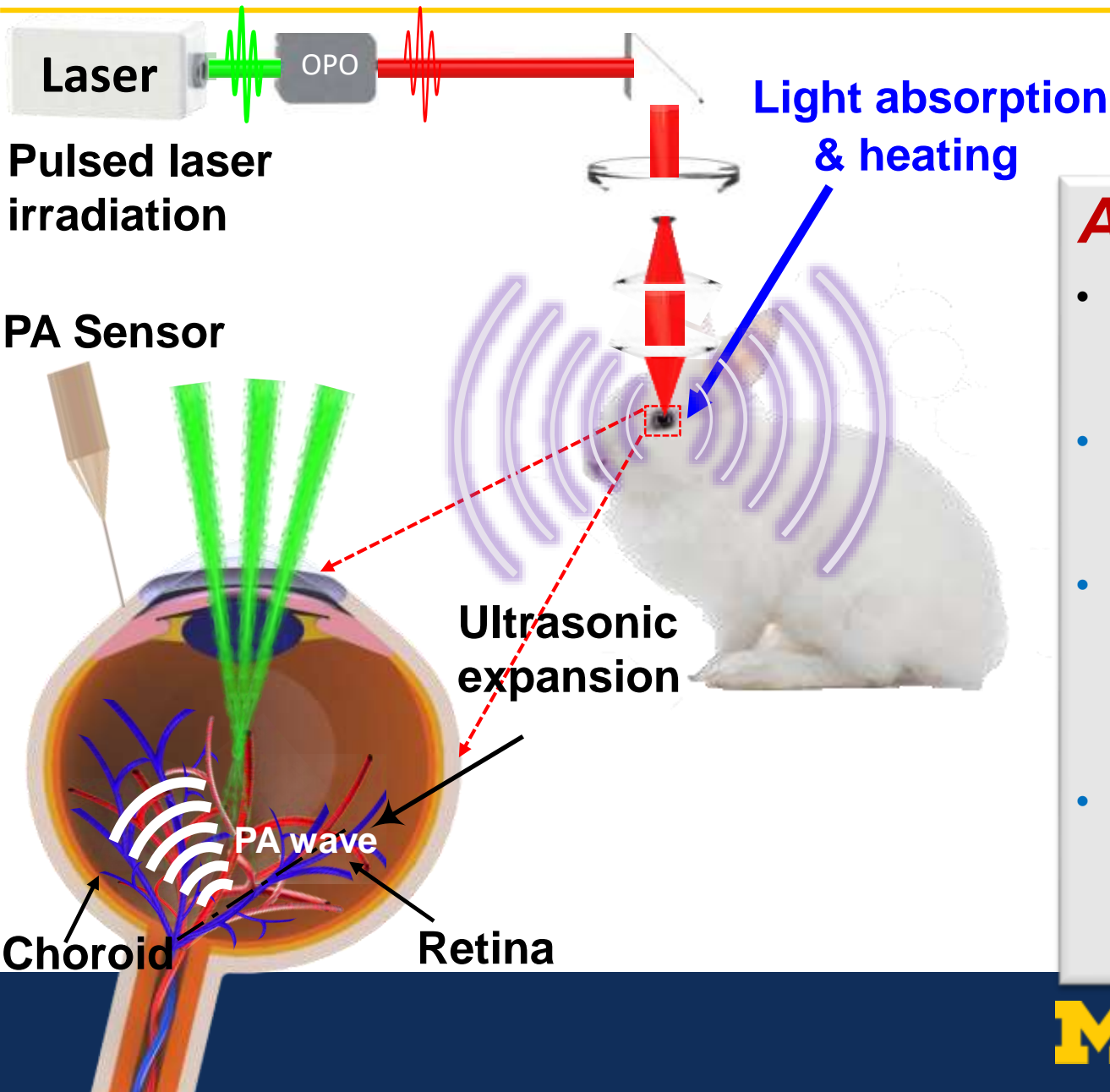
## Advantages

1. Deep Penetration
2. High Resolution
3. Speckle-Free

1880: Alexander G. Bell describes photoacoustic princ.

Need  $t < 20$  nsec. Most  $\sim 5$  nsec

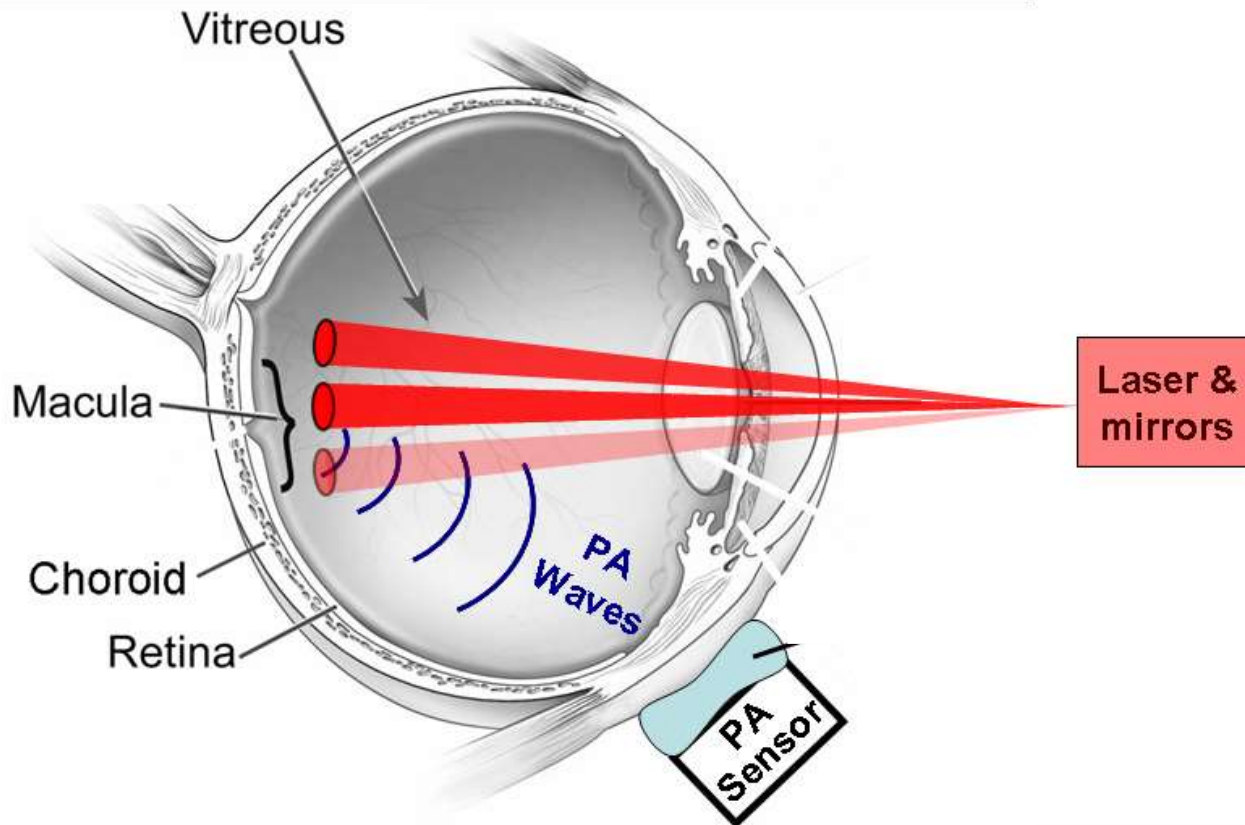
# Photoacoustic Ocular Imaging



## ***Advantages:***

- Non-invasive, and **high contrast**
- **Structural:** 3D vessel structure
- **Functional:** hemoglobin concentration, oxygen saturation, blood flow
- **Molecular information with contrast agents:** integrin, growth factor

# Photoacoustic Ocular Imaging



## Endogenous absorbers:

Hemoglobin

Oxy and deoxy

Melanin

DNA/RNA

Lipid

H<sub>2</sub>O

## Exogenous contrast:

Methylene blue

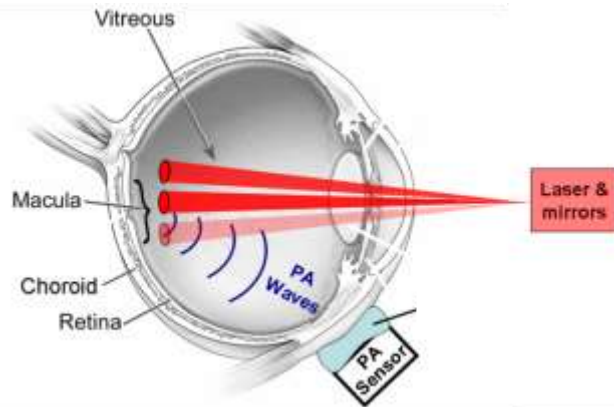
Indocyanine green

Organic nanoparticles

Gold nanorods

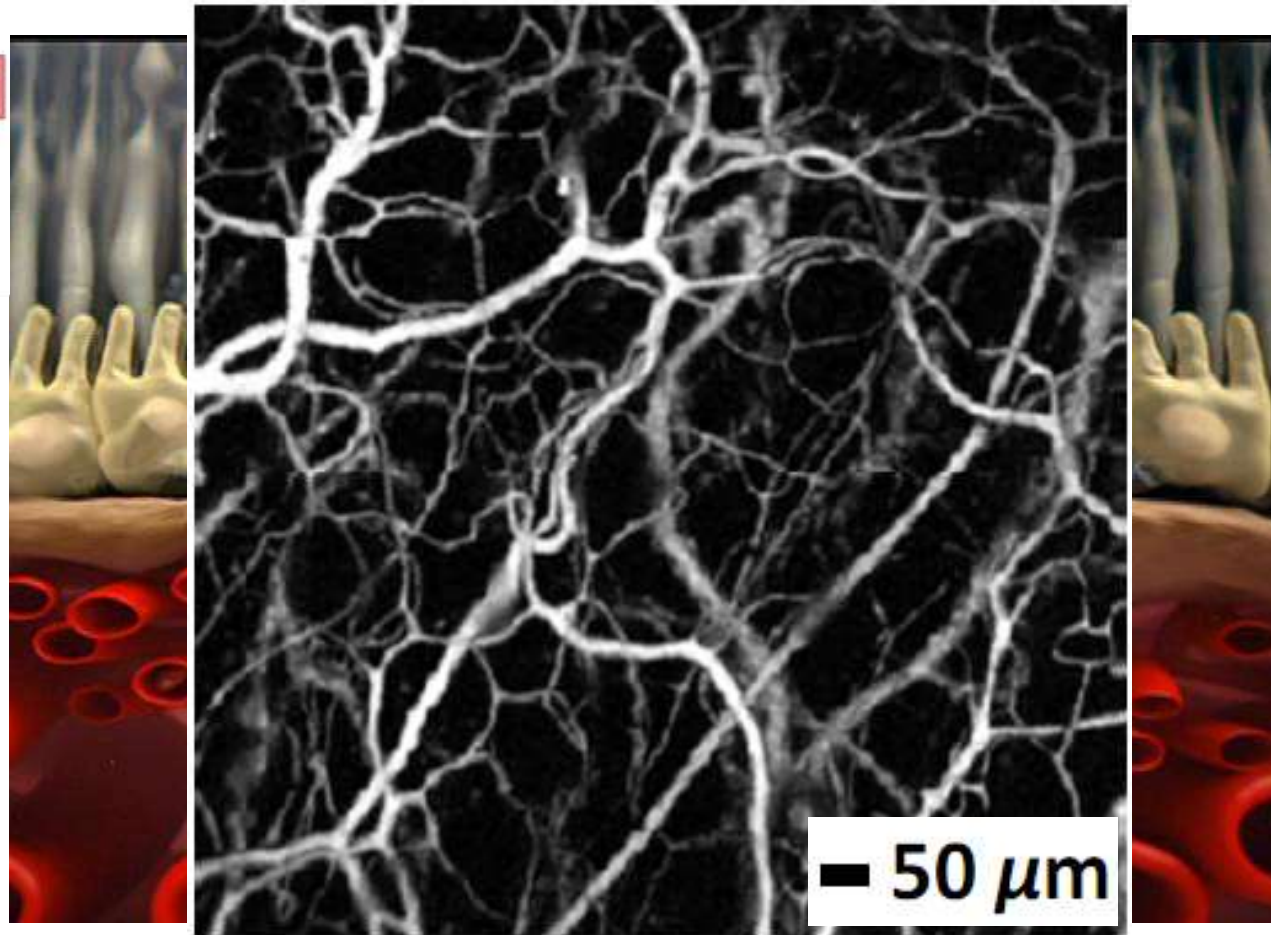
Microbubbles

# Photoacoustic Ocular Imaging



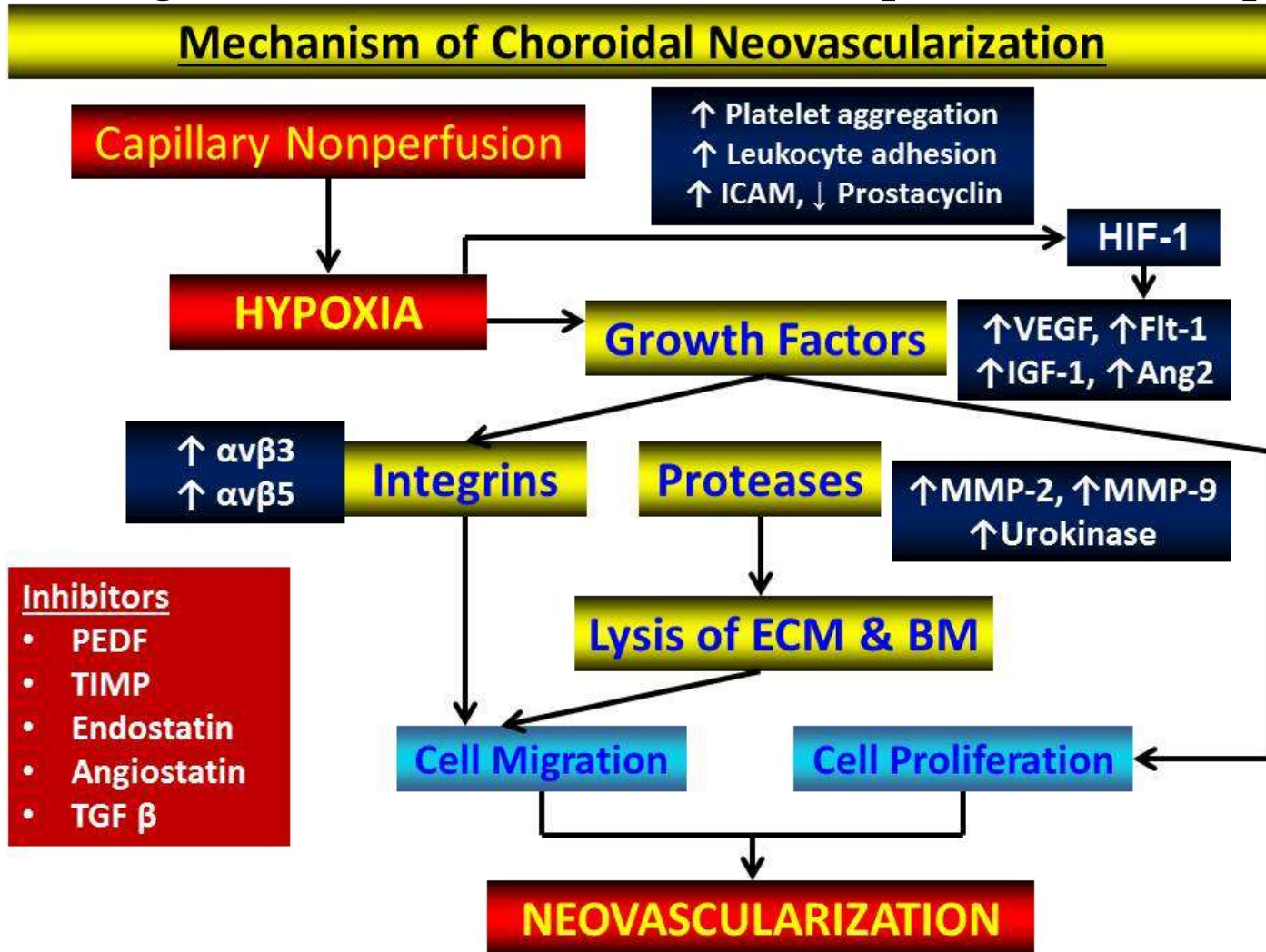
3  $\mu\text{m}$  resolution:

- ✓ OCT image
- ✓ Blood distribution (angiogenesis)
- ✓ Oxygen saturation levels (ischemia)
- ✓ Tissue blood content
- ✓ ICG Photoacoustic Angiography



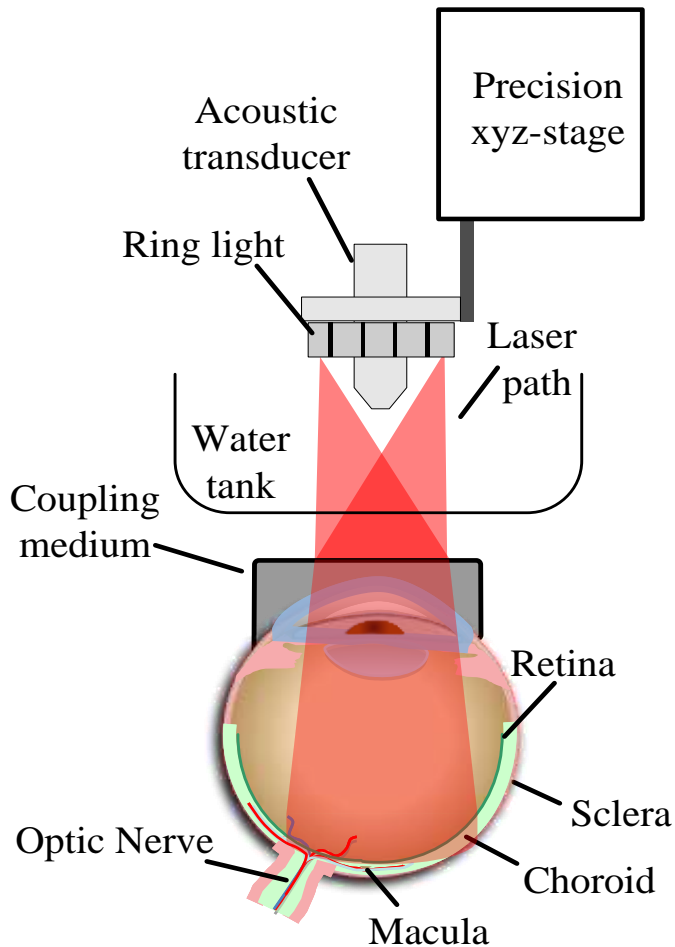
# Molecular Imaging

Anatomic changes are the end result from complex molecular pathways

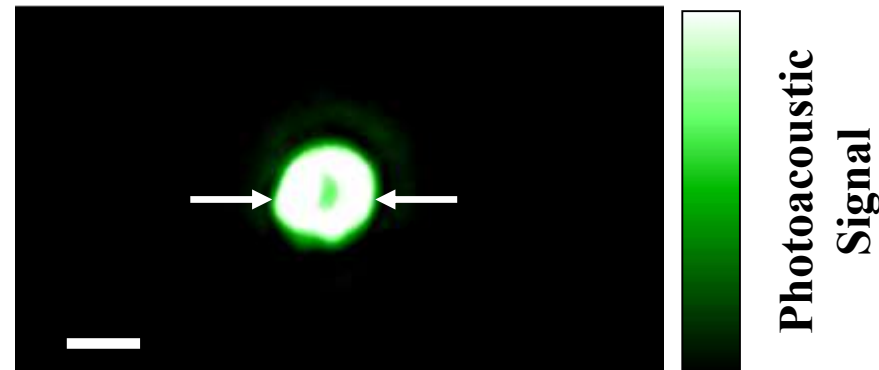
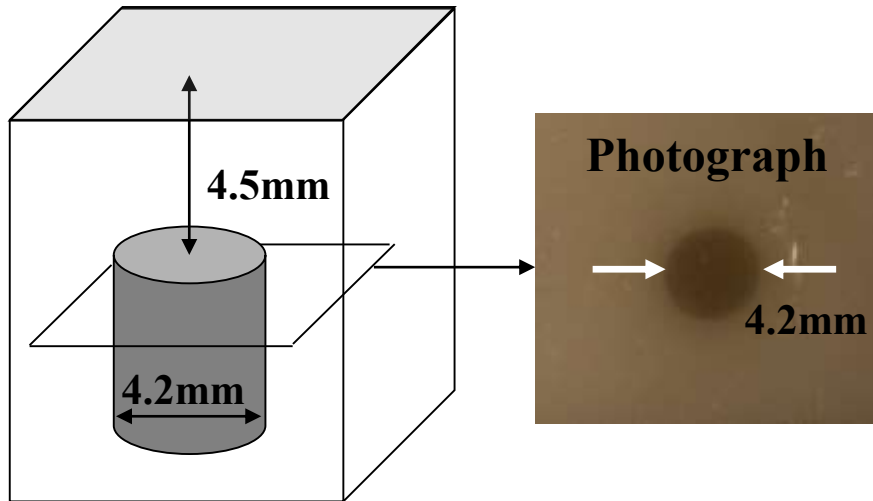
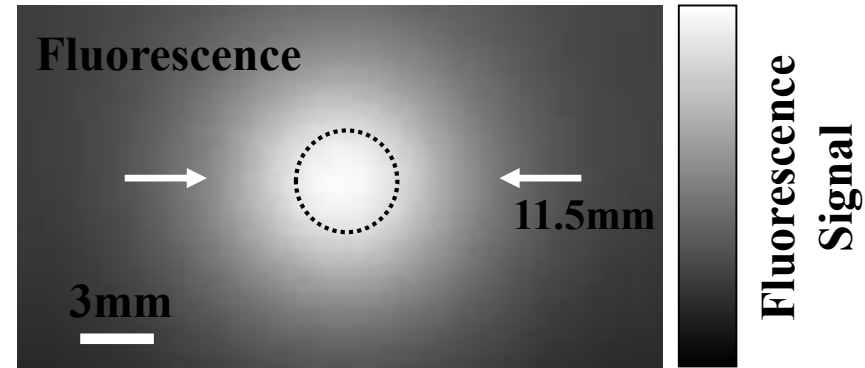
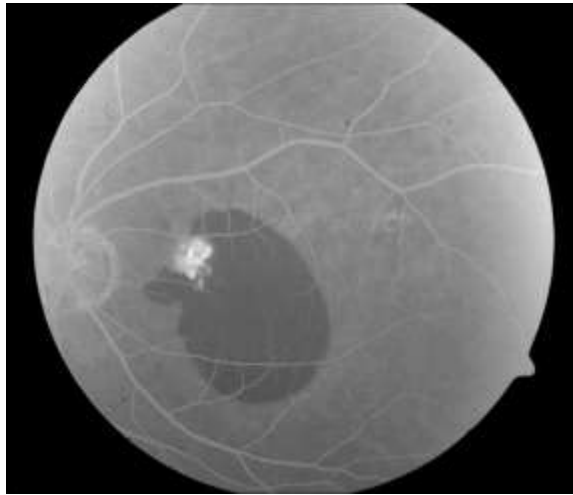


# Photoacoustic Imaging System

- Tunable, pulsed Nd:YAG laser
- Rep rate 10 Hz, pulse width 5 ns
- Fiber optic ring light
- Retinal laser density  $0.5 \text{ mJ/cm}^2$ , below ANSI limit
- US transducers 15 & 25 MHz acquire pulse-echo + PA signals
- Axial resolution: 83 & 50  $\mu\text{m}$
- Lateral resolution: 200 & 240  $\mu\text{m}$



# ICG Photoacoustic Imaging



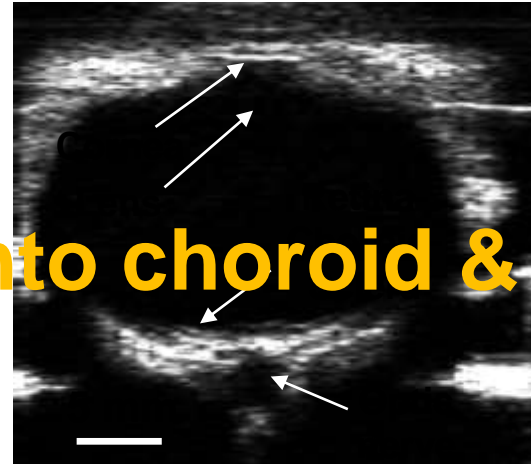
Tissue mimicking phantom filled with ICG



# Results: Enucleated Pig Eye

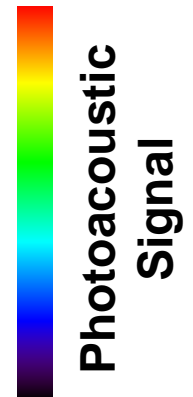
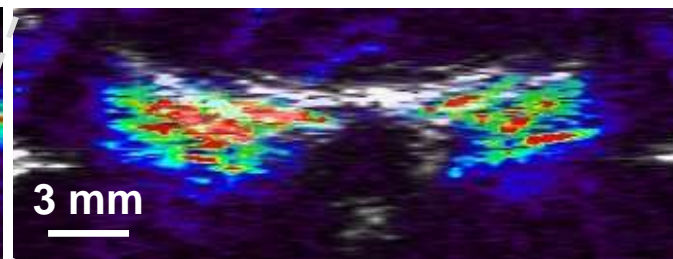
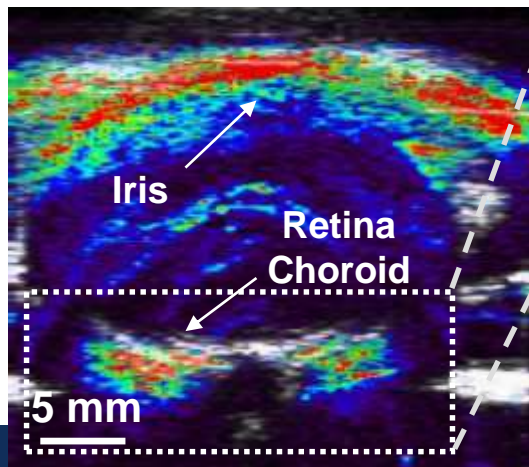
Pig's Eye

Ultrasound



➤ Deep penetration into choroid & sclera

Photoacoustic



Pig eye: 22 mm

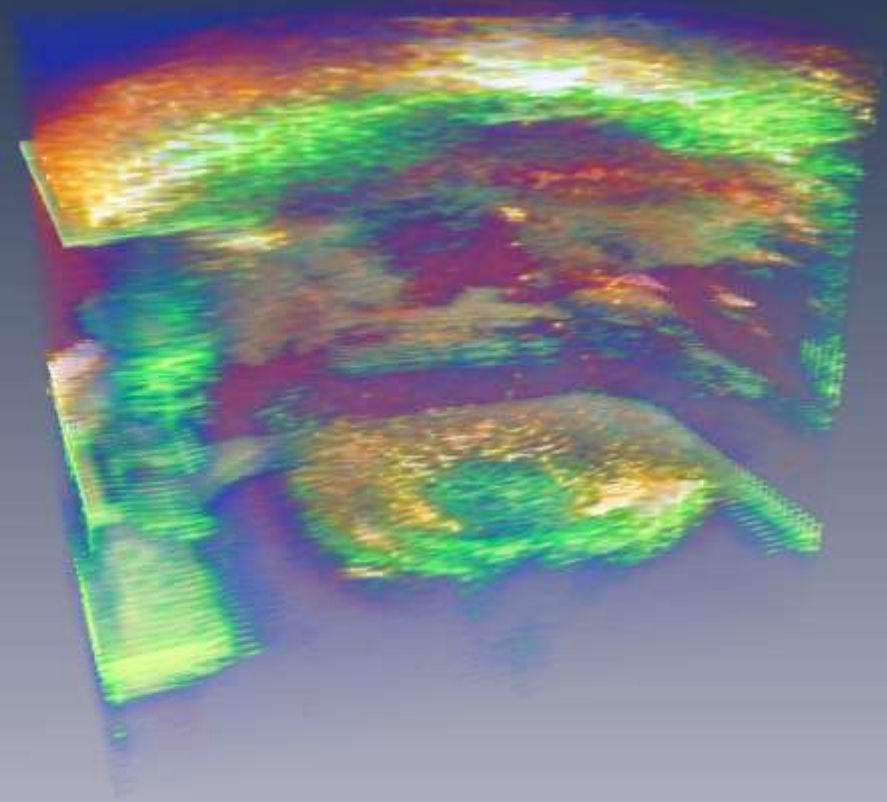
Human eye: 24 mm

# Enucleated Pig Eye



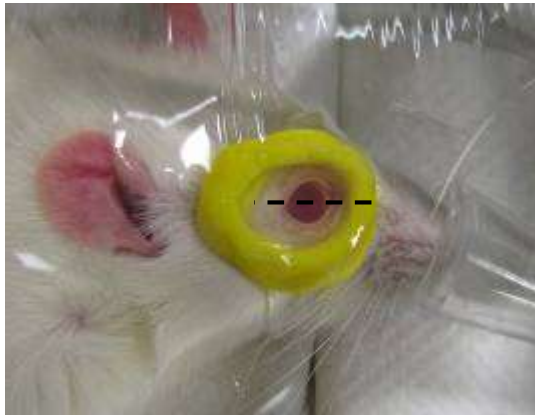
**Reconstructed 3D:  
Amira, Visage Imaging  
63 A-line scans  
250  $\mu\text{m}$  apart  
8 averages  
60 sec acquisition**

**Ultrasound + Photoacoustic**

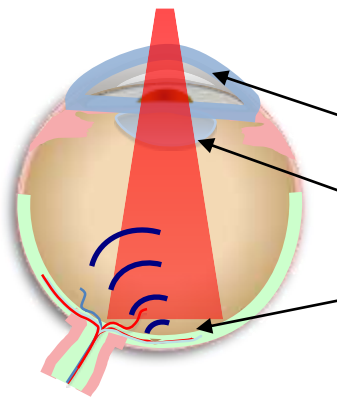


# Live Rat Eye

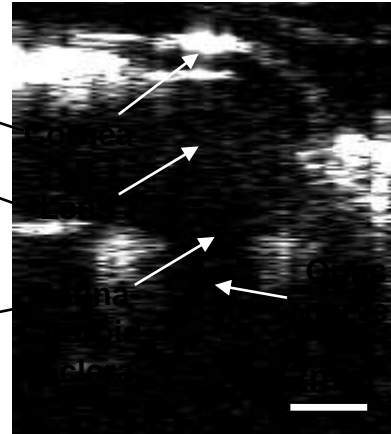
Rat's eye



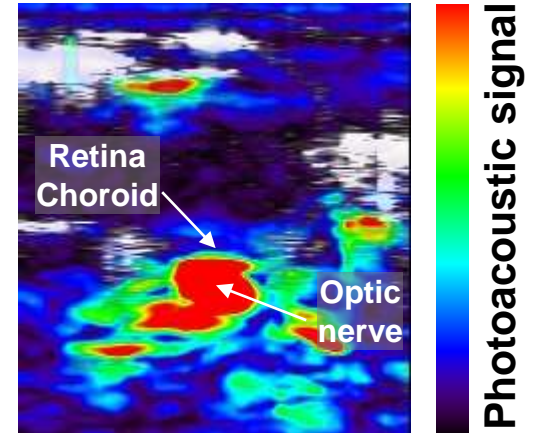
Eye Diagram



Ultrasound



Photoacoustic

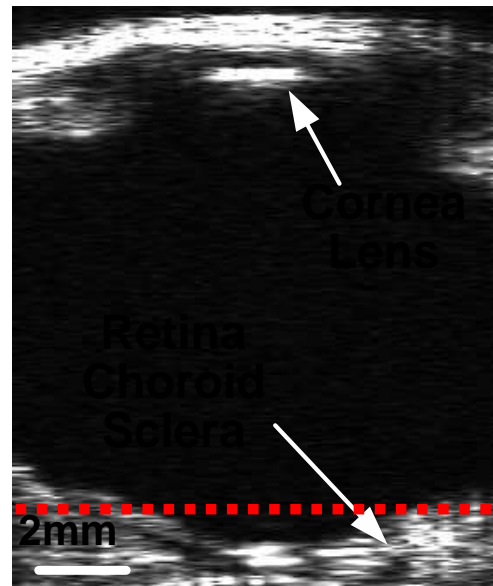


# Live Rabbit Eye

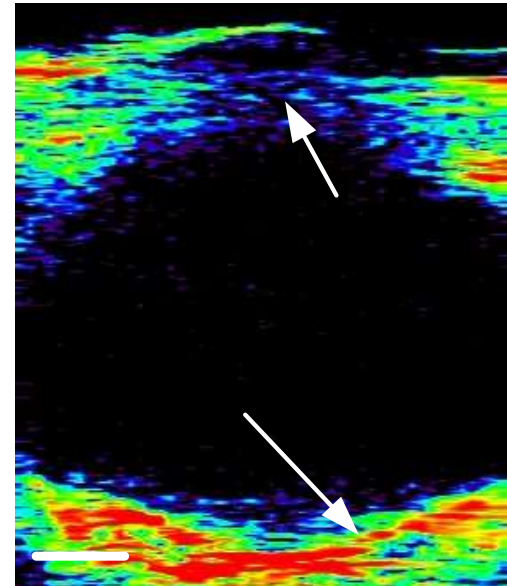
Photograph  
rabbit  
Vertical Slice



Parasagittal  
Ultrasound



Parasagittal  
Photoacoustic

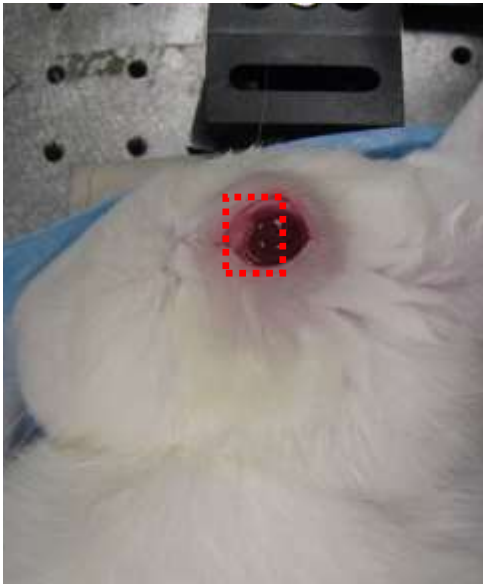


➤ **Good signal within safe laser level**

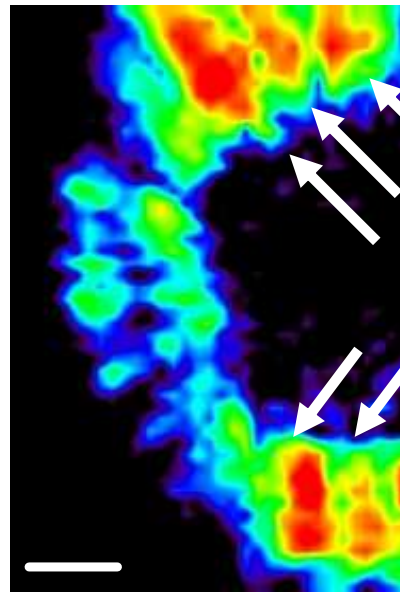
New Zealand rabbits, 6-8 weeks of age

# Live Rabbit Eye

**Coronal  
Horizontal Slice**



**Coronal  
Photoacoustic**



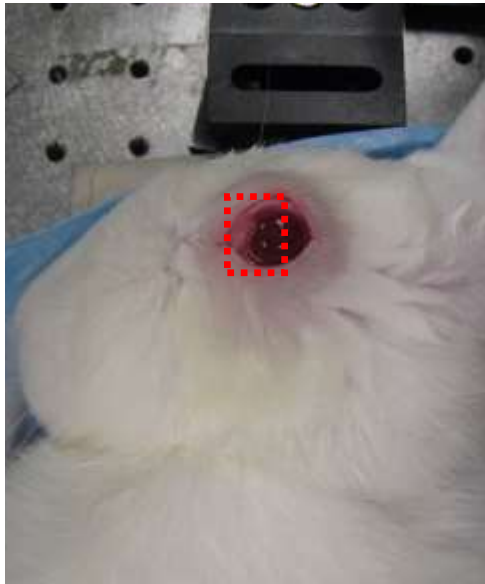
**Eye  
blood  
vessels**

➤ **Visualize individual blood vessels**

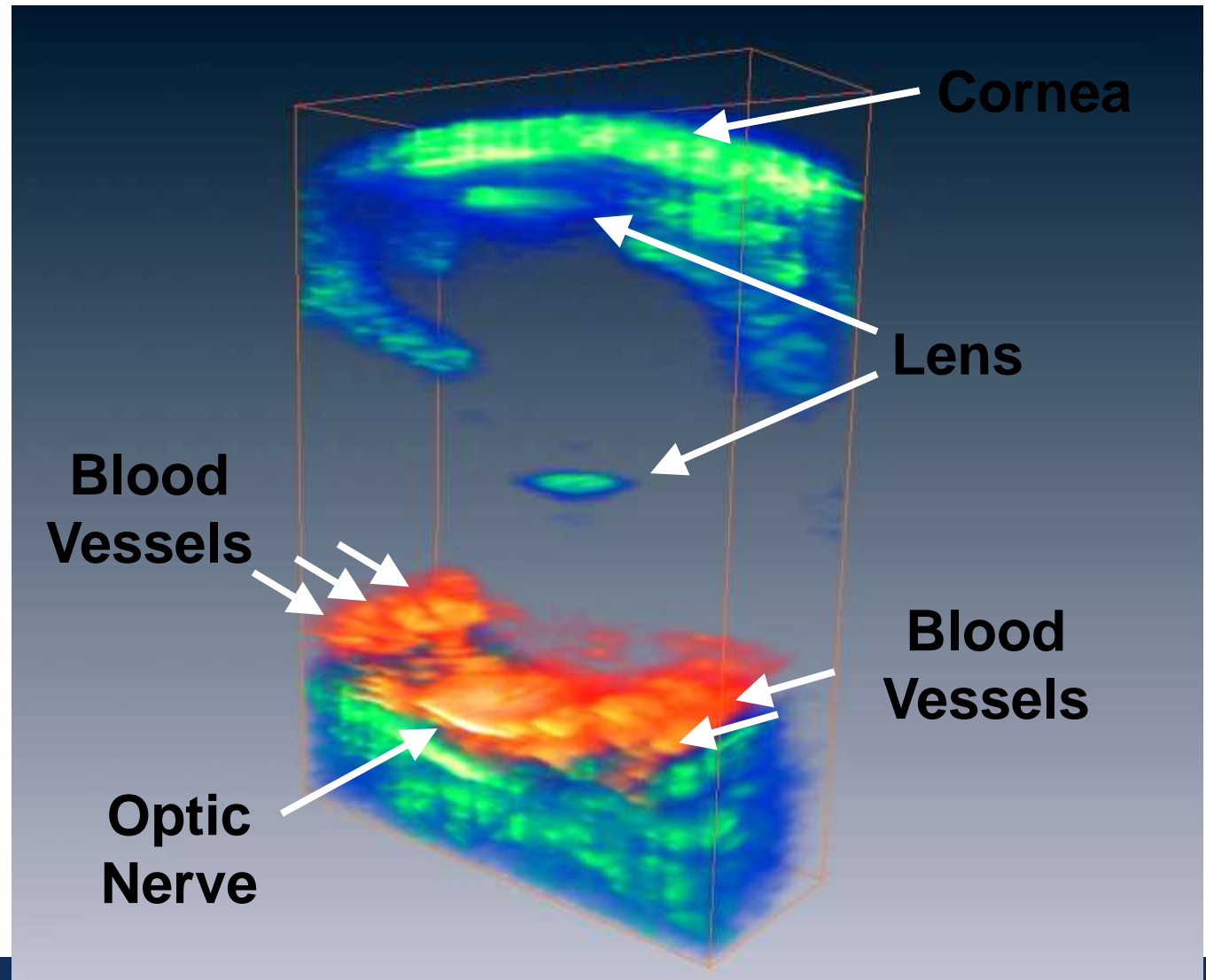
740nm laser, 5ns pulses,  $0.5\text{mJ}/\text{cm}^2$ , 25MHz focused transducer

# Live Rabbit Eye 3D Reconstruction

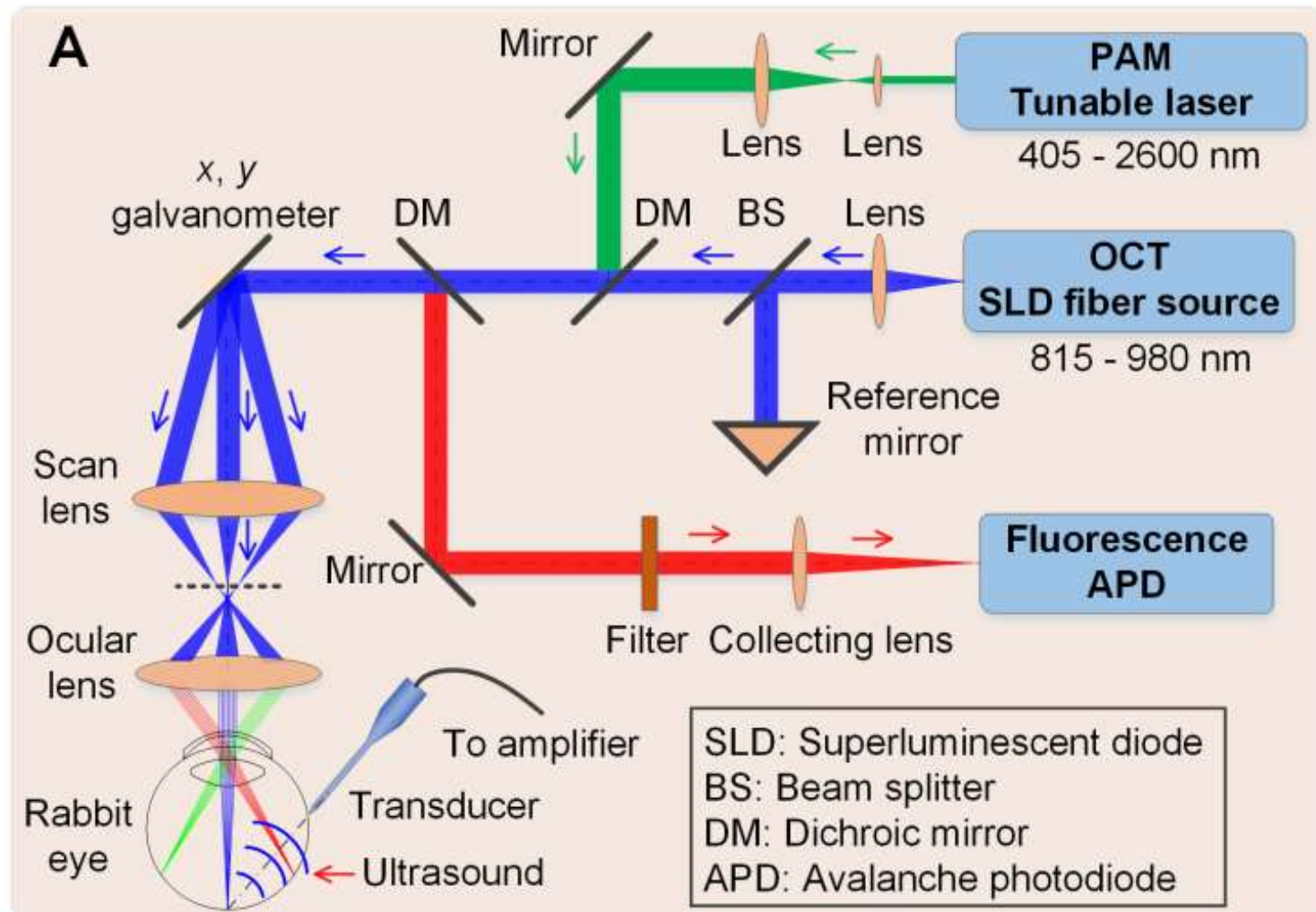
## 3D Rendering

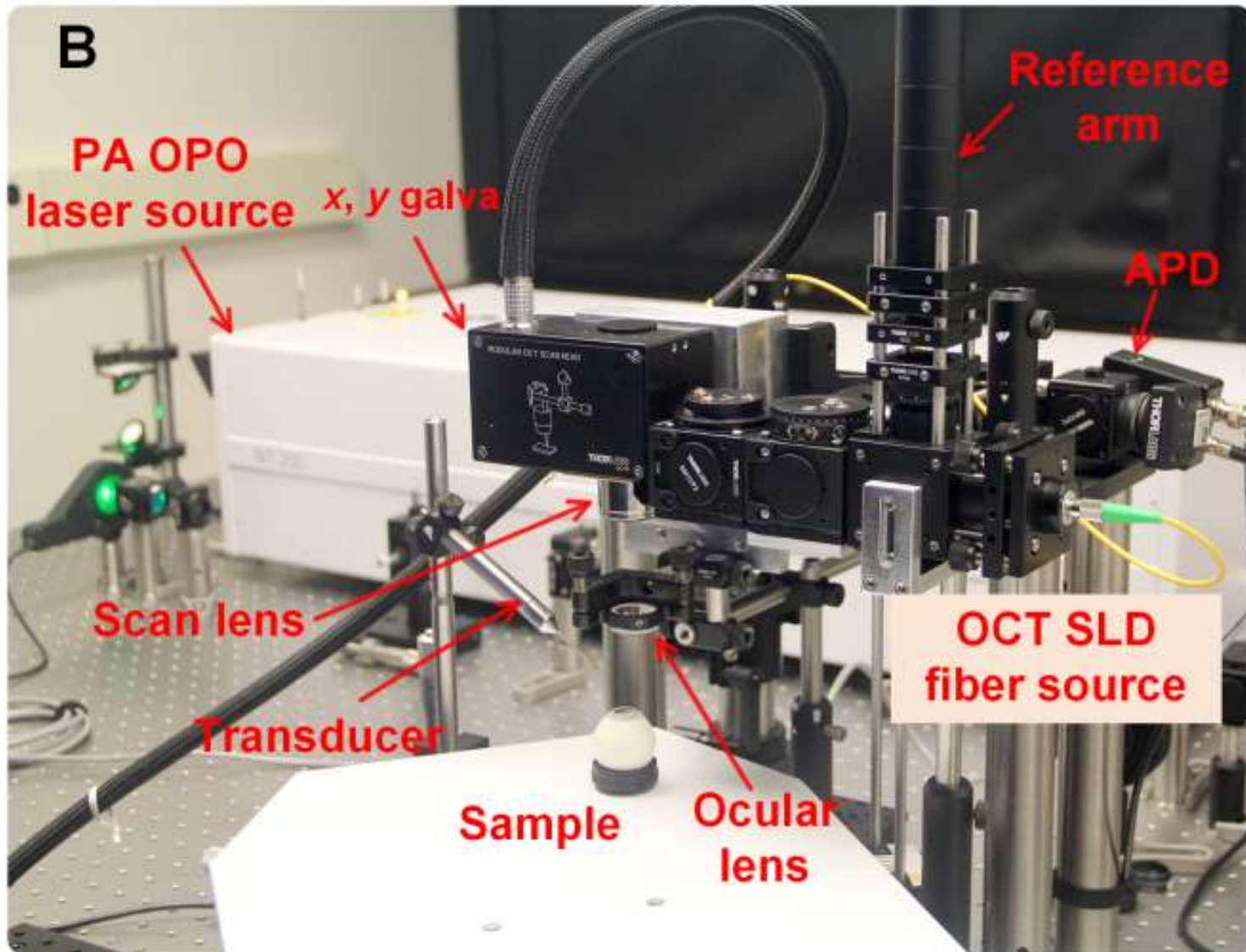


**32 averages**  
**Area: 12**  
**mm x 8 mm** 250  
**µm steps**



# Schematic multimodal photoacoustic microscopy (PAM), OCT, fluorescence microscopy





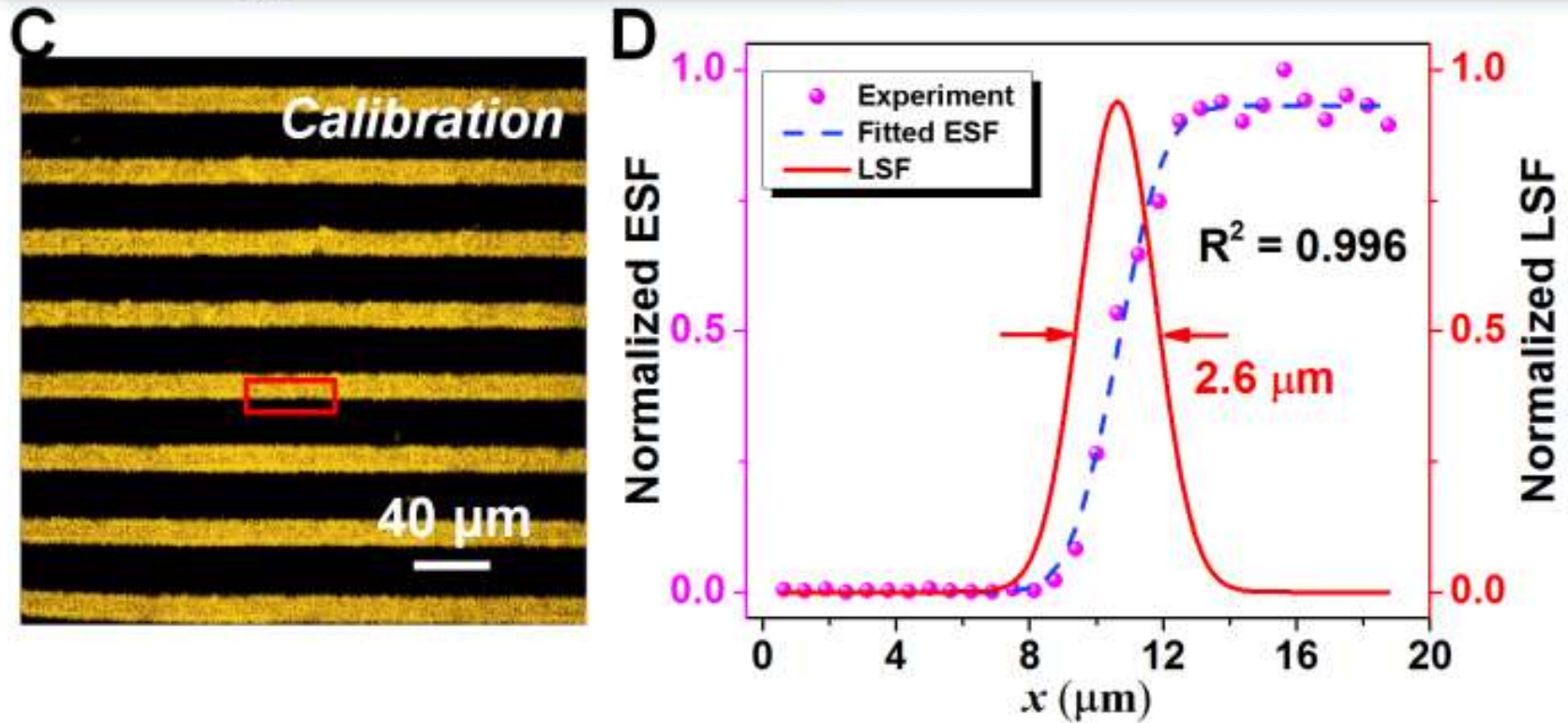
Tunable pulsed laser (Ekspla NT-242, Lithuania) with 1,000 Hz repetition rate

Ultrasound transducer center frequency: 35 MHz

Thorlabs Ganyemede-II-High resolution system 36 kHz



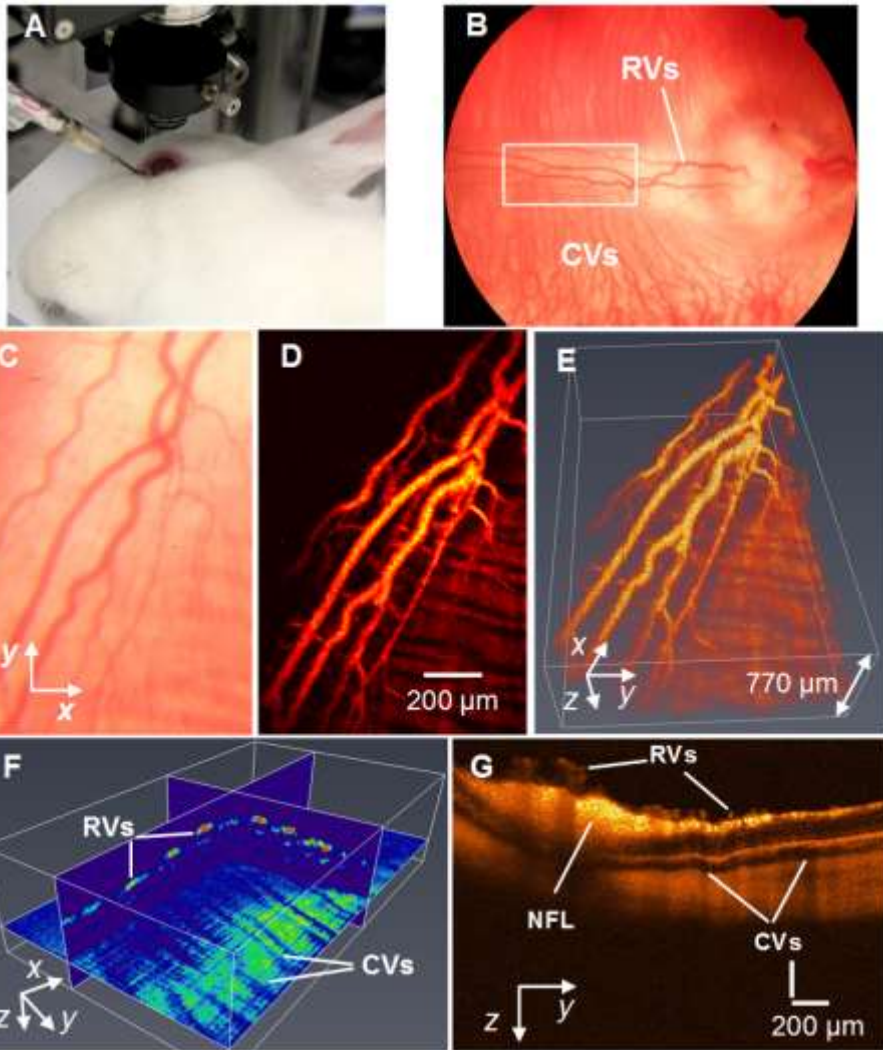
# PAM 2.6 $\mu\text{m}$ lateral resolution



A chrome grating was imaged at the focal plane of the scan lens for lateral resolution calibration.

Initial Laser pulse energy:  $\sim 30$  nJ (half ANSI safety limit)

# PAM/OCT Retina



**A:** External photo.

**B:** Fundus photograph.

**C:** Photo retinal medullary ray vessels.

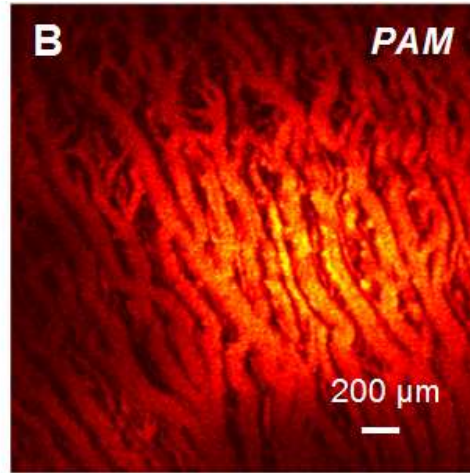
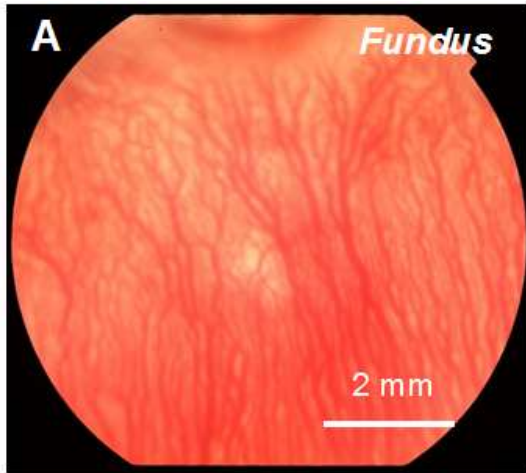
**D:** PAM of retinal (RV) and choroidal vessels (CVs).

**E:** 3D PAM.

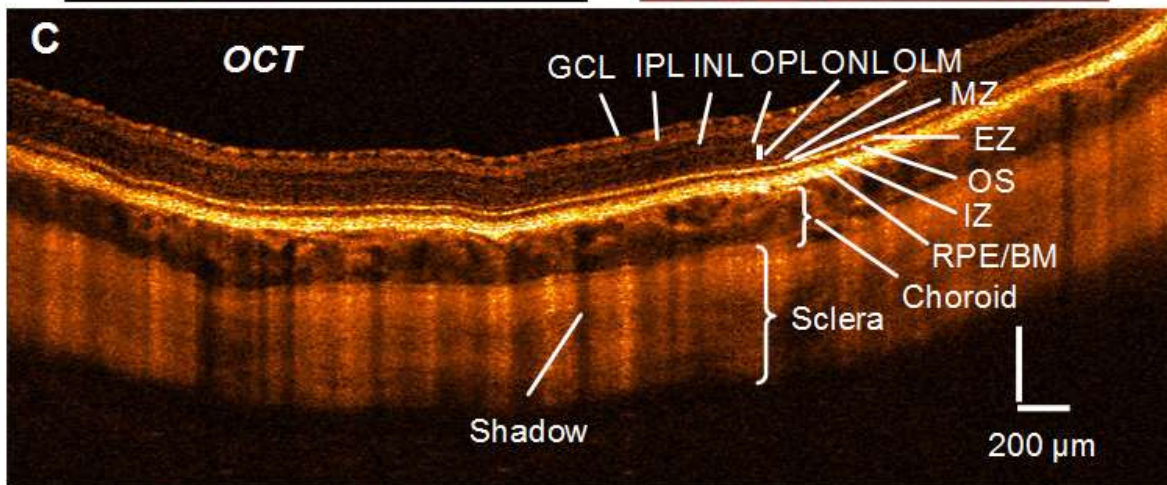
**F:** 2D orthogonal slices of the PAM image.

**G:** OCT image showing RVs, CVs, NFL (nerve fiber layer), and retinal layers.

# PAM/OCT Choroid



**A:** Fundus photograph.  
**B:** PAM signal of the choroid  
**C:** OCT image.

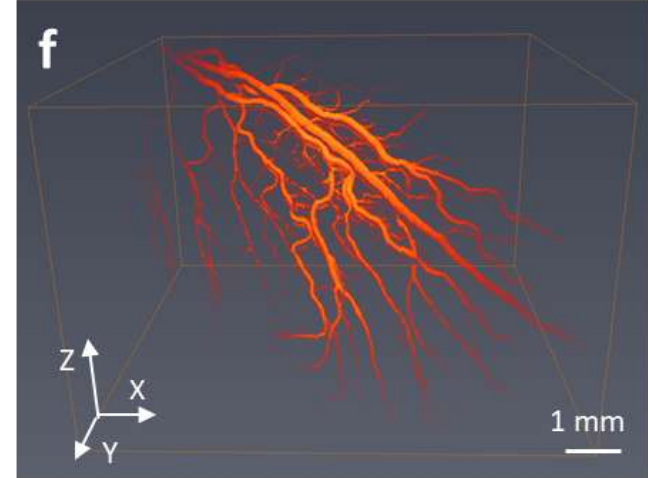
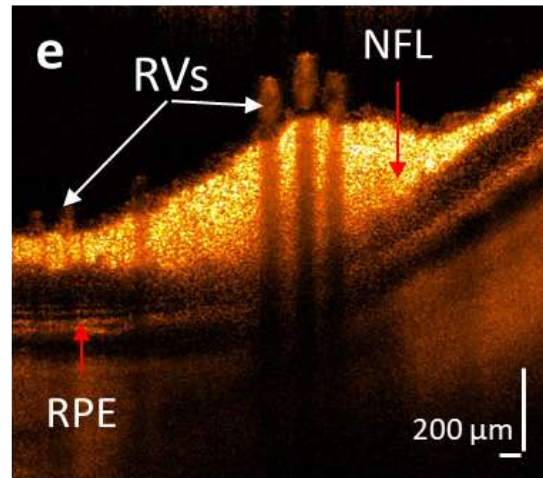
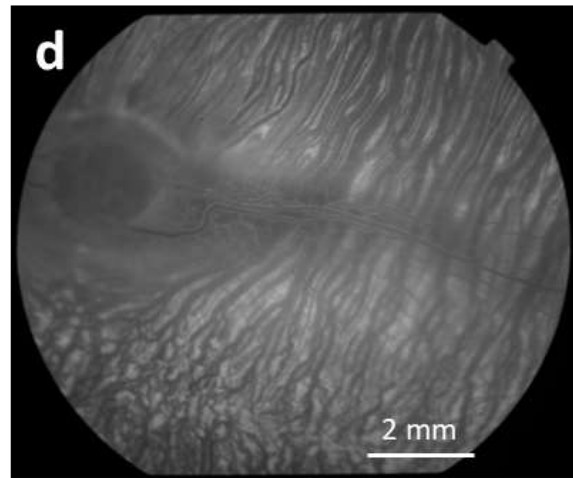
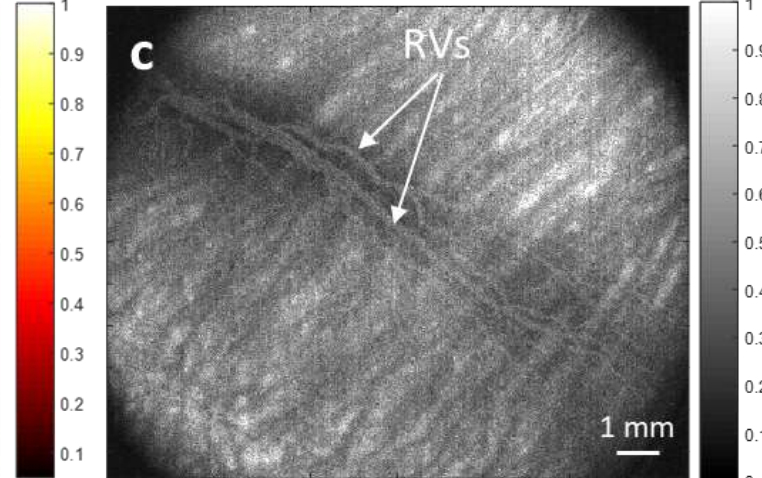
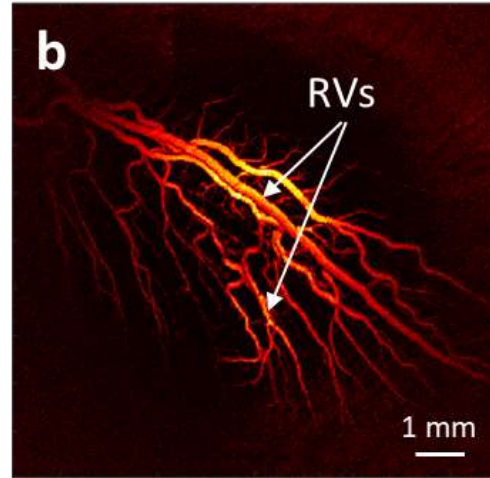
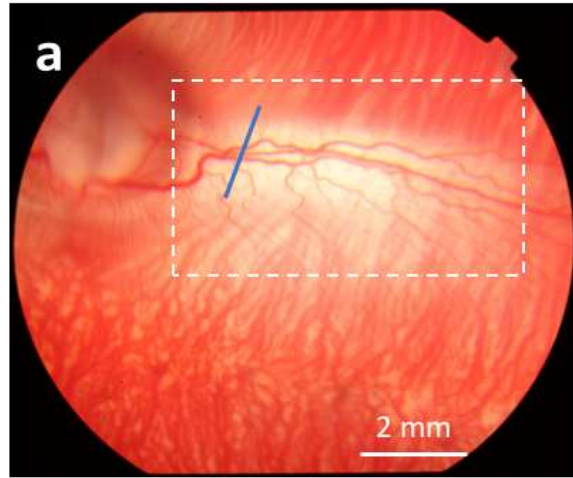


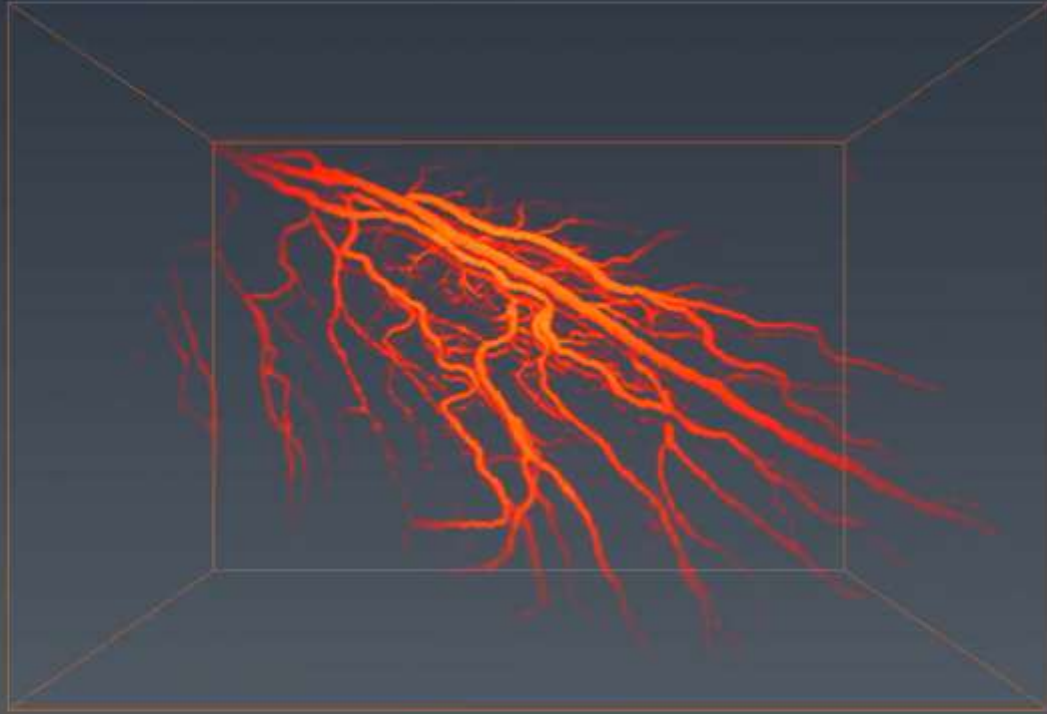
GCL: ganglion cell layer; IPL: inner plexiform layer; INL: inner nuclear layer; OPL: outer plexiform layer; ONL: outer nuclear layer; OLM: outer limiting membrane; MZ: myoid zone; EZ: ellipsoid zone; OS: outer segment; IZ: interdigitation zone; BM, Bruch's membrane.

PAM can be performed using a safe laser exposure dose (~80 nJ) below the ANSI safety limit (160 nJ) at 570 nm. Now down to 5% ANSI limit

# Normal New Zealand white rabbits

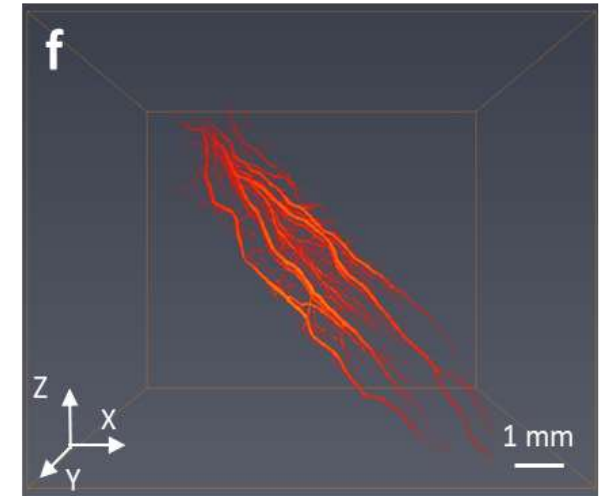
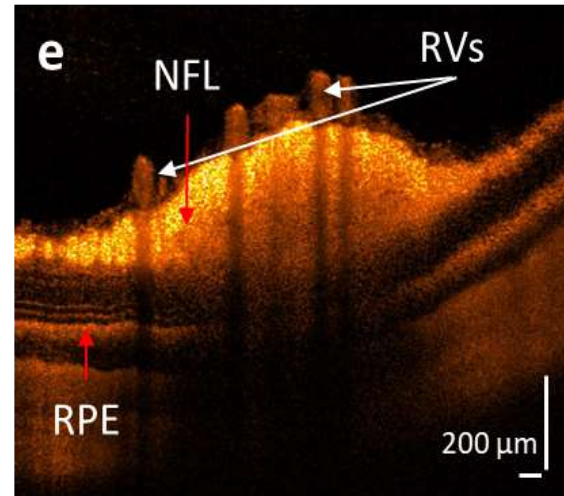
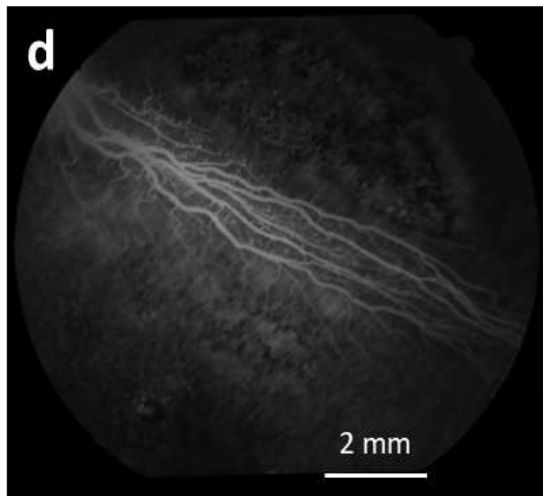
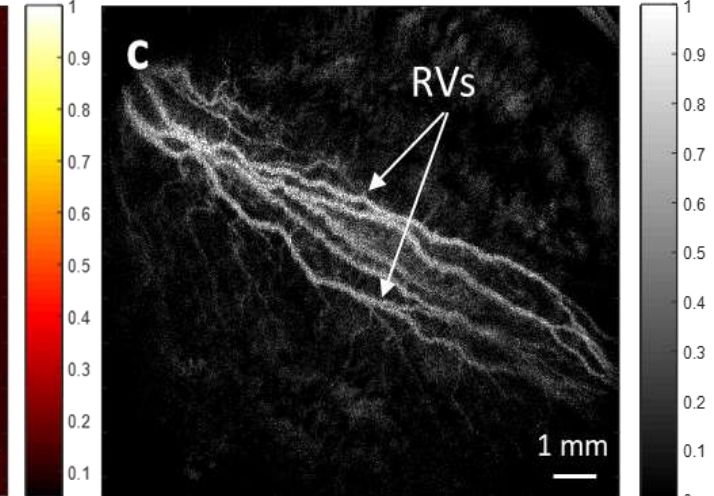
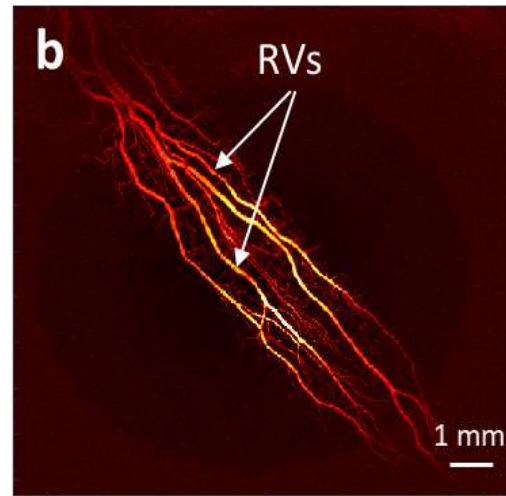
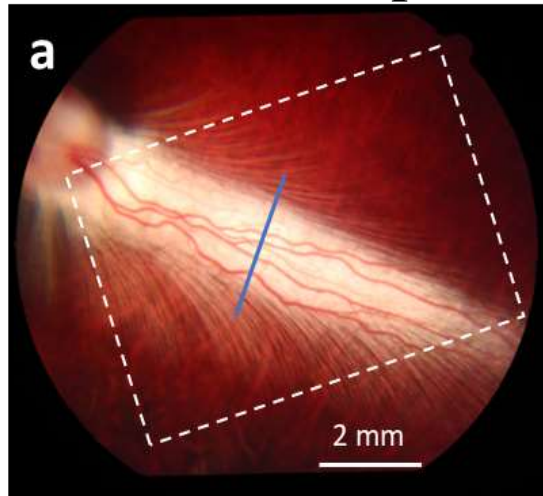
(a) Color fundus photo; (b) PAM (c) FM; (d) FA; (e) OCT; (f) 3D PAM

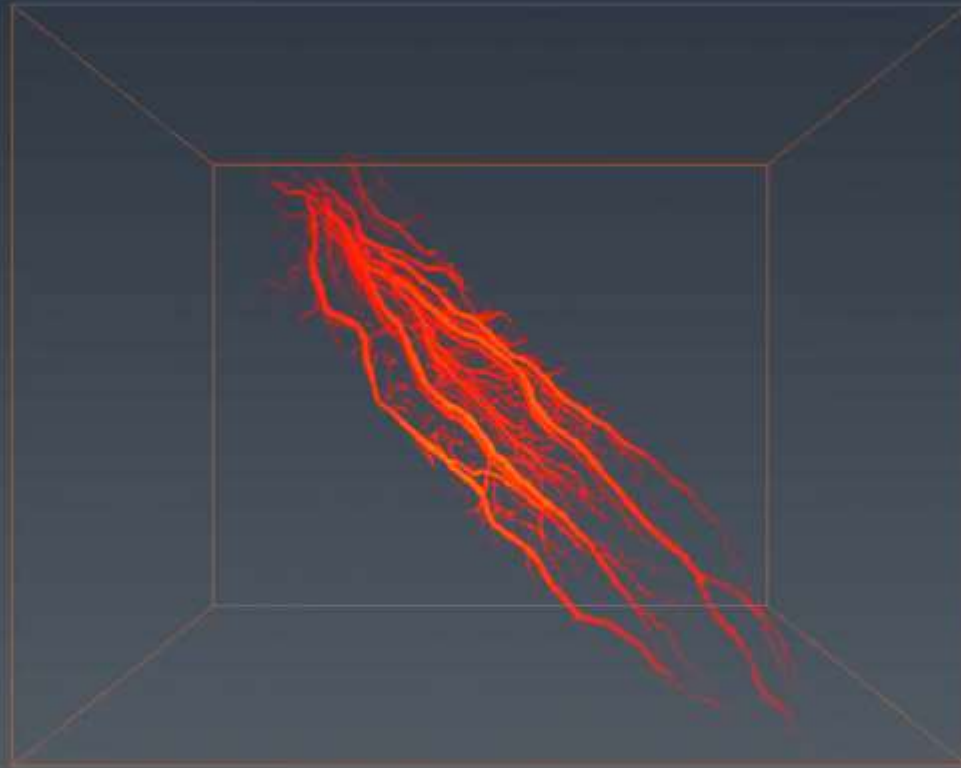




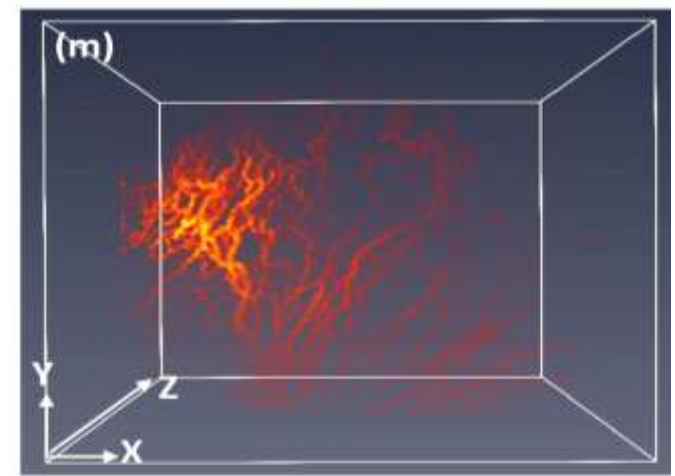
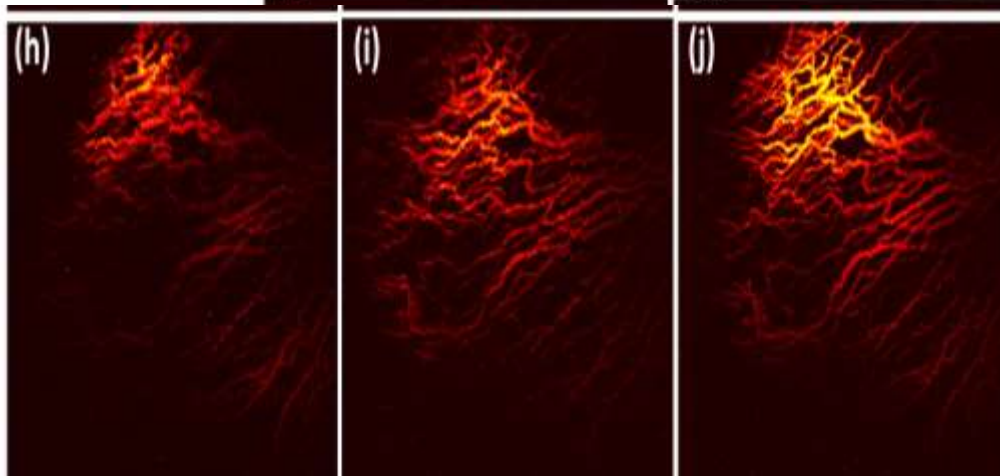
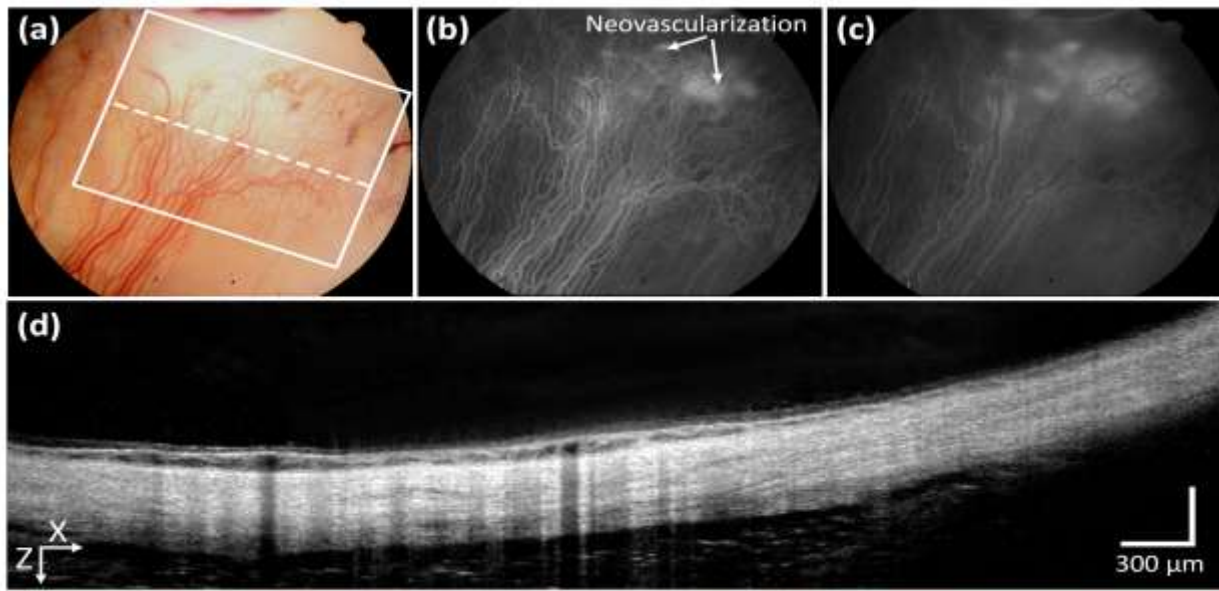
# Normal Pigmented rabbits

(a) Color fundus photo; (b) PAM (c) FM; (d) FA; (e) OCT; (f) 3D PAM





# Retinal Vein Occlusion Rabbit Model



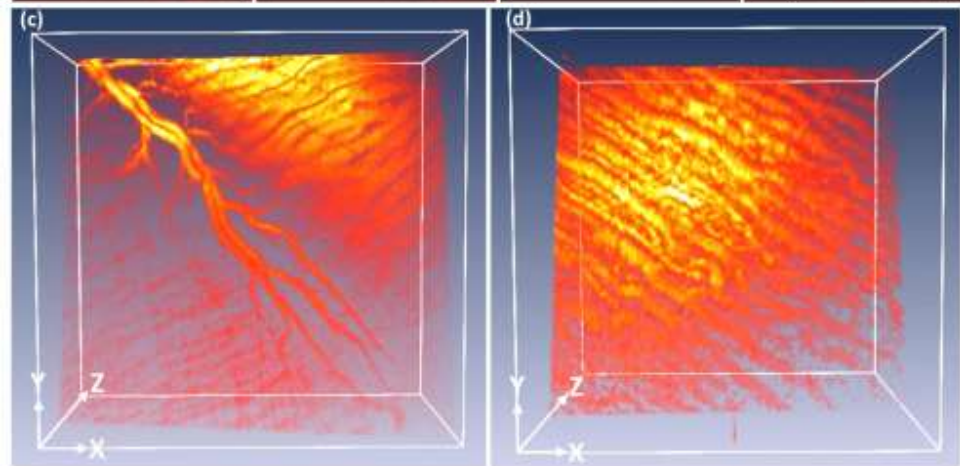
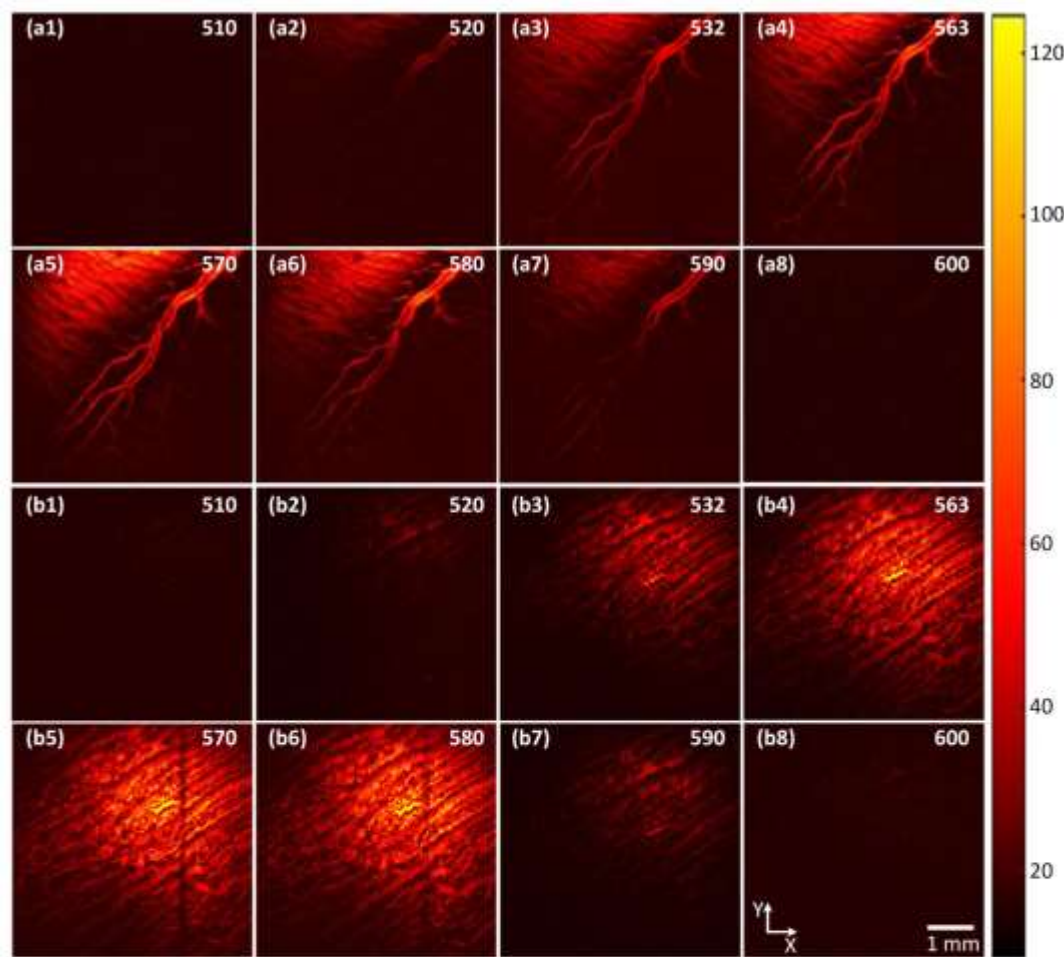


# Spectroscopic PAM analysis of the retina and choroid

A. Retinal vessels

B. Choroidal vessels

C and D. 3D reconstructions of A and B



510

520

532

563

570

580

590

600

Day 0

Day 3

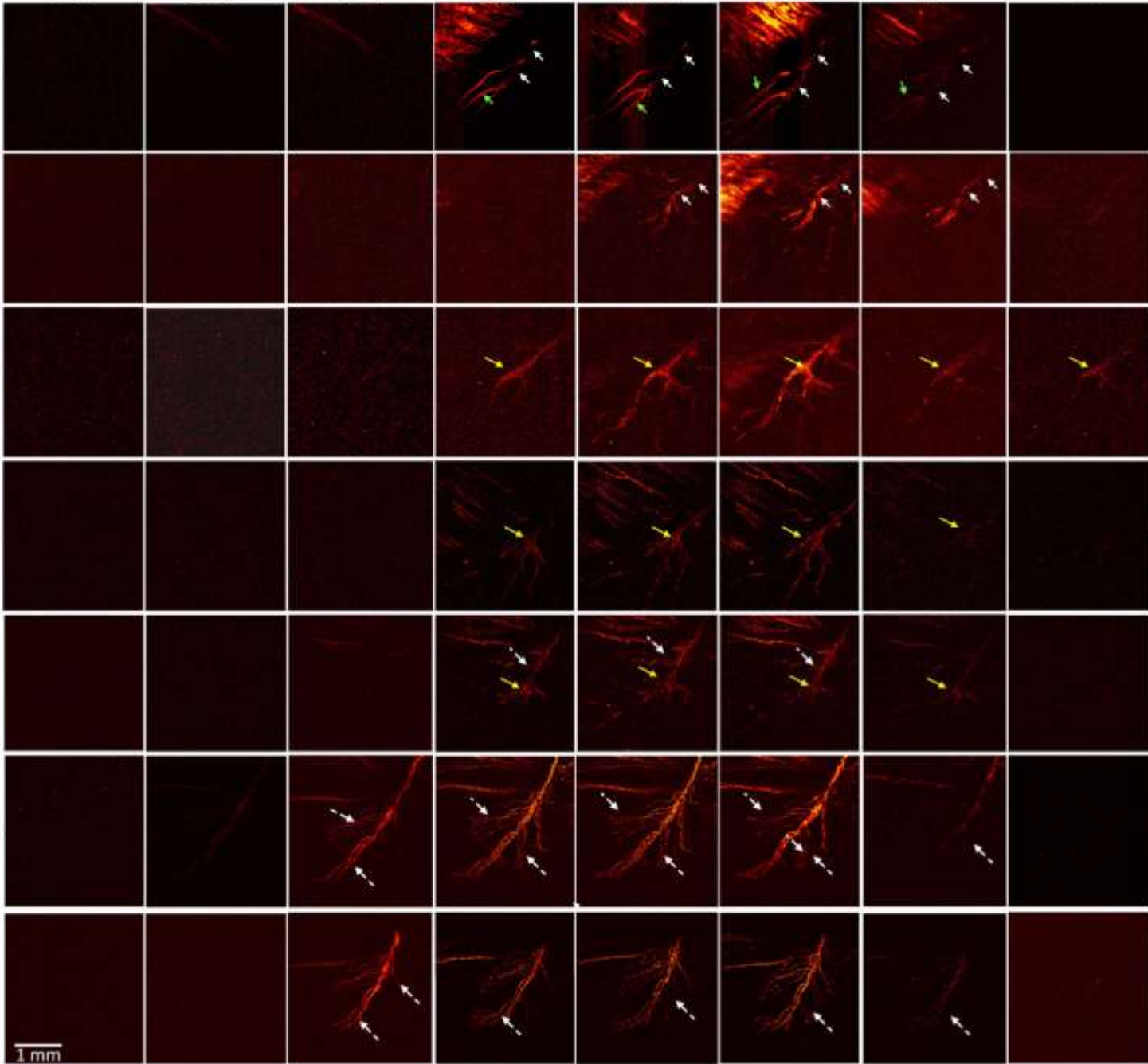
Day 7

Day 14

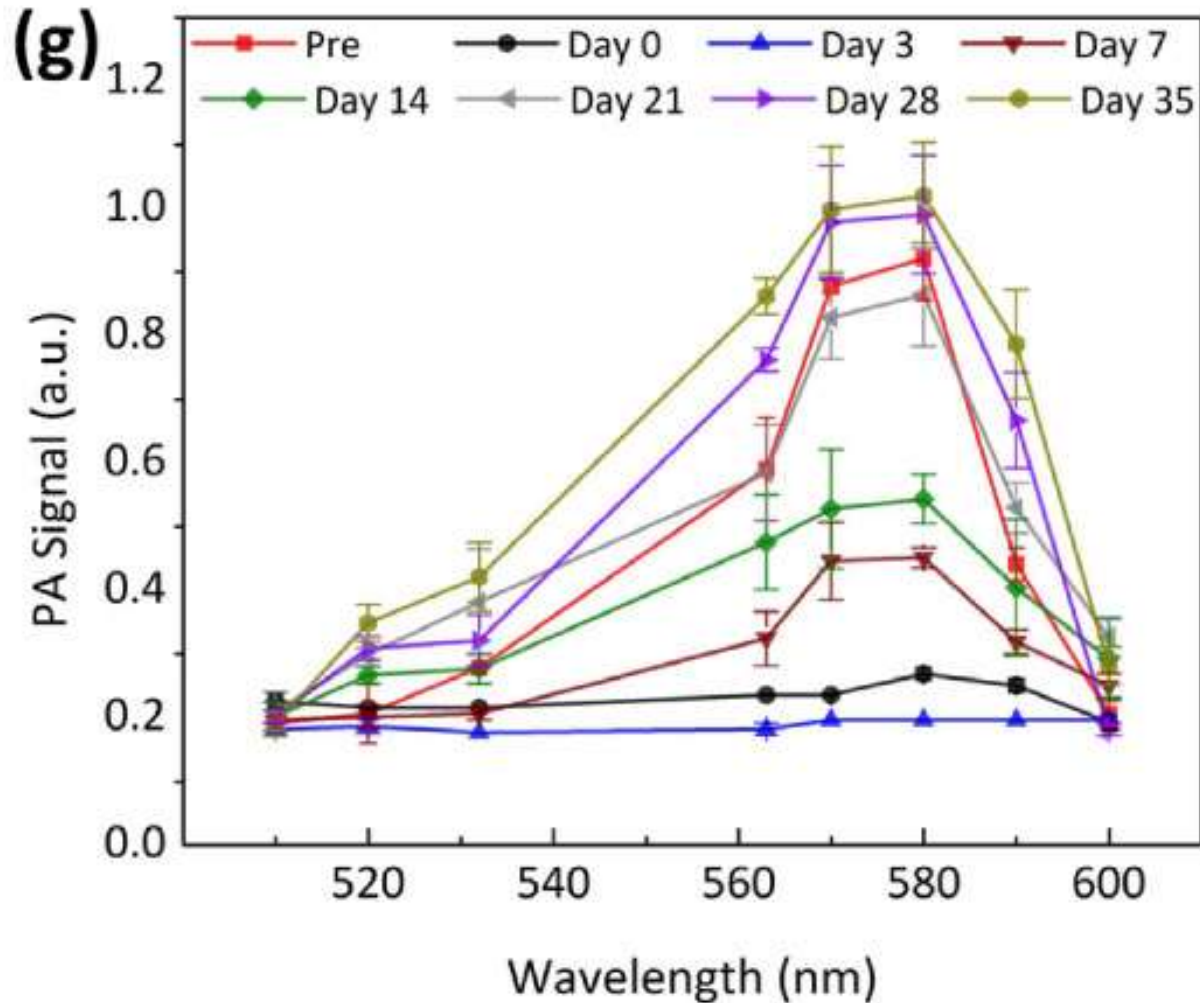
Day 21

Day 28

Day 35

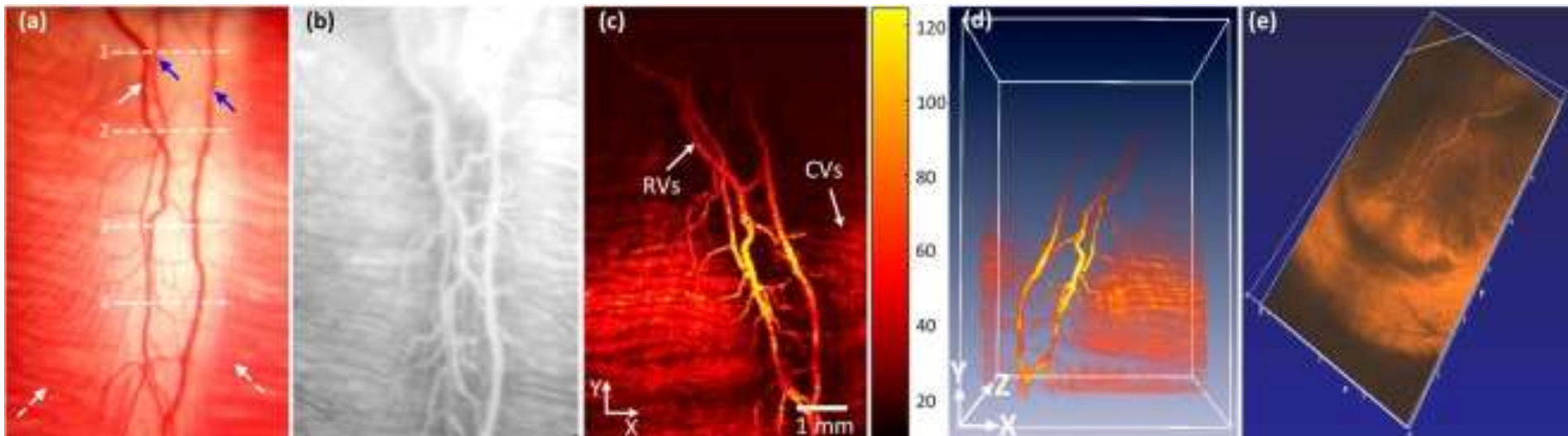


# Quantification of spectroscopic PAM

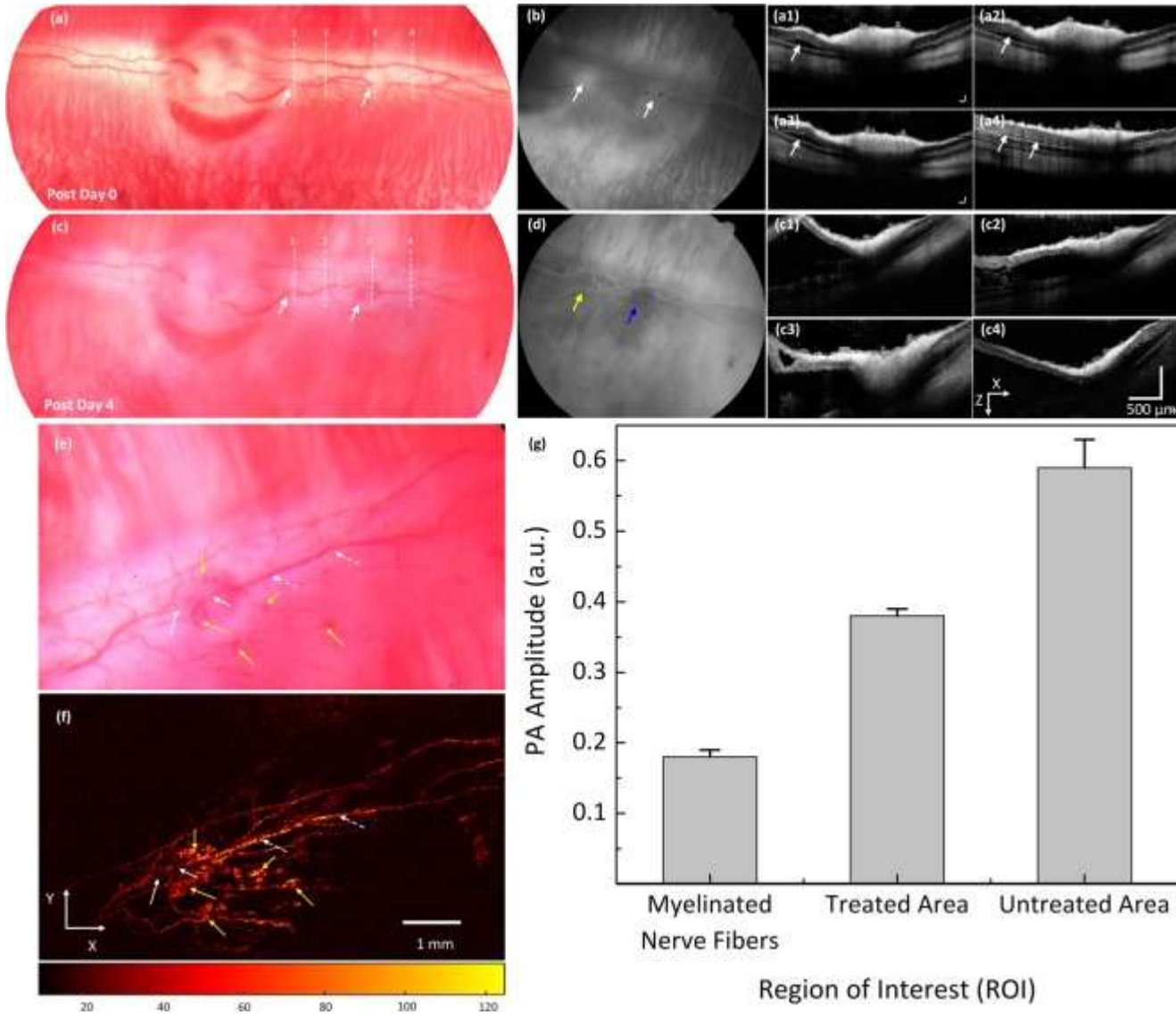


# Multimodal imaging retinal blood vessels

(a) Color photo; (b) FA (c) PAM; (d) 3D PAM; (e) 3D OCT

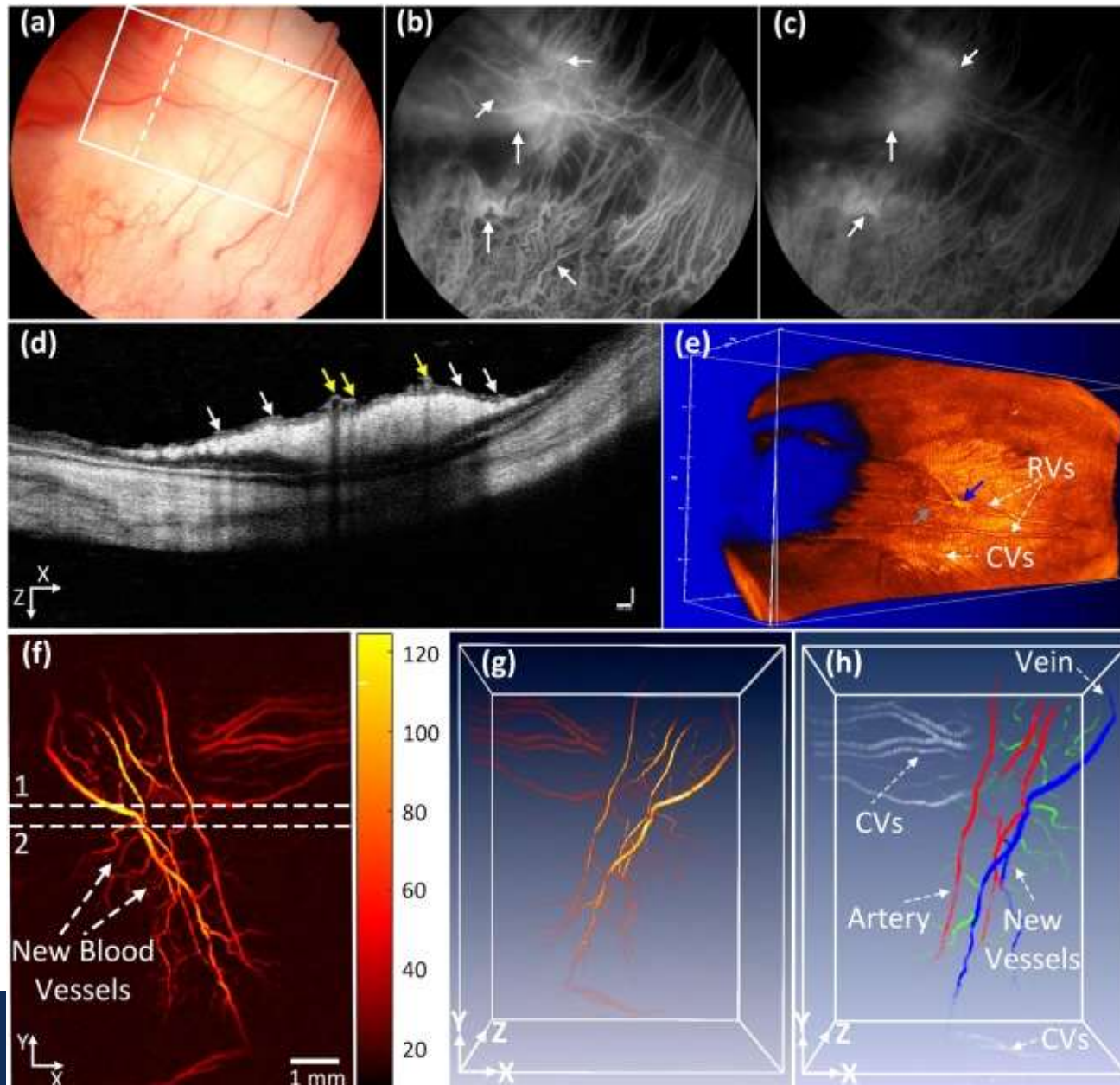


# Retinal vein occlusion



(a,c,e) Color photo;  
(b,d) FA  
(a1-4) OCT images;  
(f) PAM;  
(g) PAM amplitude

# Retinal neovascularization in RVO



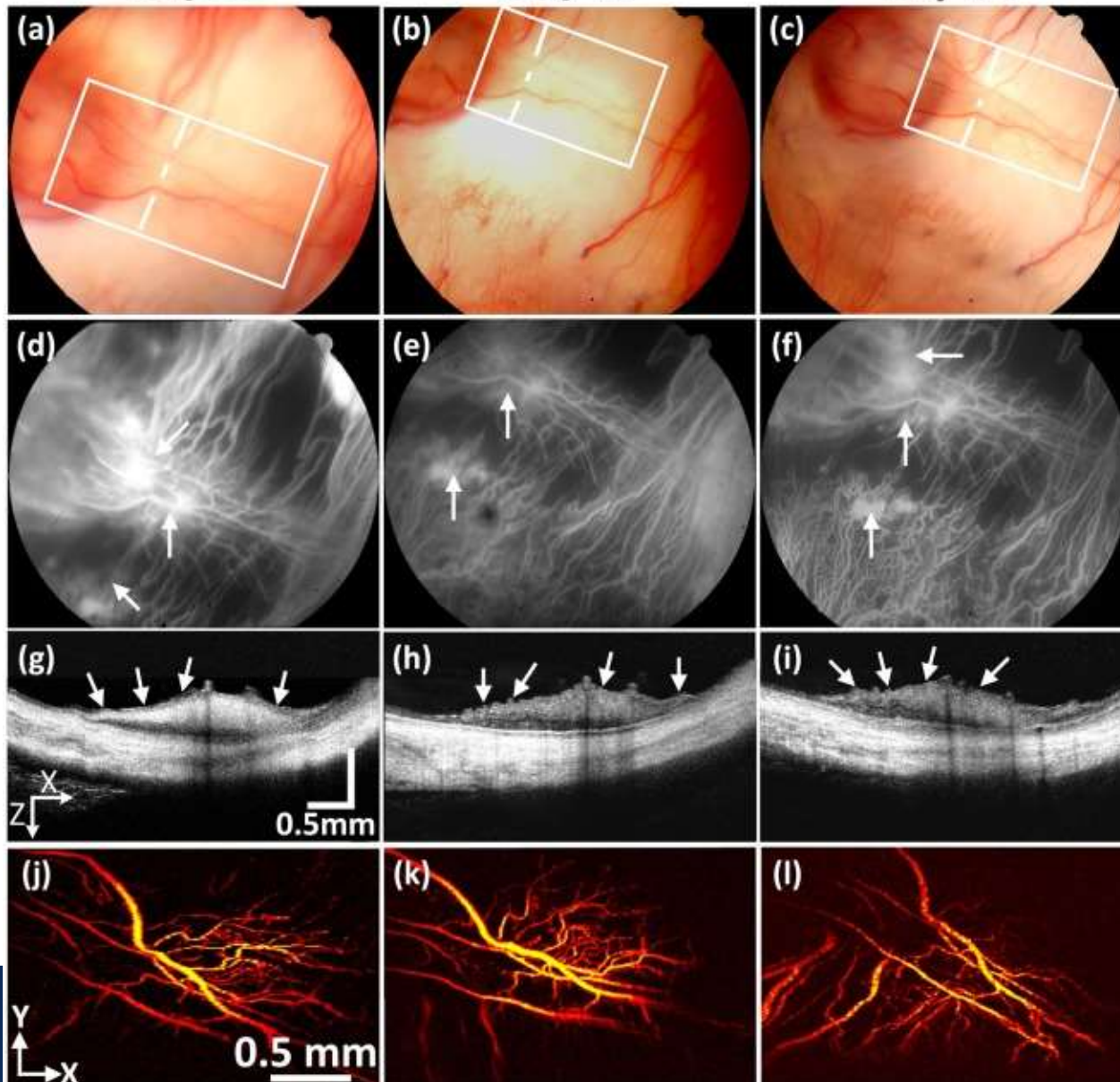
(a) Color photo;  
(b,c) FA early and late  
(d) OCT; (e) 3D OCT;  
(f) PAM;  
(g) 3D PAM;  
(h) Label type vessels

# Retinal neovascularization over time

Day 35

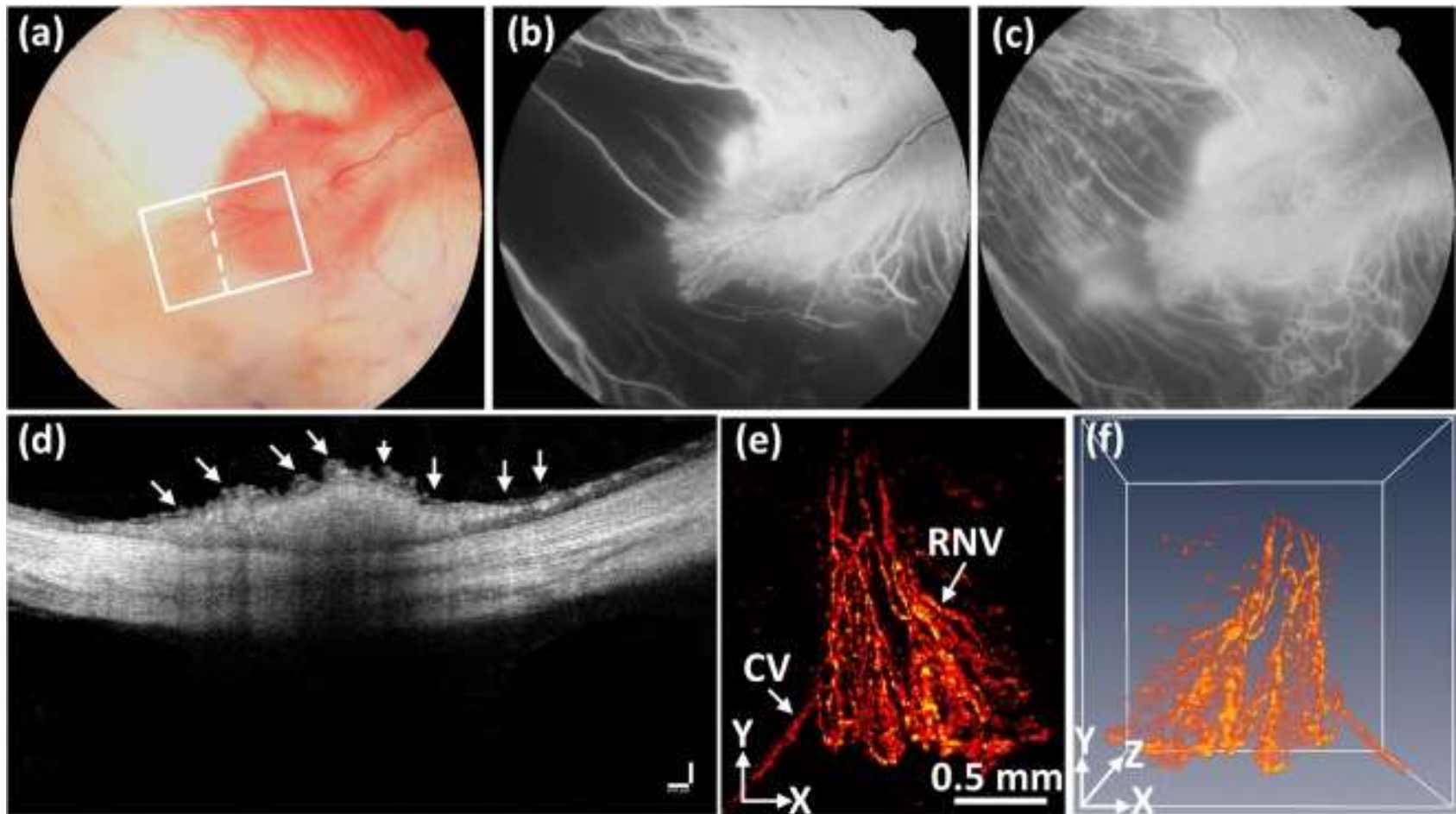
Day 49

Day 90



(a,b,c) Color photo;  
(d,e,f) FA  
(g,h,i) OCT;  
(j,k,l) PAM

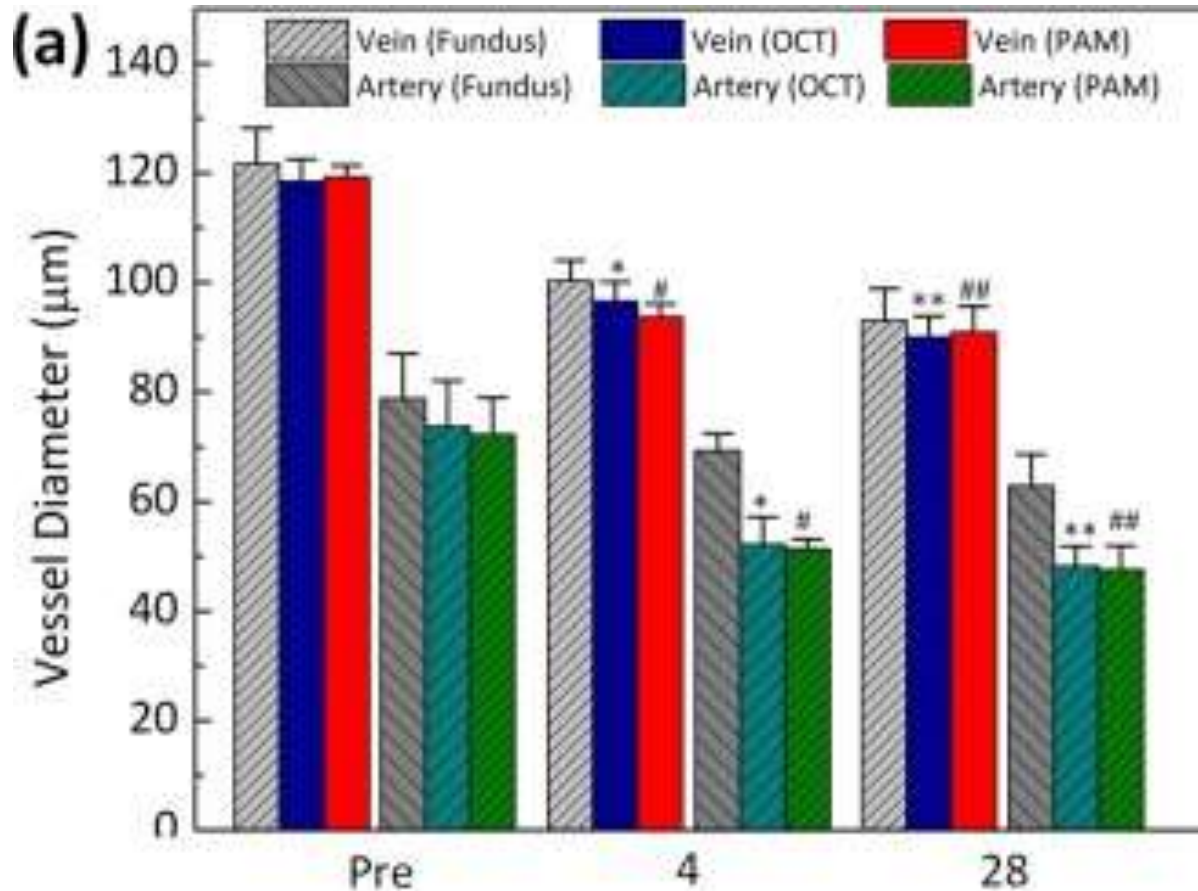
# Retinal neovascularization in RVO



(a) Color photo; (b,c) FA; (d) OCT; (e) PAM; (f) 3D PAM

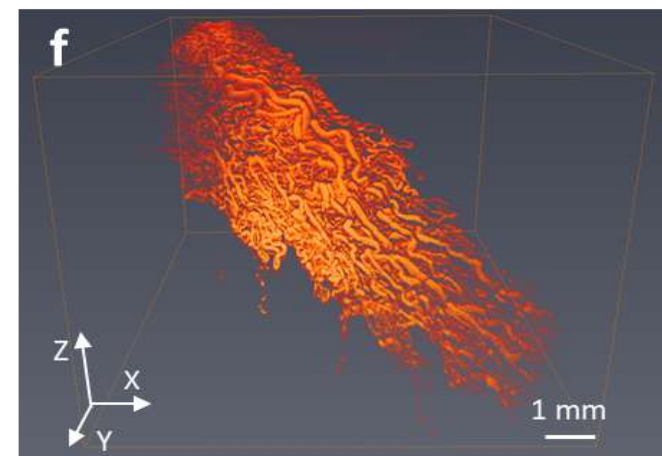
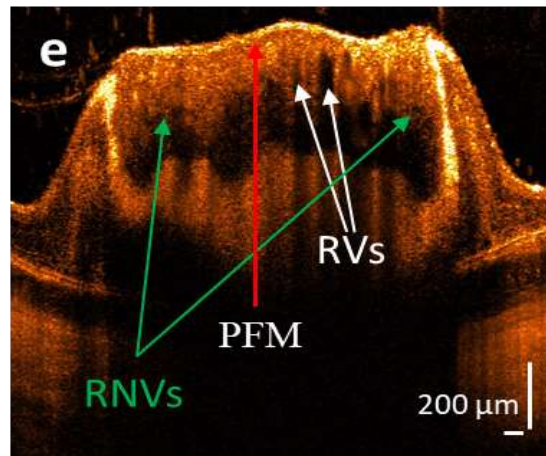
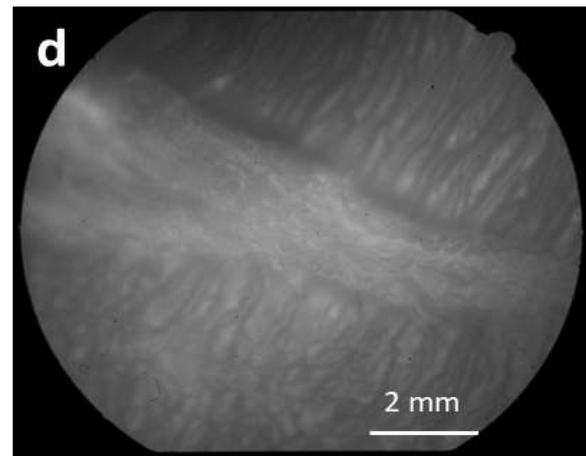
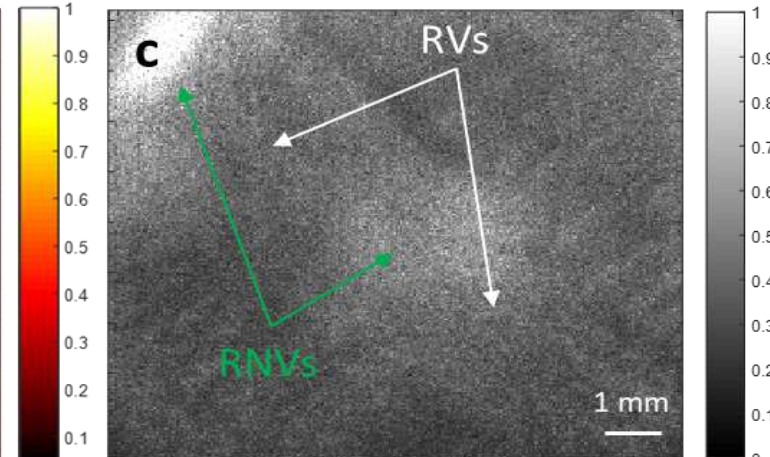
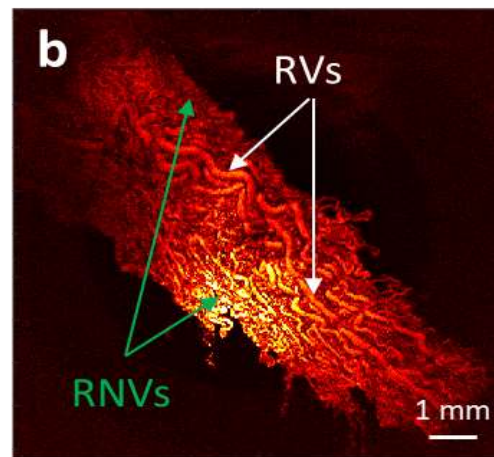
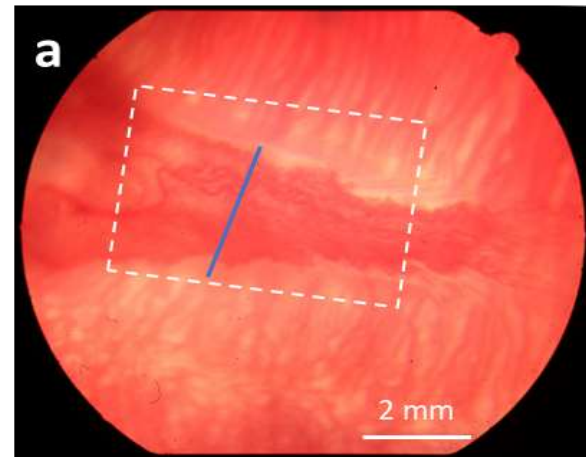


# Quantify vessel diameter photo, OCT, and PAM



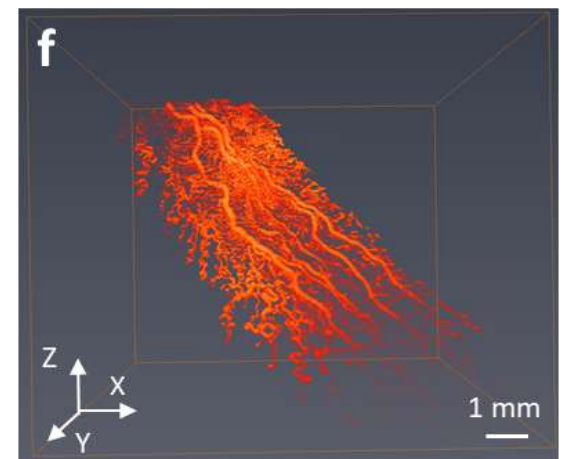
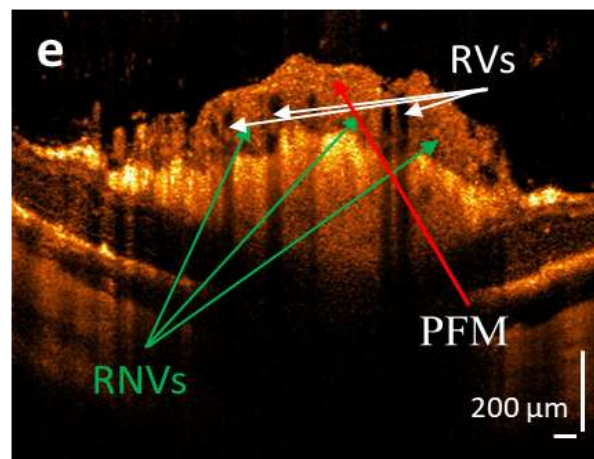
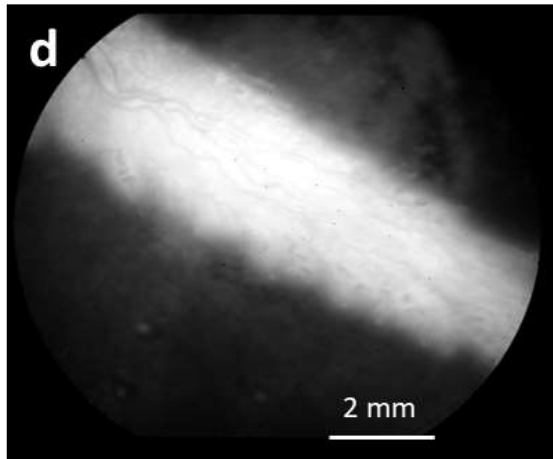
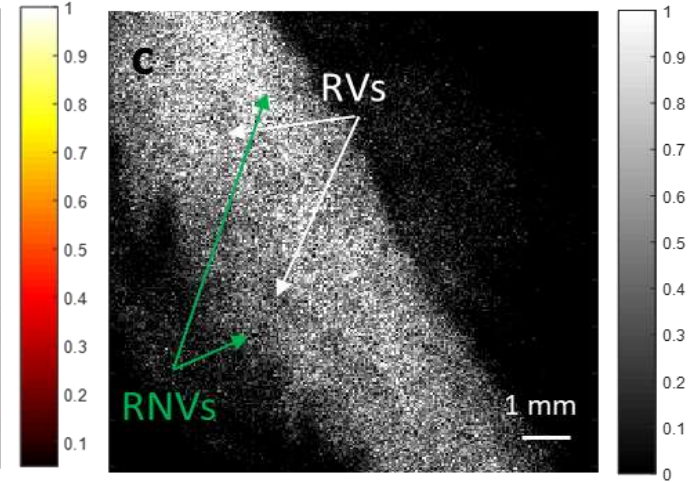
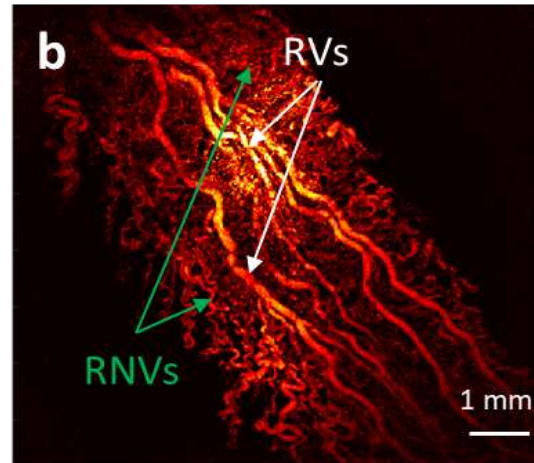
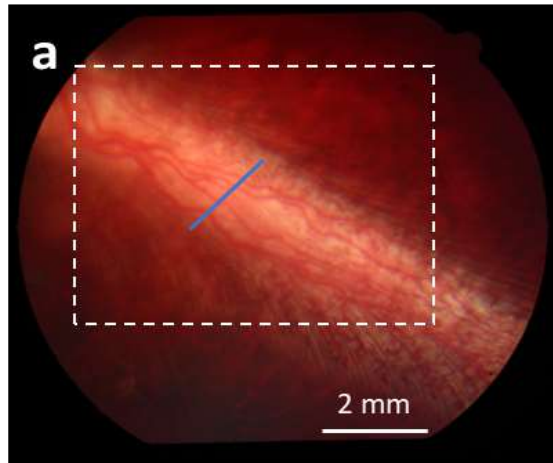
# New Zealand white rabbit RNV

(a) Color fundus photo; (b) PAM (c) FM; (d) FA; (e) OCT; (f) 3D PAM



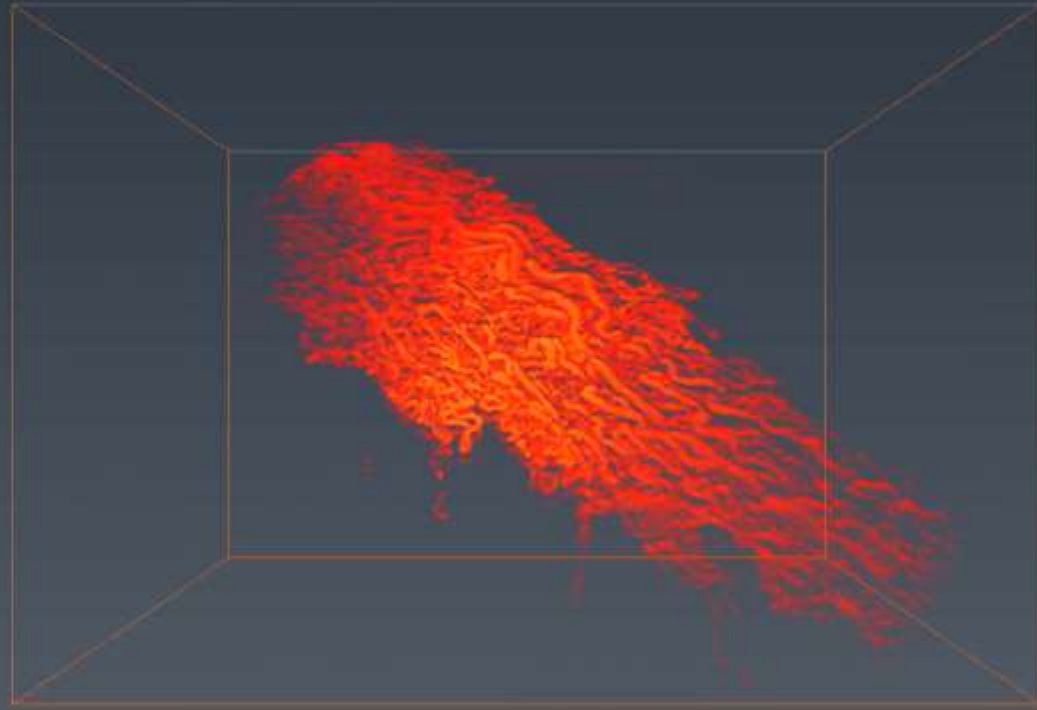
# Pigmented rabbits RNV

(a) Color fundus photo; (b) PAM (c) FM; (d) FA; (e) OCT; (f) 3D PAM

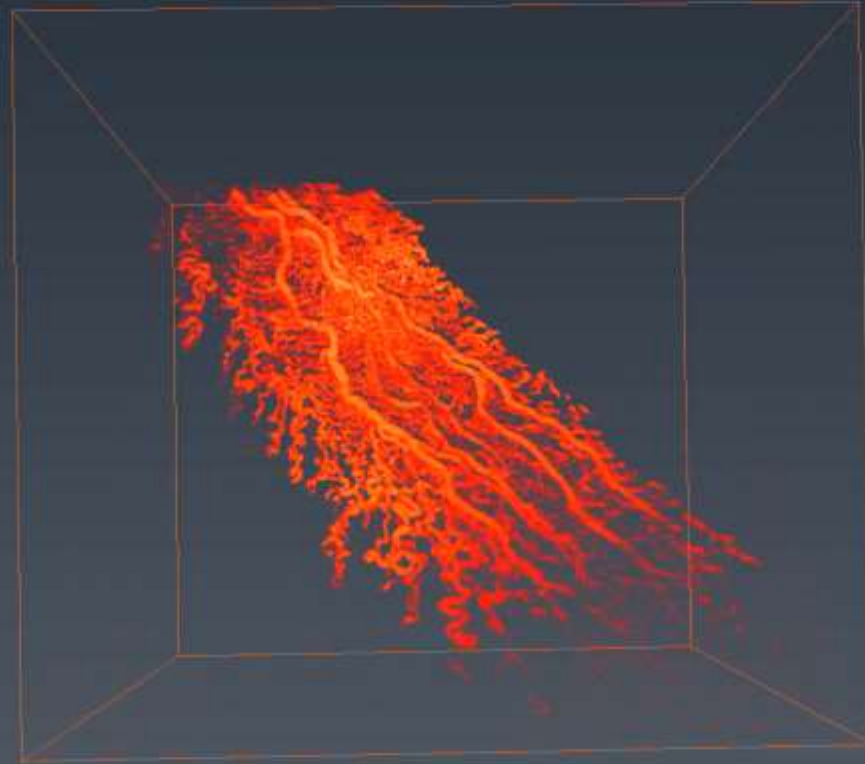


Zhang W, Li Y, Nguyen VP, Huang Z, Liu Z, Wang X, Paulus YM. High resolution, in vivo Multimodal Photoacoustic Microscopy, Optical Coherence Tomography, and Fluorescence Microscopy Imaging of Rabbit Retinal Neovascularization. *Light: Science & Applications* 2018; 7:103.

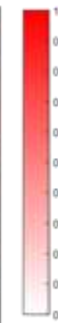
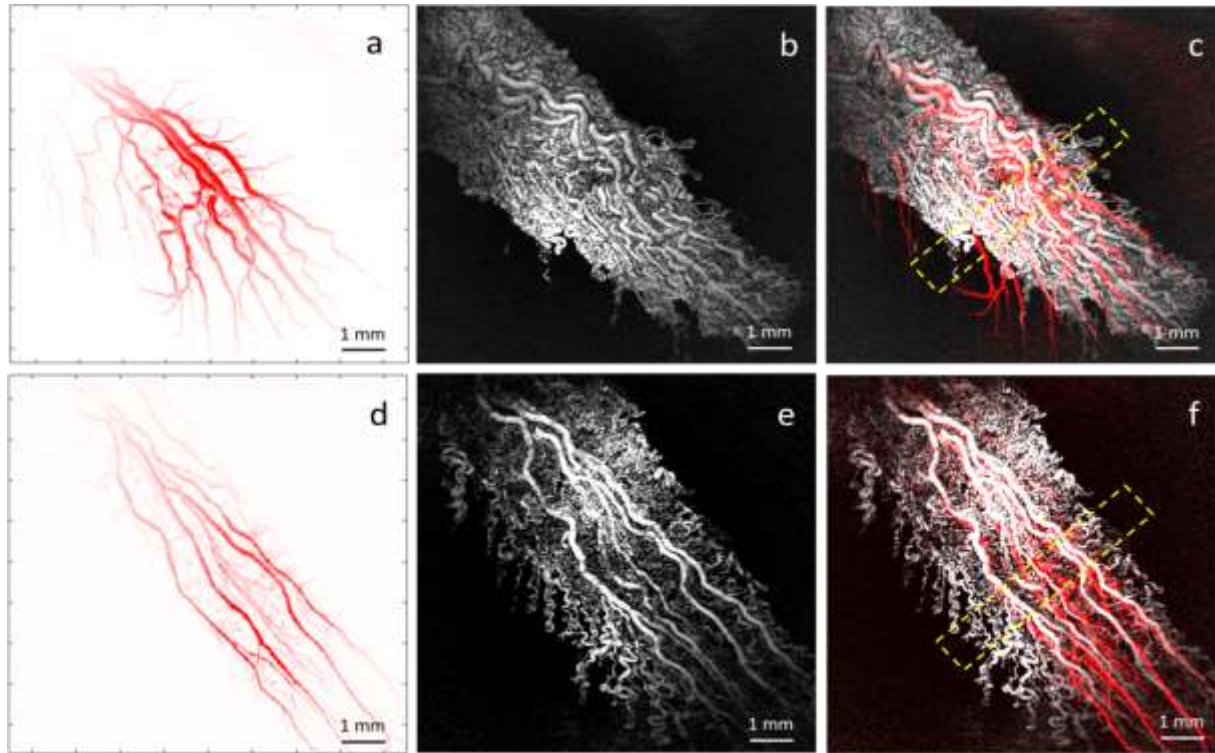
# New Zealand white rabbit neovascularization



# PAM pigmented rabbit neovascularization



# PAM normal vasculature compared to neovascularization



(a) Normal retinal vessels in New Zealand white rabbit;

(b) RNV induced by VEGF in NZ rabbit;

(c) composite pseudo color image of NZ rabbit showing the retinal vessels before and after VEGF injection;

(d) normal retinal vessels in pigmented rabbit;

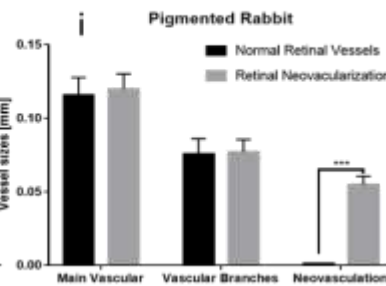
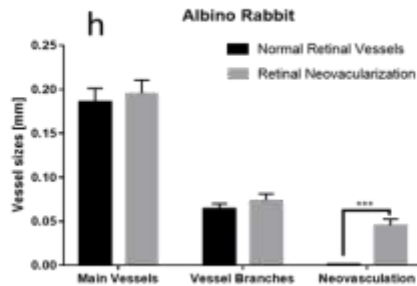
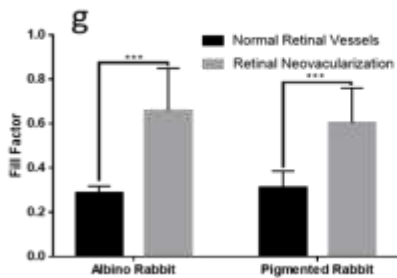
(e) RNV induced by VEGF injection in a pigmented rabbit;

(f) composite pseudo color image of pigmented rabbit showing the retinal vessels before and after VEGF injection;

(g) quantification of retinal vessels and RNV in NZ and pigmented rabbits before and after VEGF injection;

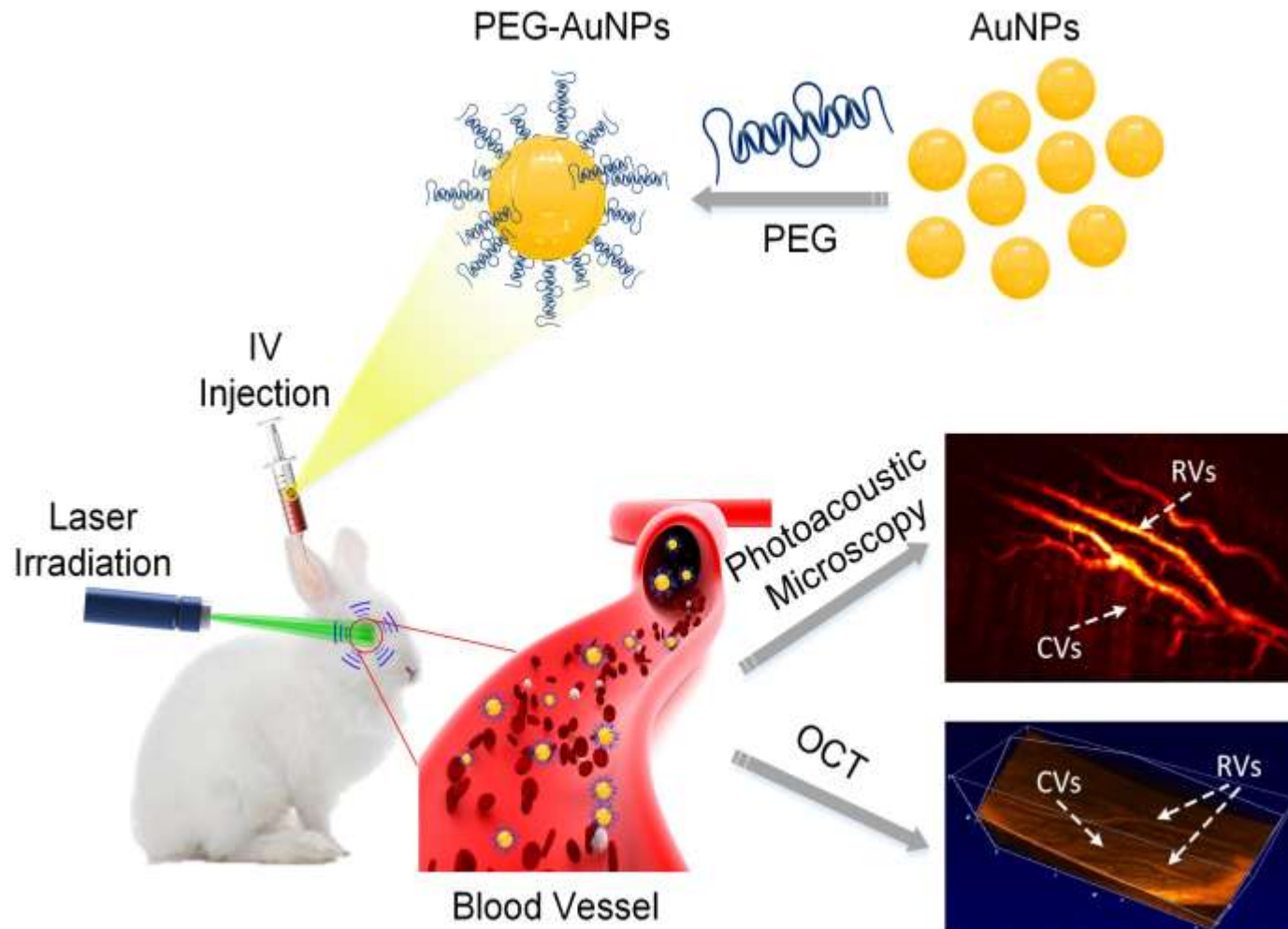
(h) quantification of retinal vessels and RNV in NZ rabbits before and after VEGF injection using the vessel size;

(i) quantification of retinal vessels and RNV in pigmented rabbits before and after VEGF injection using the vessel size



# Gold-nanoparticle enhanced PAM imaging

Photoacoustic imaging with gold nanoparticles (AuNP) can significantly enhance signal of PAM and OCT



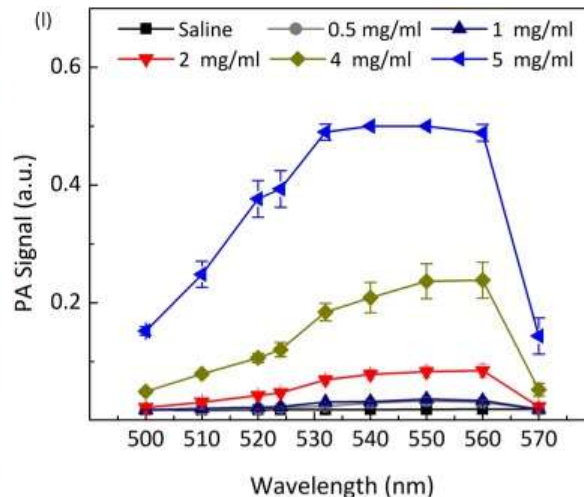
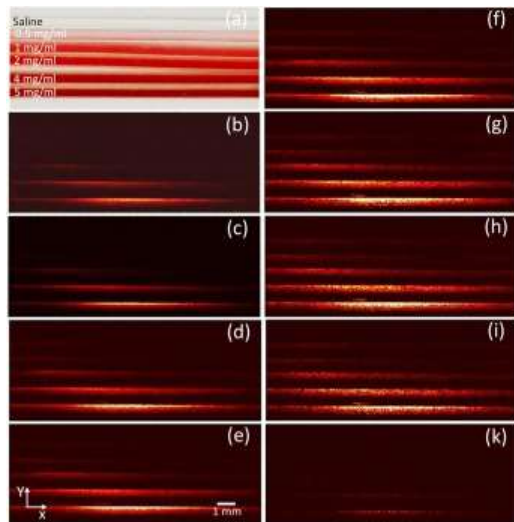
Nguyen VP, Li Y, Qiao W, Liu B, Tian C, Zhang W, Huang Z, Ponduri A, Tarnowski M, Wang X, Paulus YM. Contrast Agent Enhanced Multimodal Photoacoustic Microscopy and Optical Coherence Tomography for Imaging of Rabbit Choroidal and Retinal Vessels *in vivo*.

*Scientific Reports* 2019; 9(1):5945.



**KELLOGG EYE CENTER**  
MICHIGAN MEDICINE

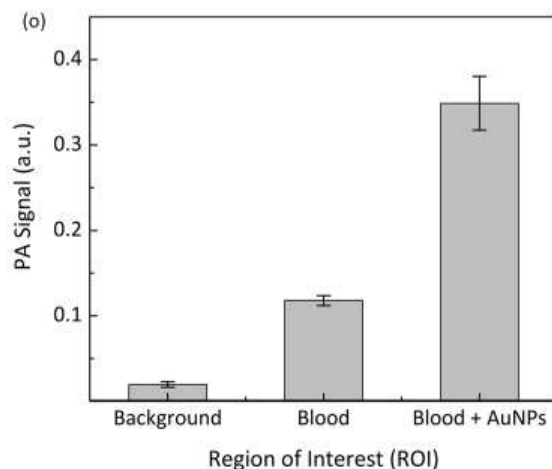
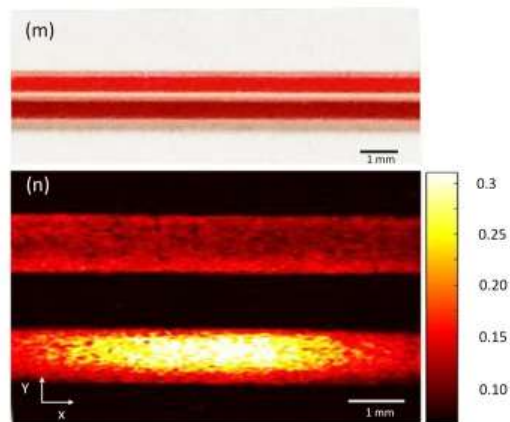
# PA signal with AuNP



(a) Photograph phantoms AuNP various concentrations

(b-k) PA images of phantoms wavelength from 500 to 570nm

(l) PA signal as function of AuNP concentration and wavelength



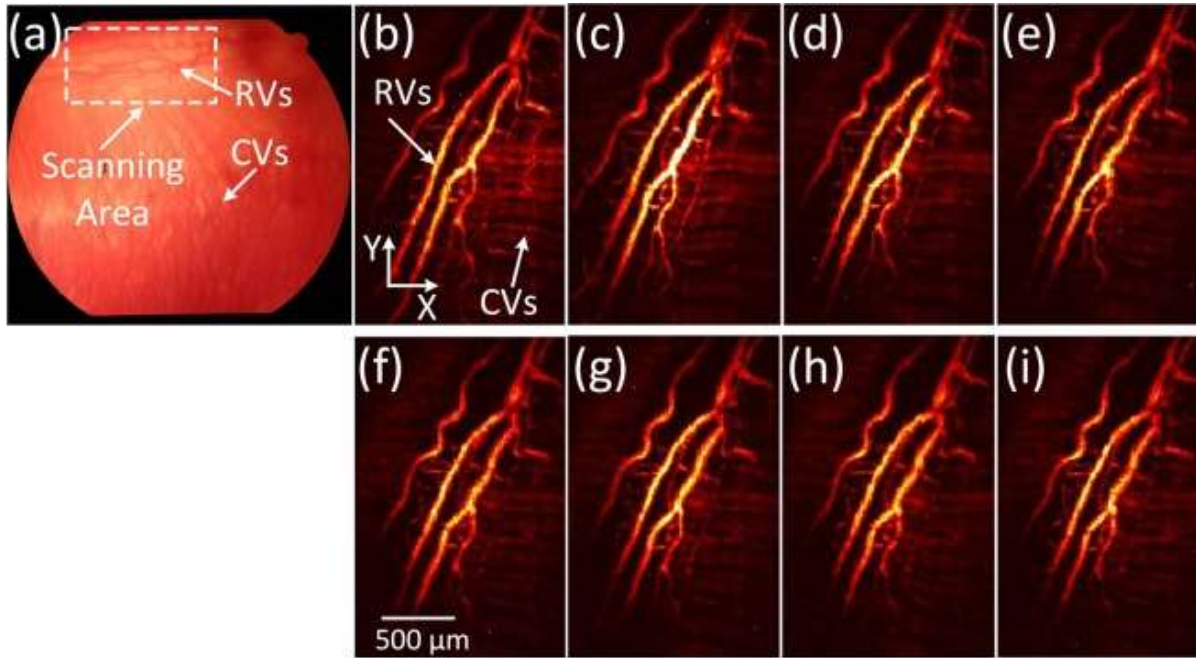
(m) photographs phantoms blood and blood + AuNP 1:1

(n) corresponding PA image

(o) PA blood 3-fold higher background; PA blood + AuNP 17.5 fold higher



# PA imaging retinal vessels with PEG-AuNP 2 mg/mL

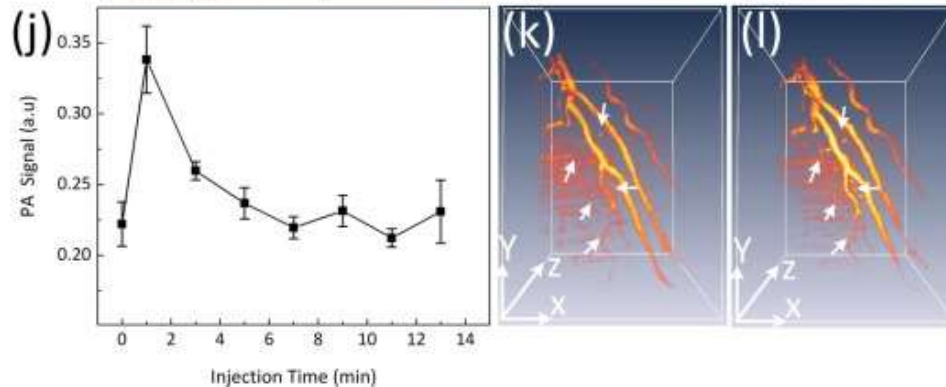


(a) Color fundus image of retina.

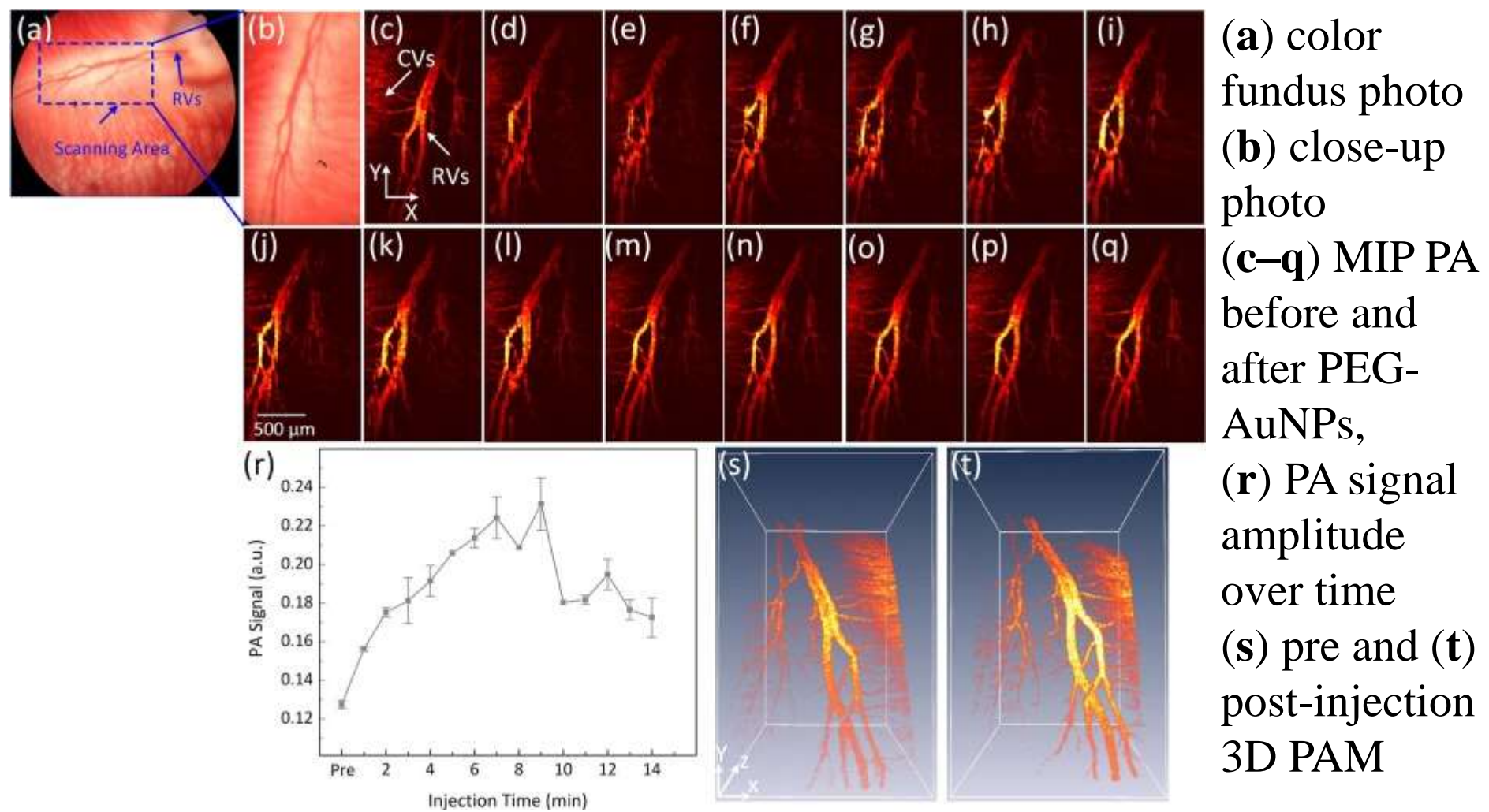
(b–i) PAM images before and after injection of PEG-AuNPs.

(j) PA signal amplitude increase 0.22 to 0.34

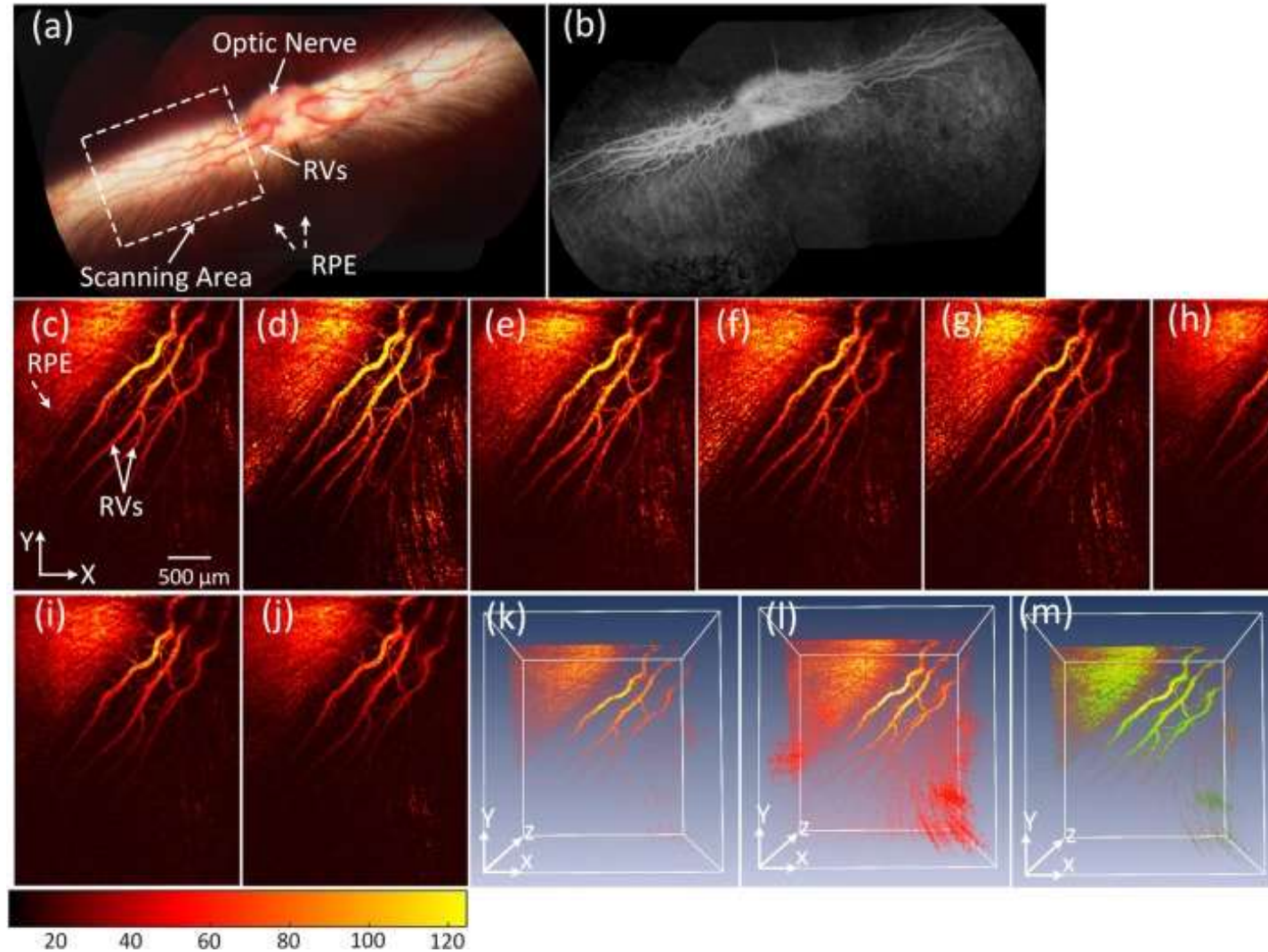
(k) Pre and (l) post injection 3D PAM



# PA imaging retinal vessels with PEG-AuNP 5 mg/mL



# PA retinal vessels in pigmented rabbits with PEG-AuNP 5 mg/mL



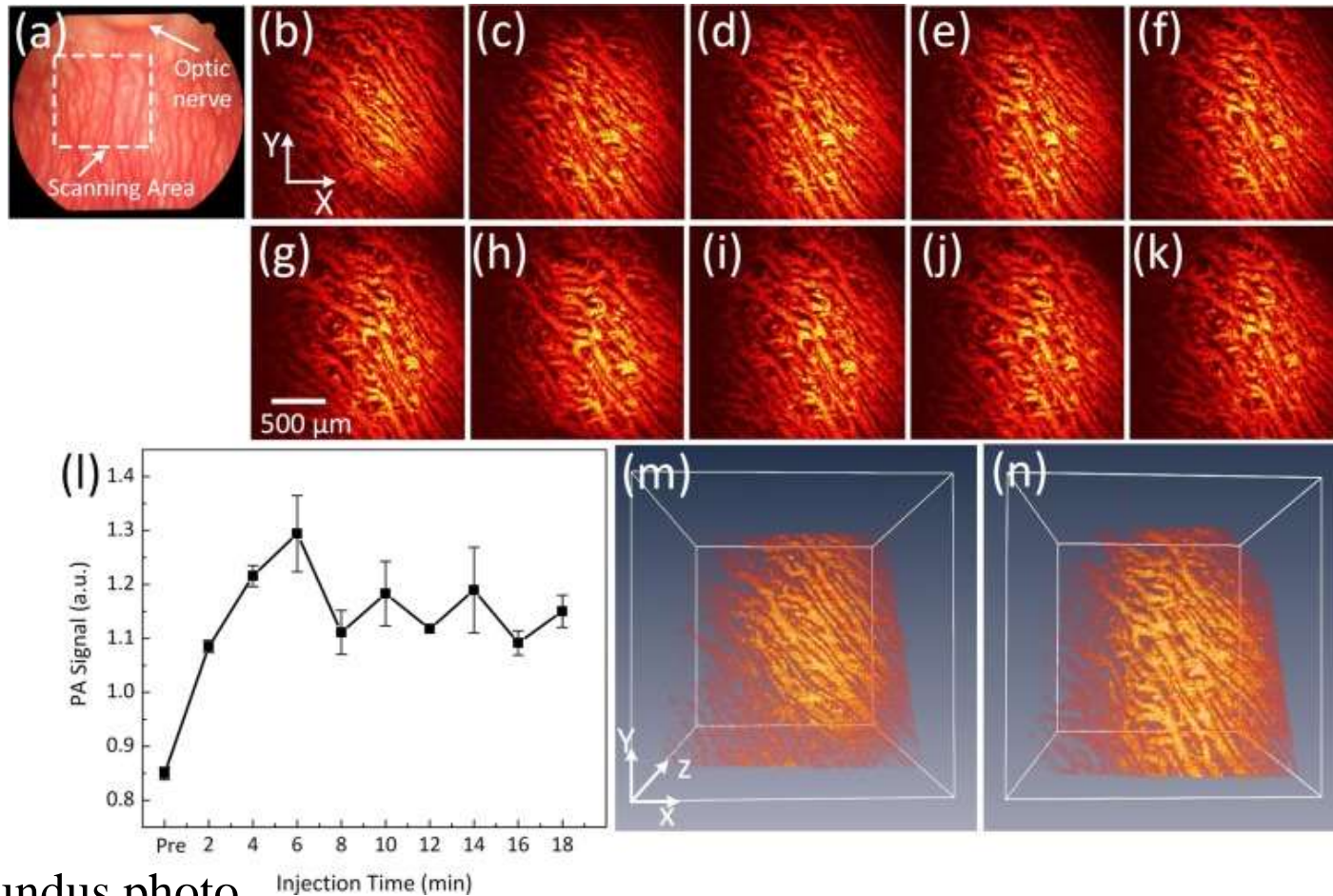
(a) color fundus photo

(b) FA

(c–j) MIP PA before and after PEG-AuNPs, (k) pre and (l) post-injection 3D PAM

(m) Subtraction of post-pre

# PAM imaging choroidal vessels with PEG-AuNP 2 mg/mL



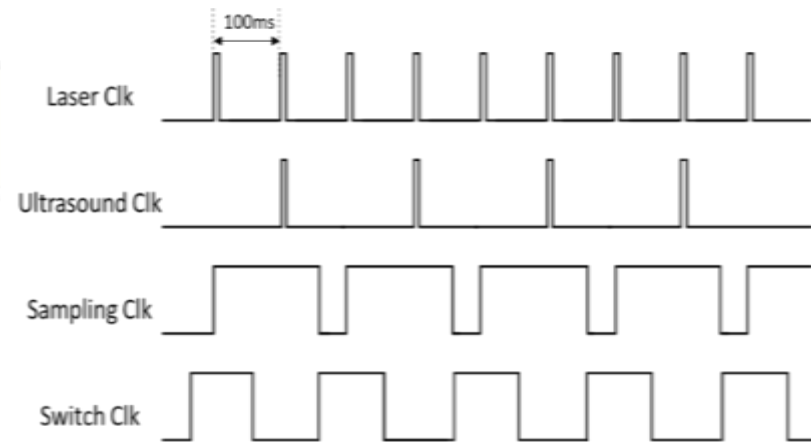
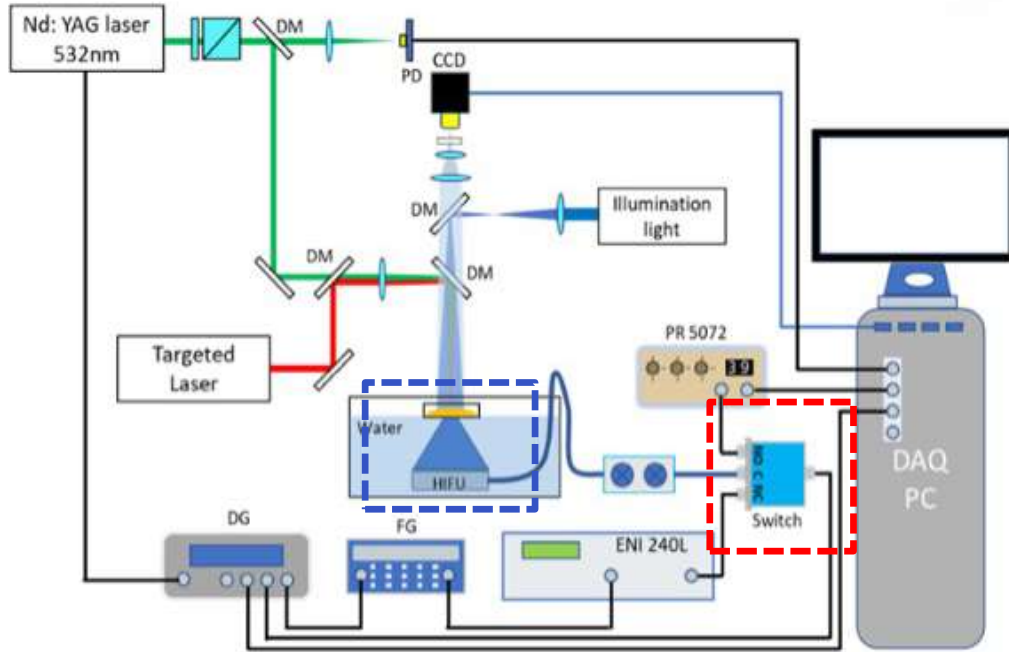
(a) Color fundus photo

(b–k) PAM images before and after injection of PEG-AuNPs.

(l) PA signal amplitude increasing after AuNP

(m) Pre and (n) post injection 3D PAM

# Real-time photoacoustic signal guided therapy



Schematic diagram of real-time PA signal guided PUT system

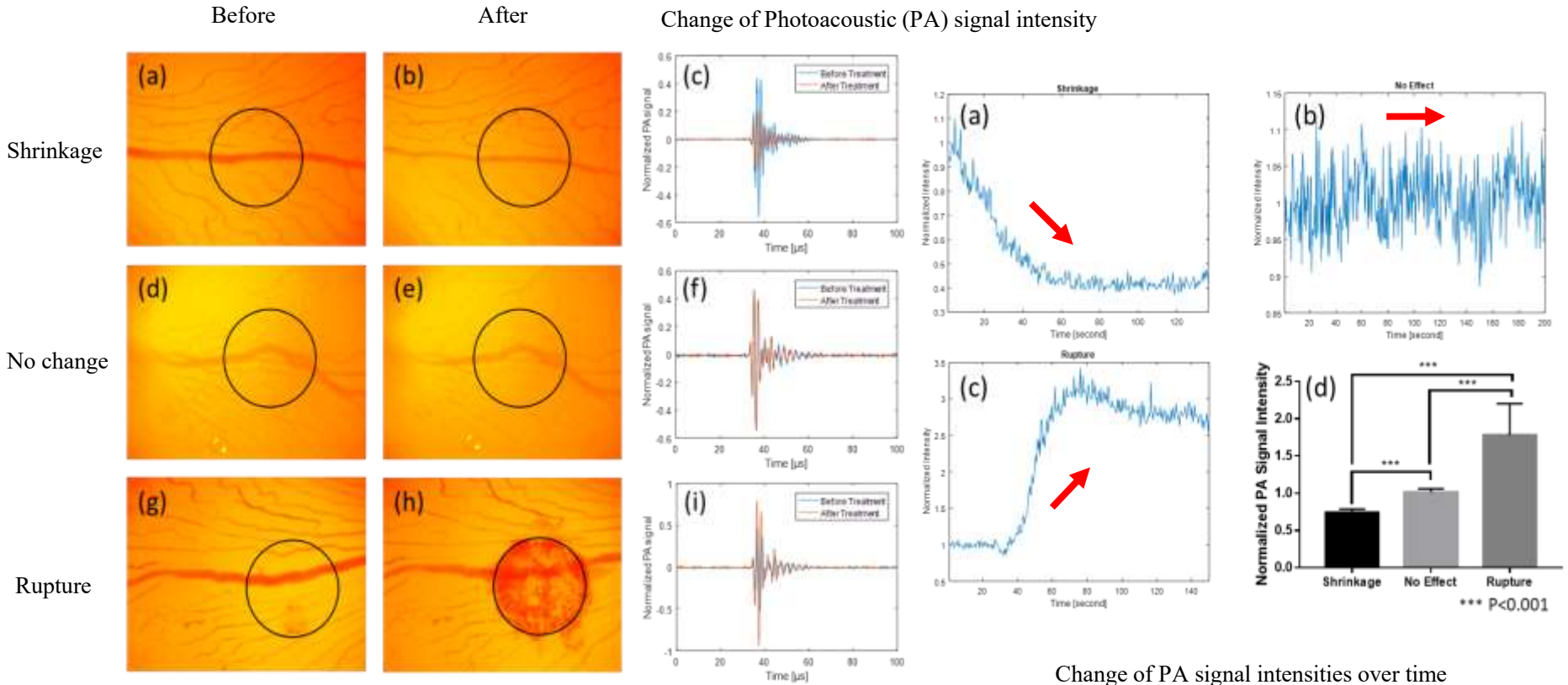
HIFU: High-intensity focused ultrasound

Switch: Pulse/Receiver Switch

DG: Delay generator FG: Function generator

Time sequencing of the system

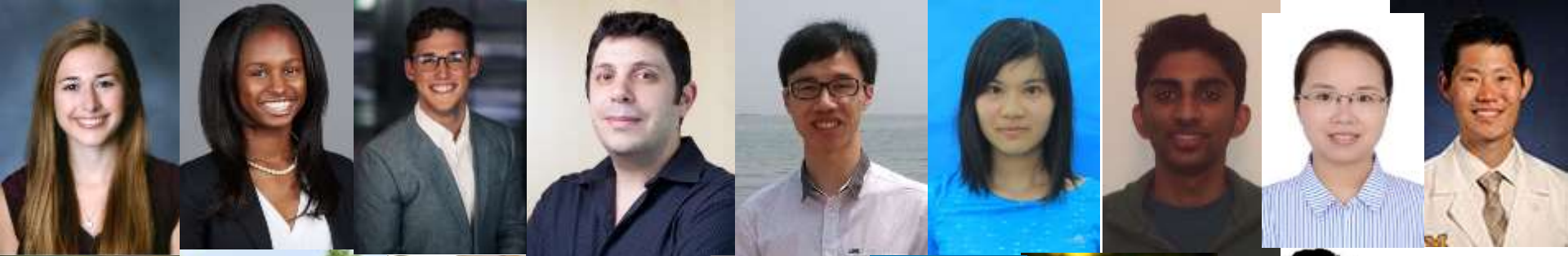
# Real-time photoacoustic signal guided therapy



Change of PA signal intensities over time

# Conclusions

- Photoacoustic imaging is a promising modality to noninvasively image blood distribution with a high depth of penetration (retina and choroid)
- Photoacoustic microscopy can be achieved with a high resolution (2.6  $\mu\text{m}$  lateral resolution)
- PAM can be utilized with a safe laser intensity below the ANSI safety limit
- PAM can visualize blood vessels in human-sized eyes (rabbit)
- PAM can visualize retinal vascular pathology such as retinal neovascularization (like in diabetes) and retinal vein occlusion
- Gold nanoparticles can serve as contrast agents to enhance PAM imaging
- Photoacoustic monitoring of retinal vasculature for automated dosimetry and reproducible burns “Smart laser”



# Paulus Advanced Retinal Imaging and Laser Laboratory



# Retina Division UM





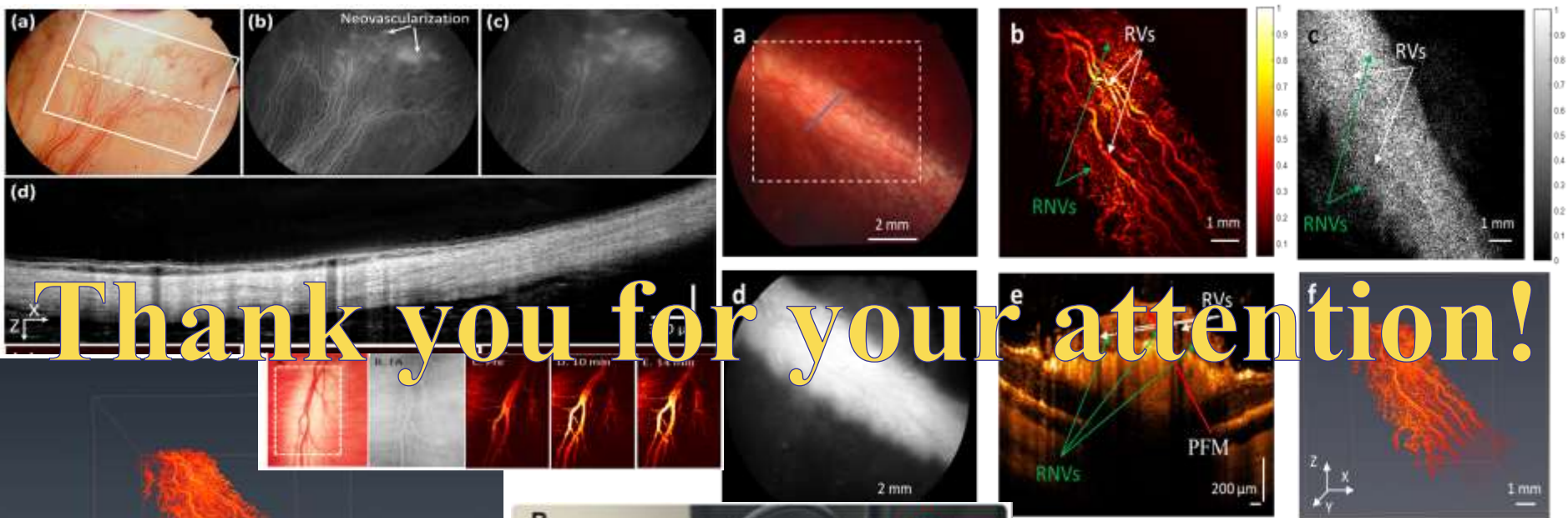
Special thanks



# Funding acknowledgements

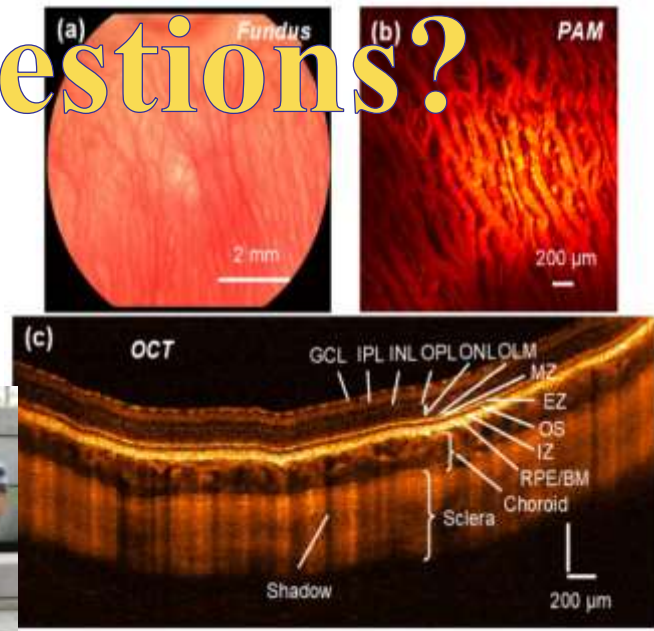
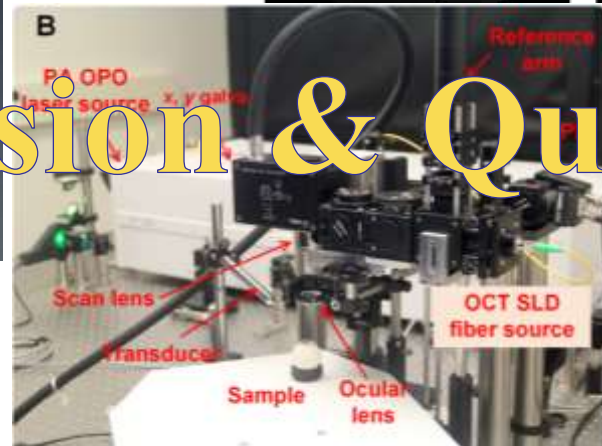
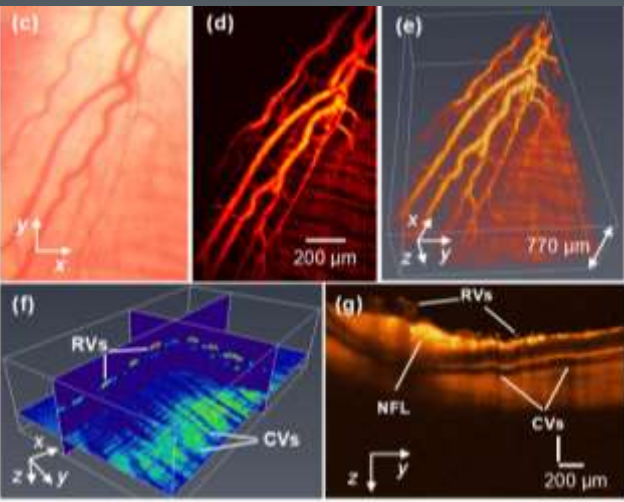
- National Eye Institute 1K08EY027458, 1R01EY029489, 4K12EY022299
- University of Michigan Office of Research Equipment Grant
- Fight for Sight - International Retinal Research Foundation FFSGIA16002
- Research to Prevent Blindness
- Alliance for Vision Research
- Knights Templar Eye Foundation
- University of Michigan Center for Entrepreneurship
- University of Michigan Translational Research and Commercialization for Life Sciences (MTRAC) # N021025
- Core Center for Vision Research funded by P30 EY007003 from the National Eye Institute
- University of Michigan Department of Ophthalmology and Visual Sciences





Thank you for your attention!

Discussion & Questions?



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