

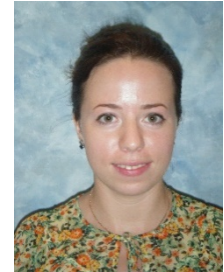
Recent Advances in Tissue Biomechanics Using Dynamic Optical Coherence Elastography

Presented by:



Therapeutic
Laser Applications
Technical Group

Executive Committee



Chair: Elina A. Vitol

Advisor: David R. Busch, UT Southwestern Medical Center at Dallas

Committee Member: Yannis M. Paulus, Assistant Professor, University of Michigan

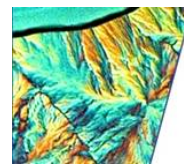
Committee Member: Guido Perrone, Associate Professor, Politecnico di Torino

Committee Member: Felix Fanjul-Velez, Associate Professor, University of Cantabria

Industry Liaison: Robbie Thomas, Photon Force, Ltd.

Social Media Liaison: Mariia Shutova, Texas A&M University

Where to find information about the group



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Therapeutic
Laser Applications
Technical Group

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Therapeutic Laser Applications (BA)

Get Involved

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- Chapters and Sections Map
- Technical Groups
 - Bio-Medical Optics
 - Microscopy and Optical Coherence Tomography (BM)
 - Molecular Probes and Nanobio-Optics (BP)
 - Optical Biosensors (BB)
 - Optical Trapping and Manipulation in Molecular and Cellular Biology (BT)
 - Therapeutic Laser Applications (BA)**
 - Tissue Imaging and Spectroscopy (BS)
 - Photobiomodulation (BL)
 - Fabrication, Design & Instrumentation
 - Information Acquisition, Processing & Display

Therapeutic Laser Applications

This group focuses on the use of lasers in surgery or in other treatments of disease. This includes the use of lasers as surgical tools for tissue cutting, welding, and coagulation, as well as the use of optics to initiate cell-damaging photochemical reactions for the treatment of diseases such as cancer. In addition, optics, spectroscopy, and imaging provide unique tools that may allow real-time diagnostics of the efficacy of clinical procedures. For many of these applications, the development of optical tools for appropriate light delivery, especially for fiber-based or endoscopic delivery to tissues that are not directly accessible, is critical. In addition, this group emphasizes basic science studies of the mechanisms by which light can affect tissue in adverse or therapeutic ways.

Upcoming Technical Group Webinars

Photoacoustic Imaging of the Eye

Hosted By: Therapeutic Laser Applications Technical Group

24 October 2019, 10:00 AM - 11:00 AM

[Register Now](#)

Announcements

If you are a member of the Therapeutic Laser Applications Technical Group and have ideas for activities and initiatives to help engage this community, please [share them with the chair, Elina Vitol](#).

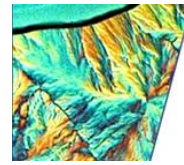
View [OSA Technical Group webinars](#) on-demand at any time or register for any of our upcoming webinars [online](#). Each webinar is an hour long and features a technical presentation on a topic selected by your OSA Technical Groups.

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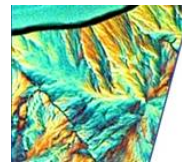
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- Select **Therapeutic Laser Applications** as one of 5 technical groups of interest at your OSA membership account page
- Attend our networking events, webinars and poster sessions
- Join us on LinkedIn and Facebook to keep in touch  
- Look out for emails from the committee about group activities
- Interested in presenting your research? Have ideas for technical group events?
Want to reach out to your fellow group members?
 - Contact us at elina.vitol@gmail.com or TGactivities@osa.org

Recent and upcoming webinars



OSA

Therapeutic
Laser Applications
Technical Group



24 October 2019

Photoacoustic imaging of the eye

Yannis M. Paulus, M.D., F.A.C.S., University of Michigan

Recording is
available from
OSA website

https://www.osa.org/en-us/get_involved/technical_groups/technical_group_webinars/#ondemand

21 January 2020, 11am EST*

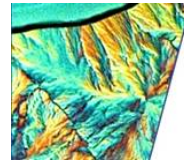
Thermomechanical effect of infrared laser for cartilage regeneration

Yulia M. Alexandrovskaya, PhD

Institute of Photon Technologies, Federal Scientific Research
Centre “Crystallography and Photonics” of the Russian
Academy of Sciences (RAS)

Registration will
open in December

Welcome to today's webinar!



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RECENT ADVANCES IN TISSUE BIOMECHANICS USING DYNAMIC OPTICAL COHERENCE ELASTOGRAPHY

Kirill V. Larin, Ph.D.
Professor, Department of Biomedical Engineering
University of Houston

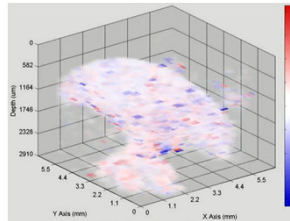
November 1, 2019

Recent Advances in Tissue Biomechanics Using Dynamic Optical Coherence Elastography

Kirill V. Larin

Professor
Fellow SPIE, Fellow OSA

University of Houston

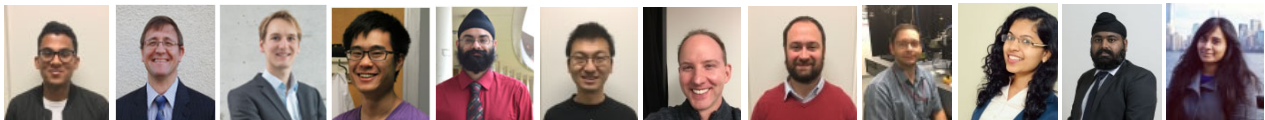


Major Projects in Biomedical Optics Lab

Optical
Elastography/Biomechanics

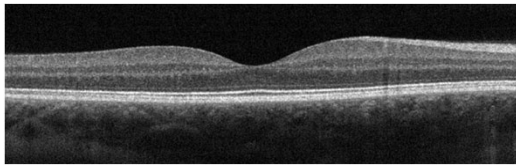


Developmental
Biology



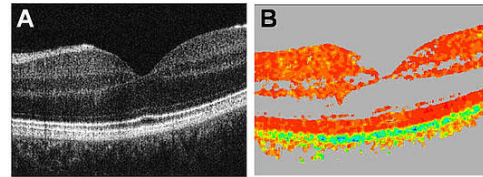
Major Breakthroughs in OCT Functional and Contrast Enhanced Imaging

Morphological Imaging



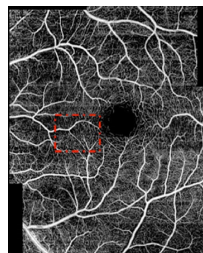
Introduced by Fujimoto et al in 1991

Polarization sensitive OCT



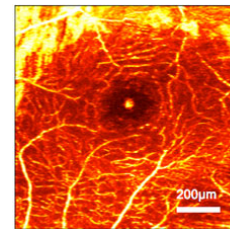
Hitzenberger, et al. Optics Express, 2001

OCT Angiography

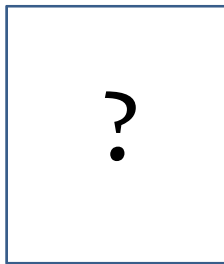


R. Wang Lab, UW

Doppler OCT

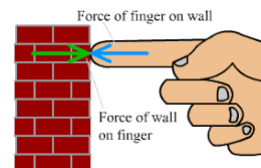
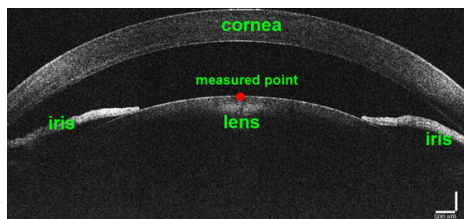


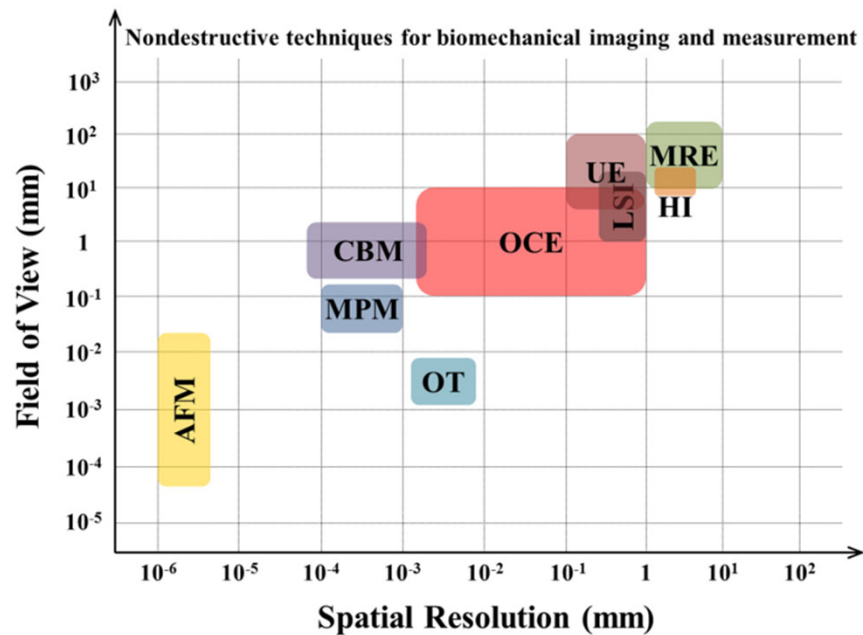
Leitgeb, et al. Optics Express, 2003



Optical Coherence Elastography:

OCT + Force





S. Wang and K. Larin, J of Biophotonics, (2015)

Optical Elastography and Tissue Biomechanics

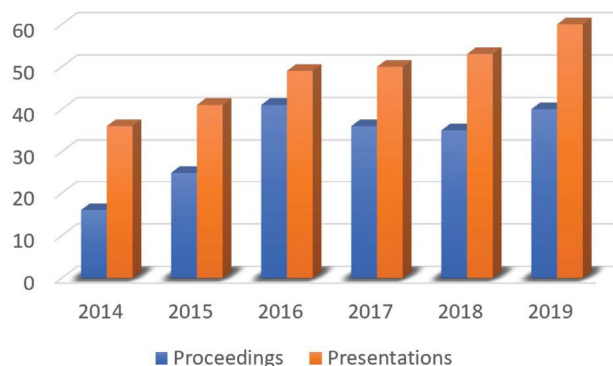
Photonics West, 1 - 6 February 2020

Conference Chairs

- Kirill V. Larin, Univ. of Houston (United States)
- Giuliano Scarcelli, Univ. of Maryland, College Park (United States)

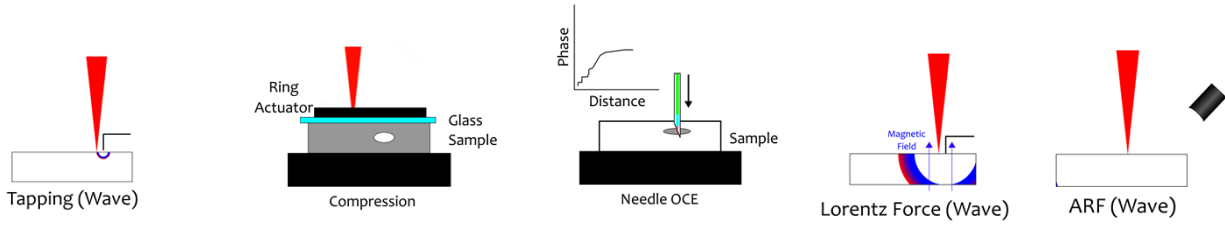
Program Committee

- Steven G. Adie, Cornell Univ. (United States)
- Albert Claude Boccara, Institut Langevin Ondes et Images (France)
- Brett E. Bouma, Wellman Ctr. for Photomedicine (United States)
- Stefan Catheline, Institut National de la Santé et de la Recherche Médicale (France)
- Zhongping Chen, Beckman Laser Institute and Medical Clinic (United States)
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- Kandice Tanner, National Cancer Institute (United States)
- Peter Török, Imperial College London (United Kingdom)
- Ruikang K. Wang, Univ. of Washington (United States)
- Tianshi Wang, Erasmus MC (Netherlands)
- Vladislav V. Yakovlev, Texas A&M Univ. (United States)
- Seok Hyun A. Yun, Wellman Ctr. for Photomedicine (United States)
- Vladimir Y. Zaitsev, Institute of Applied Physics of the RAS (Russian Federation)
- Qifa Zhou, The Univ. of Southern California (United States)

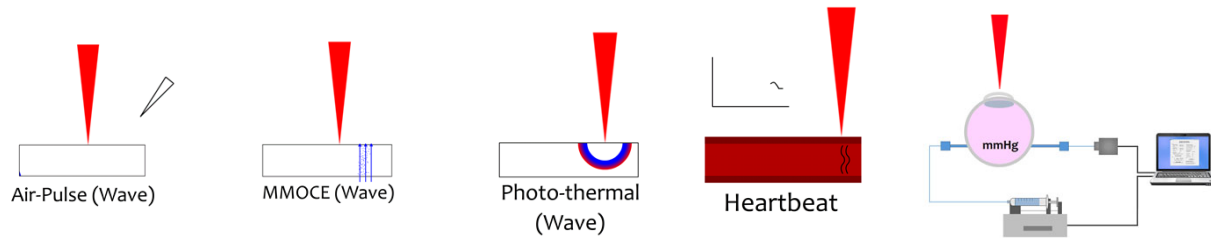


Excitation Methods in OCE

Contact



Non-Contact



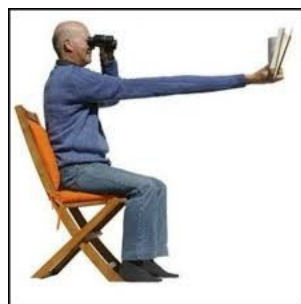
K. Larin and D. Sampson, Biomed Optics Express, 2017

OCE in Ophthalmology

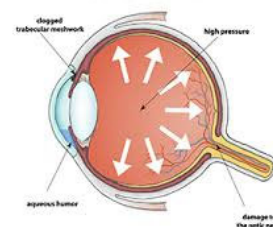
Keratoconus



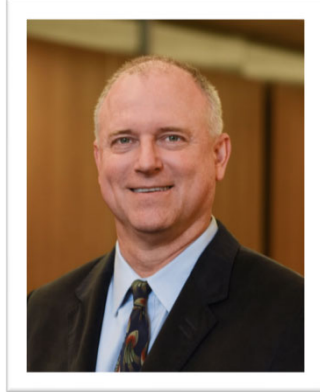
Presbyopia



GLAUCOMA



Biomechanics of the Cornea

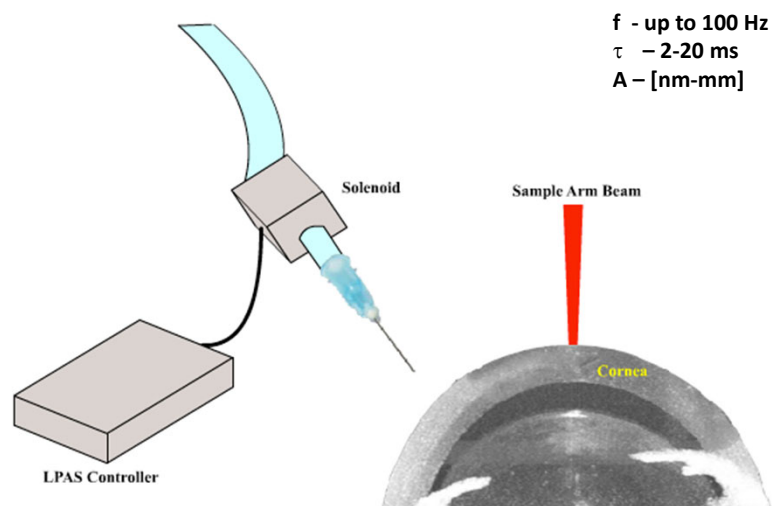


Michael Twa, Ph.D., O.D.
Professor, Dean of the College of
Optometry,
University of Houston

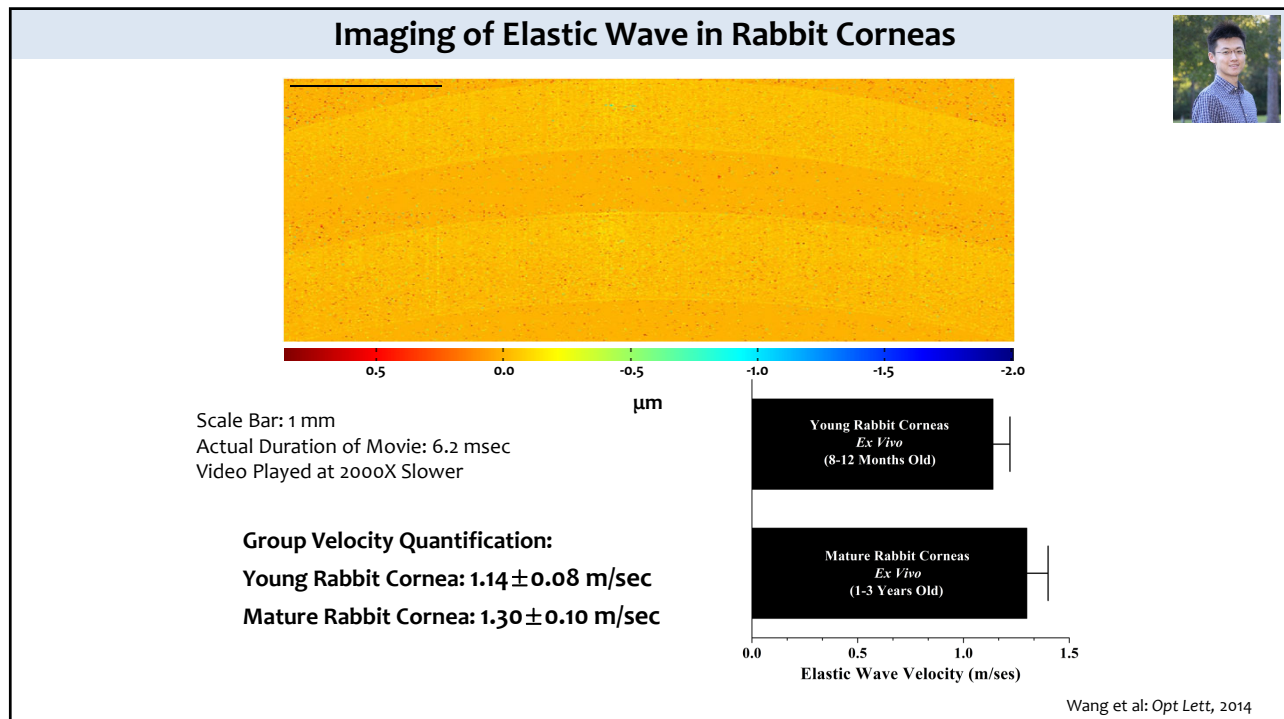
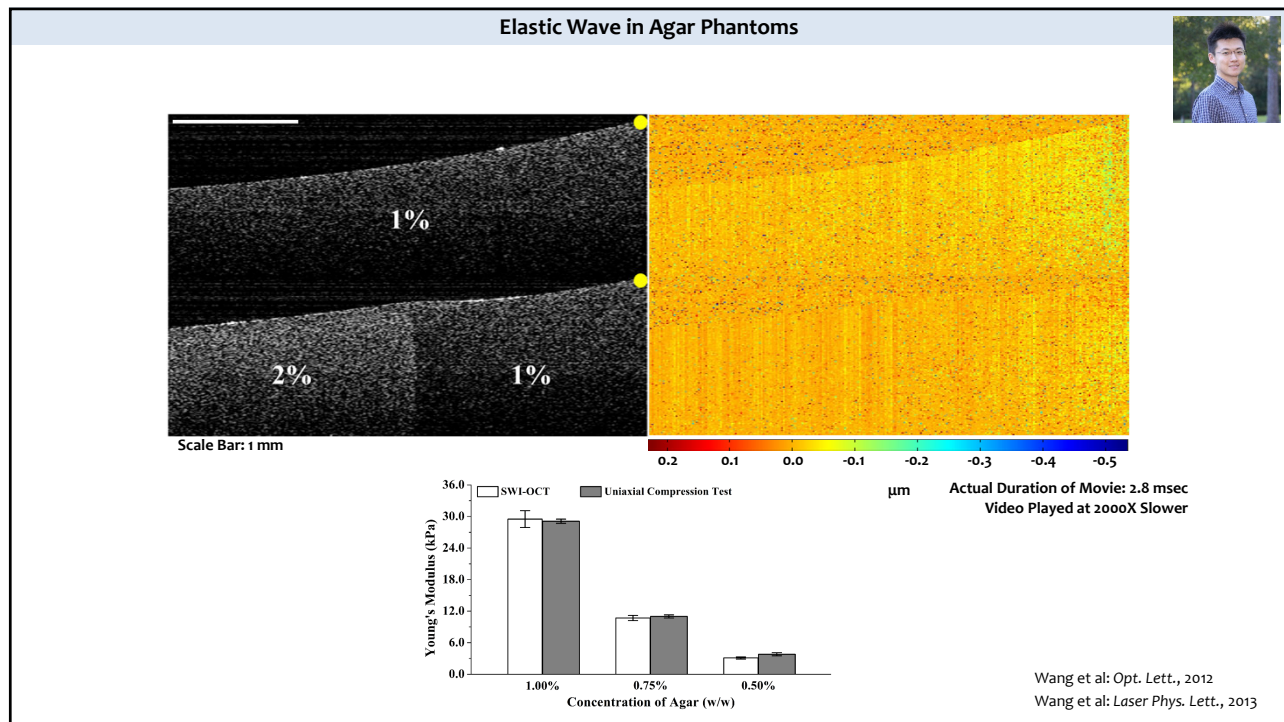


Stanislav Emelianov, Ph.D.
Professor, Department of
Biomedical Engineering,
Georgia Tech

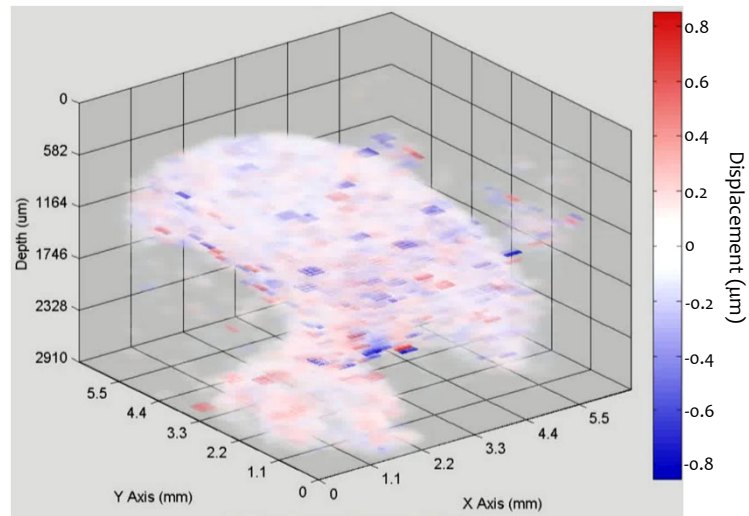
Excitation Method: Focused Air Puff



Wang et al: Opt. Lett., 2012
Wang et al: Laser Phys. Lett., 2013

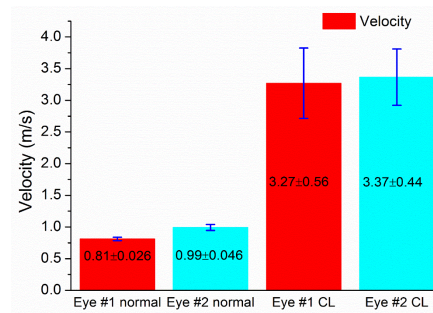
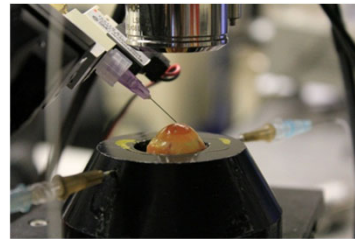
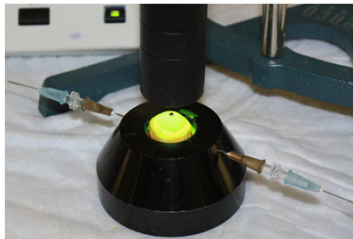
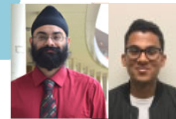


3D Elastic Wave Imaging



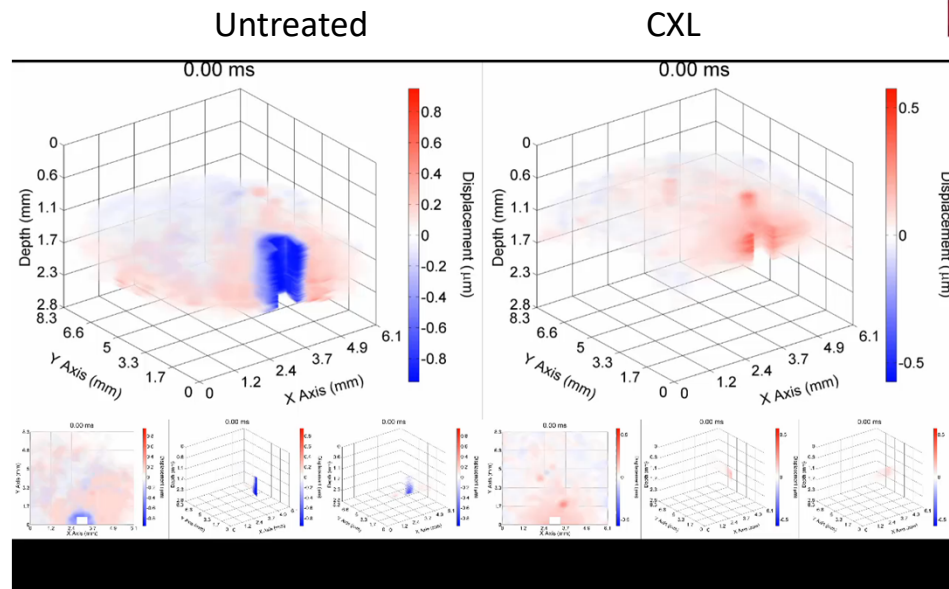
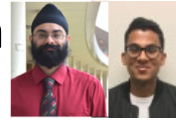
Singh et al: IEEE JSTQE, 2016

CXL

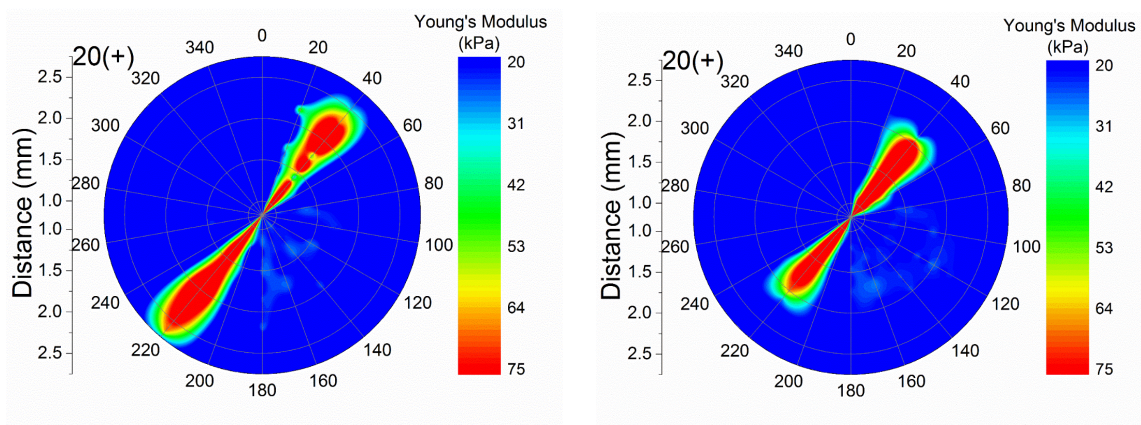
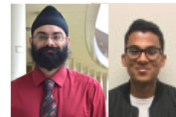


Vantipalli et al, IOVS, 2017

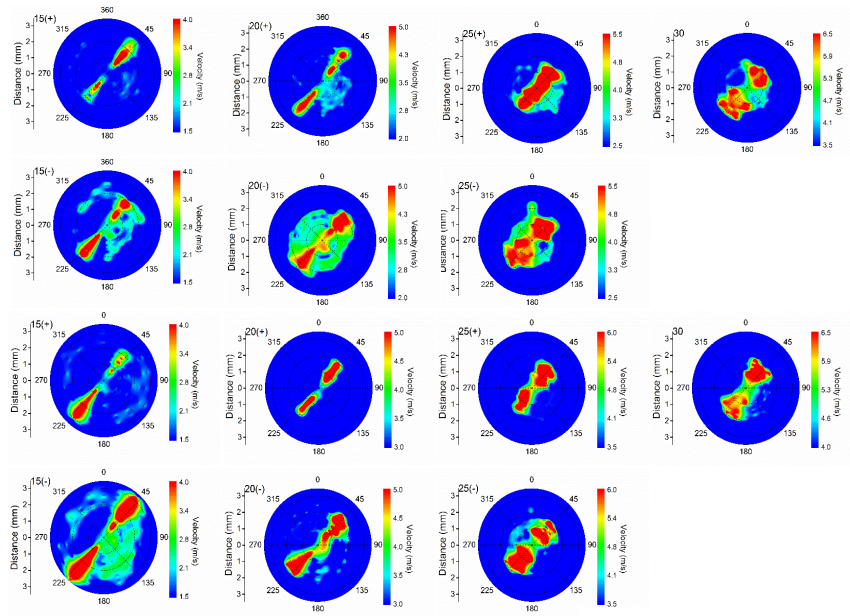
3D elastic wave propagation in cornea



Corneal Anisotropy

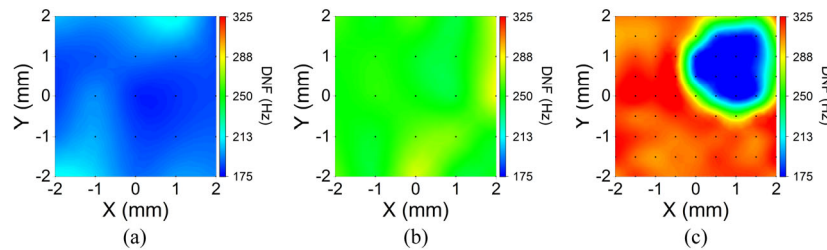
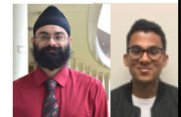
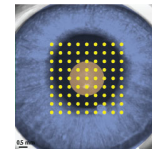


Corneal Anisotropy



Singh et al: *J Refract Surg*, 2016
Singh et al: *Biomed Optics Exp*, 2017

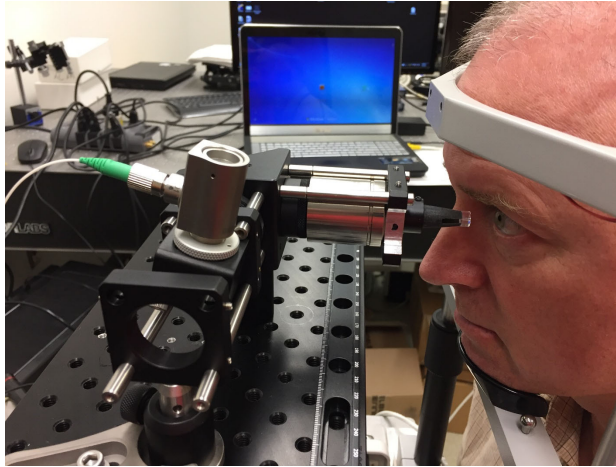
Customized CLX



Next: corneal reshaping (*experimenting*)

Singh et al: *J Biomed Optics*, 2017

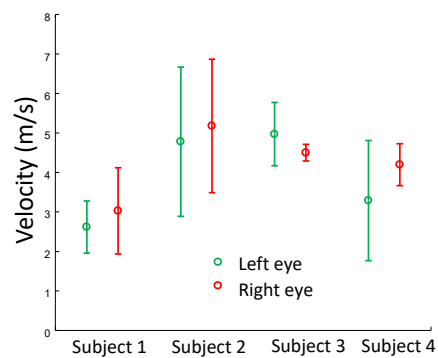
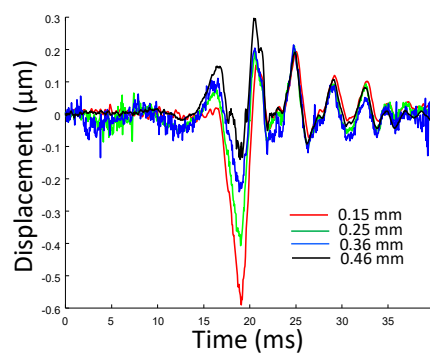
Pilot Human Studies



Twa et al, IOVS, 2017

Human Corneal OCE *in vivo*

n=11



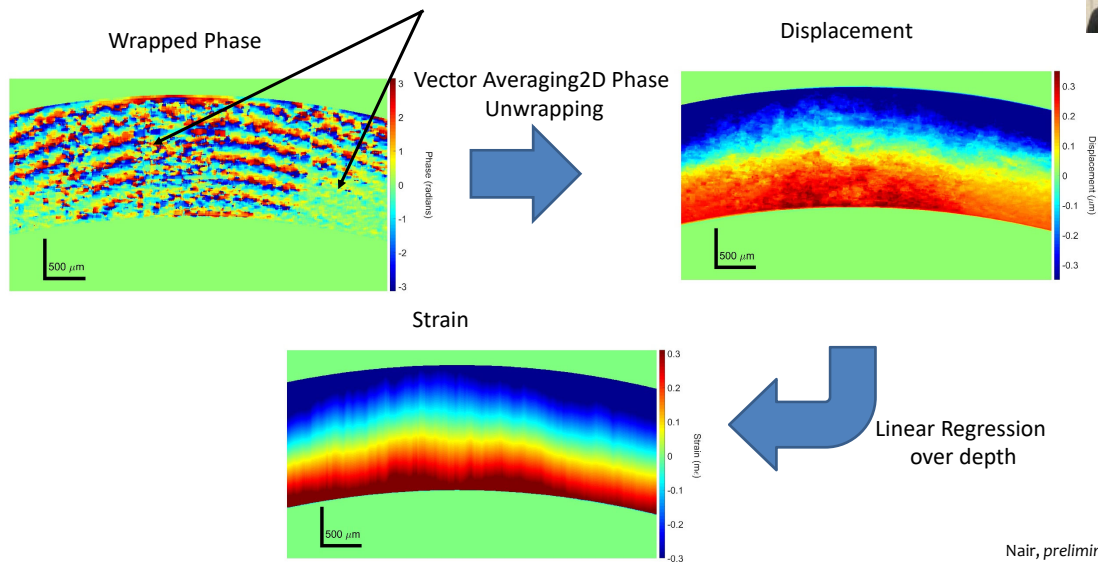
Twa et al, presented at ARVO, 2019

Can we assess corneal biomechanics without **any** external excitation?

Can we assess corneal biomechanics without **any** external excitation?

Heartbeat OCE

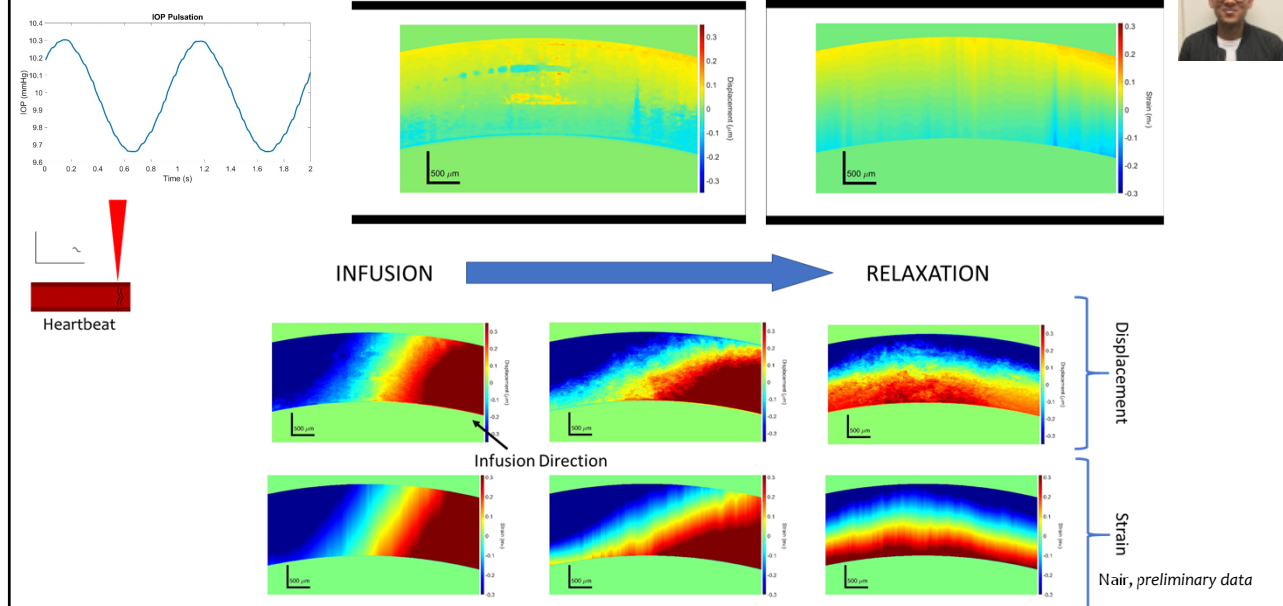
Vector Averaging reduces noise



Nair, preliminary data

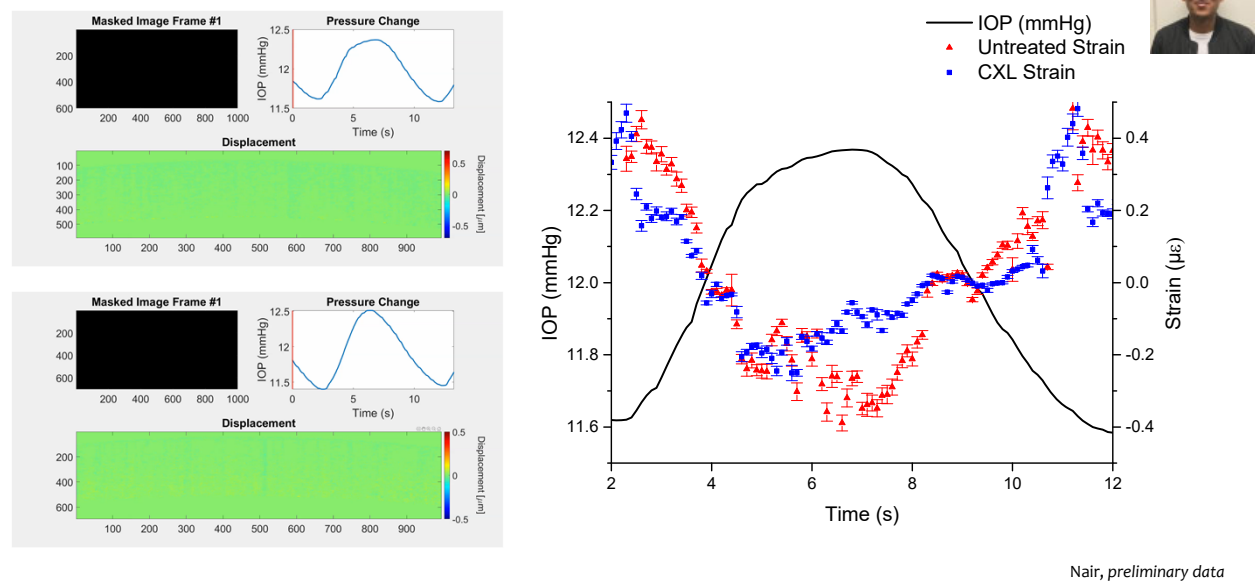
Can we assess corneal biomechanics without **any** external excitation?

Heartbeat OCE



Can we assess corneal biomechanics without **any** external excitation?

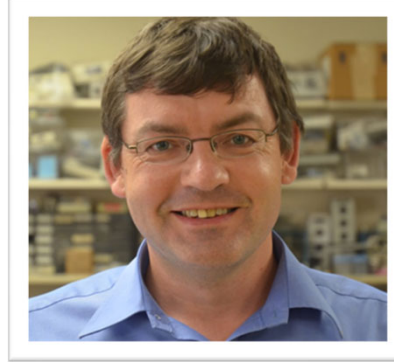
Heartbeat OCE



Biomechanics of the Lens

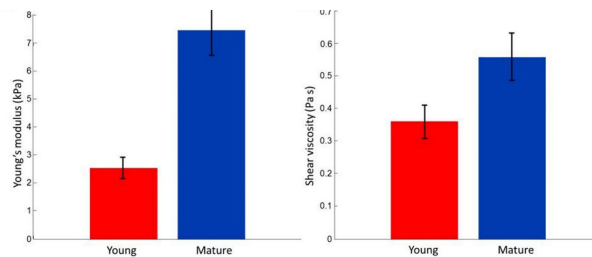
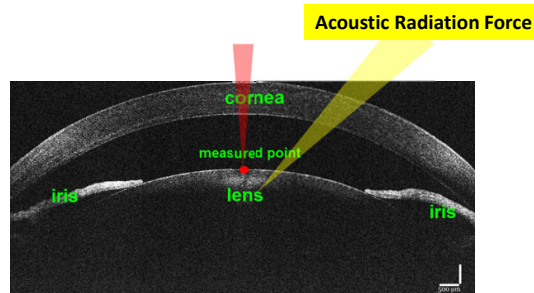


Giuliano Scarcelli, Ph.D.
Assistant Professor, Fischell
Department of Bioengineering,
University of Maryland



Fabrice Manns, Ph.D.
Professor, Department of
Biomedical Engineering,
University of Miami and Bascom
Palmer Eye Institute

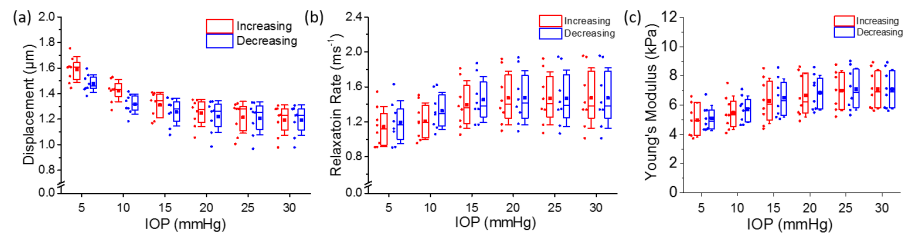
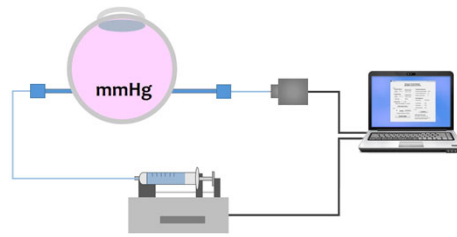
Crystalline Lens



Wu, C., et. al. Investigative ophthalmology & visual science, 56(2), 2015

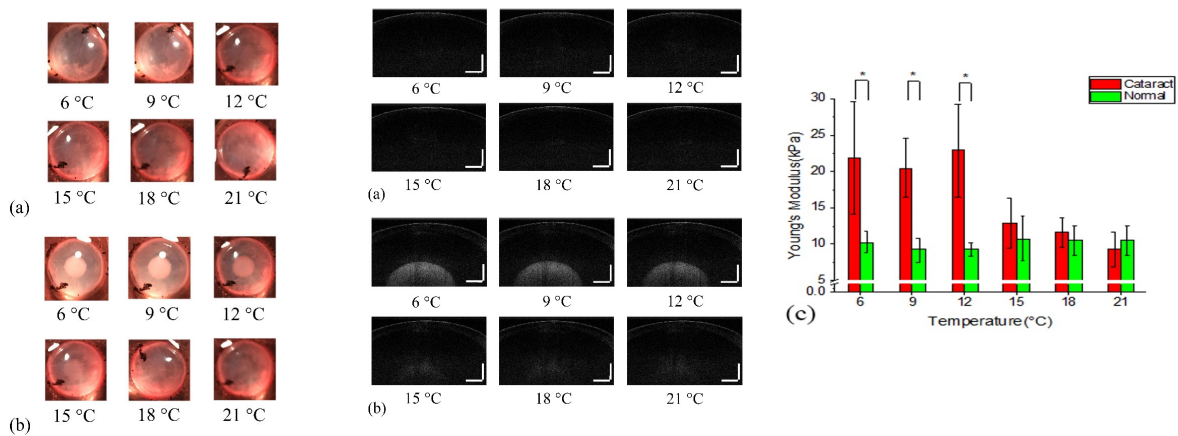
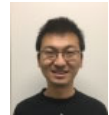


Crystalline Lens: function of IOP



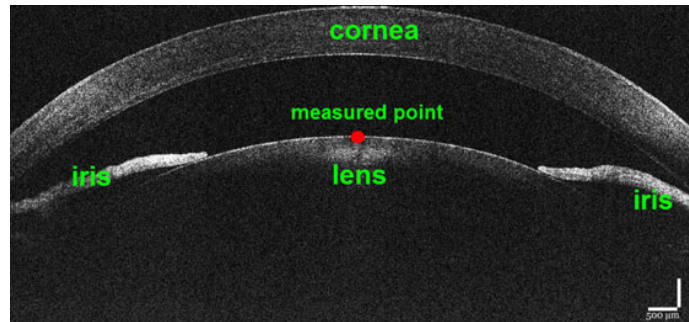
Wu, C., et. al. Quantum Electron, 2019

Crystalline Lens: function of cataract

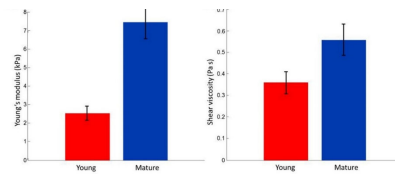
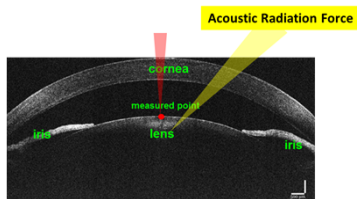


Zhang, H., et. al. JBO, 2019

Crystalline Lens: can we measure mechanical properties in 3D?



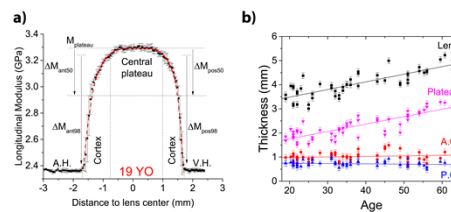
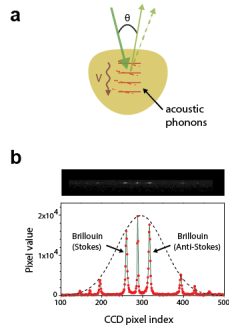
OCE:



Wu, C., et. al. Investigative ophthalmology & visual science, 56(2), 2015

Pros: Quantitative
Cons: Lens surface only

Brillouin Spectroscopy:

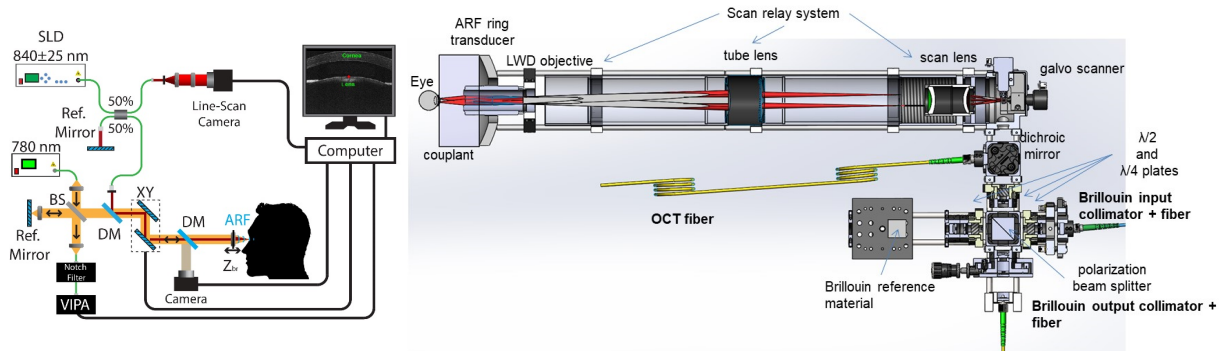


Scarcelli Lab, UMD

Pros: 3D ready
Cons: Qualitative

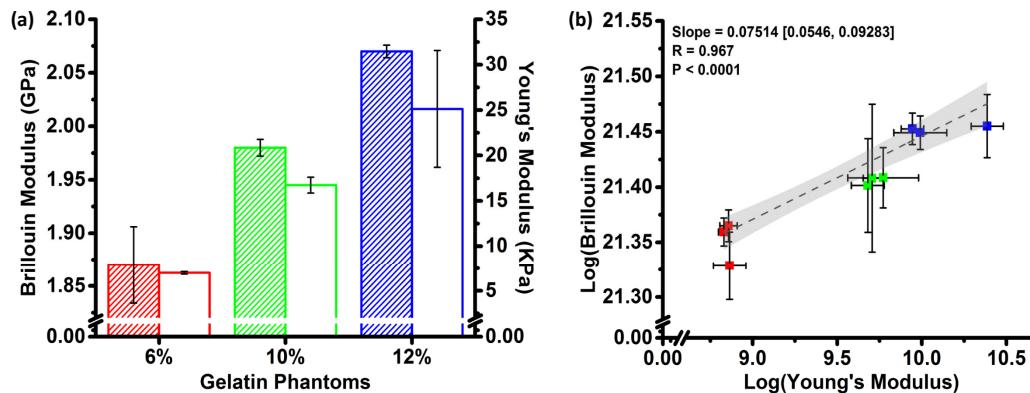
OCE + Brillouin Spectroscopy

For truly **quantitative** elastography of the Lens **in 3D**



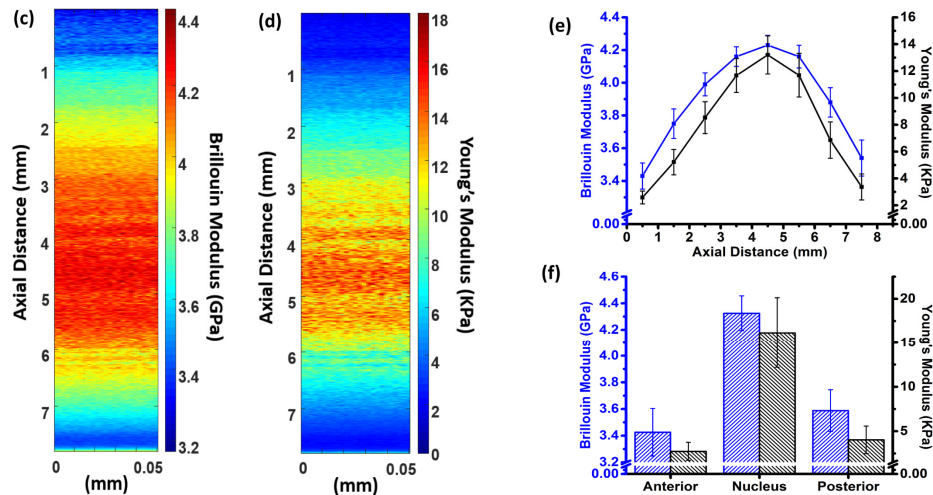
Can we **quantify** mechanical properties of the lens in 3D?

OCE + Brillouin Spectroscopy



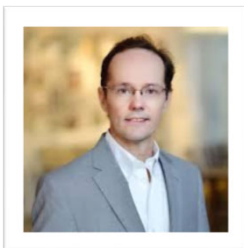
Ambekar et al (in preparation)

Can we **quantify** mechanical properties of the lens in 3D? OCE + Brillouin Spectroscopy

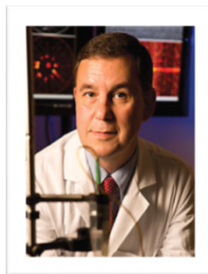


Ambekar et al (in preparation)

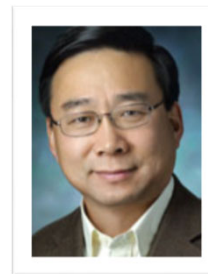
Biomechanics of the Cardiac Muscle



James Martin, M.D., Ph.D.
Professor, Department of
Physiology and Biophysics,
Baylor College of Medicine



Stanislav Emelianov, Ph.D.
Professor, Department of
Biomedical Engineering,
Georgia Tech

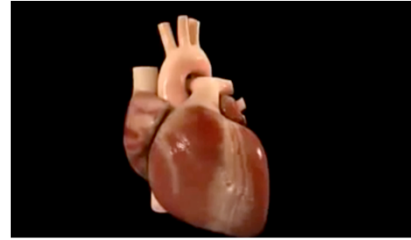


Xingde Li, Ph.D.
Professor, Department of
Biomedical Engineering,
John Hopkins University

Mechanical Properties of Cardiac Muscle

Myocardium and the beating of the heart

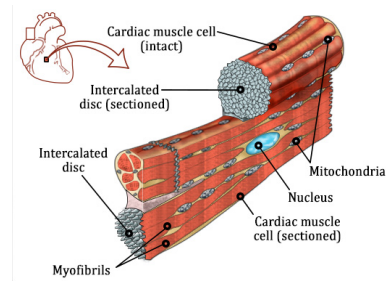
- ❖ Dynamic process: relaxation and contract;
- ❖ The elasticity of the cardiac muscle determines the normal function of the heart;
- ❖ Spatial heterogeneity: mechanical strength of the muscle fiber and between the muscle fiber.



Recovery of the heart after ischemic disease

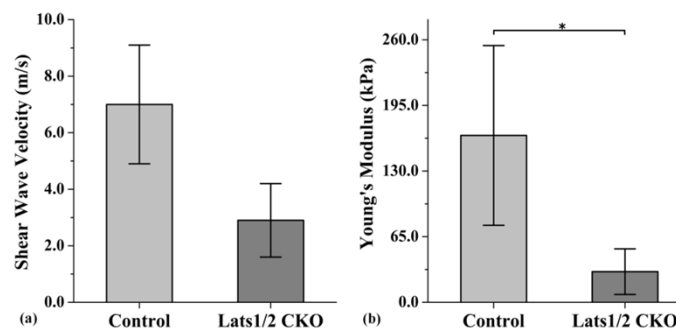
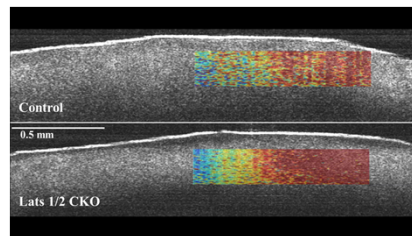
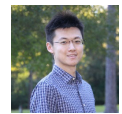
- ❖ Self-regeneration of cardiomyocytes.

Stiffness of the cardiac muscle increases with age



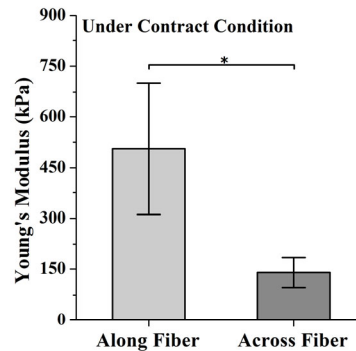
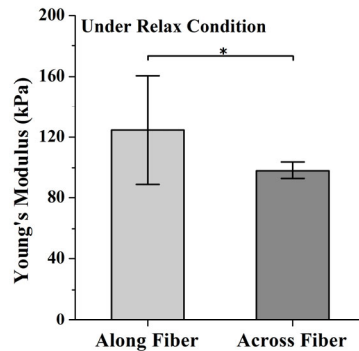
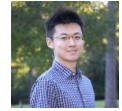
From Buzzle.com
<http://www.buzzle.com/articles/cardiac-muscle-structure.html>

Biomechanics of the Cardiac Muscle



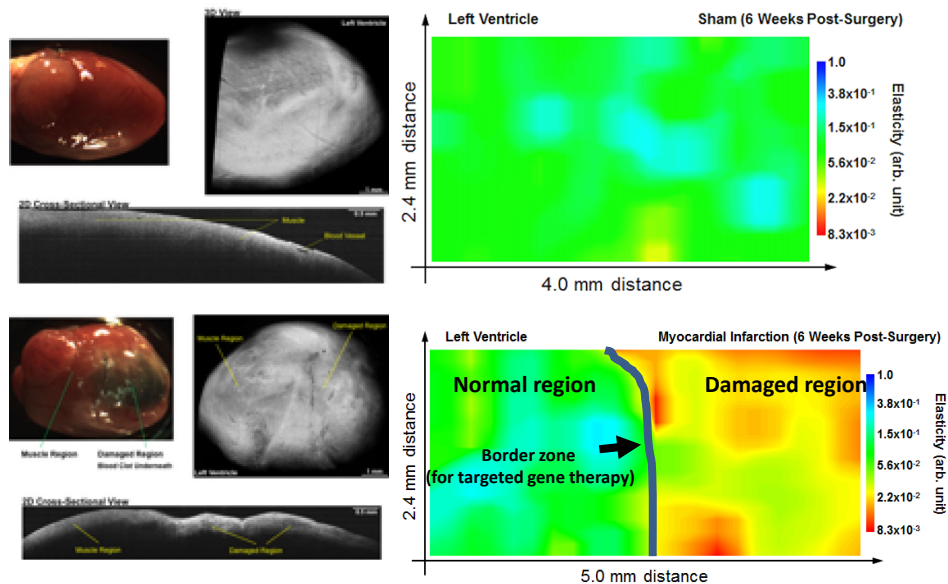
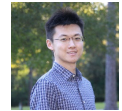
Wang et al: Biomedical Optics Express, 2014

Biomechanics of the Cardiac Muscle



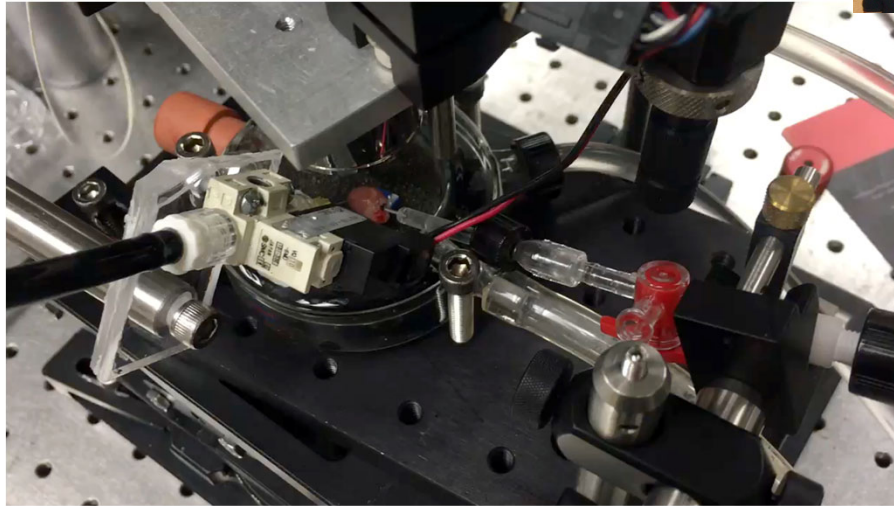
Wang, et al: (unpublished)

Biomechanics of the Cardiac Muscle during MI



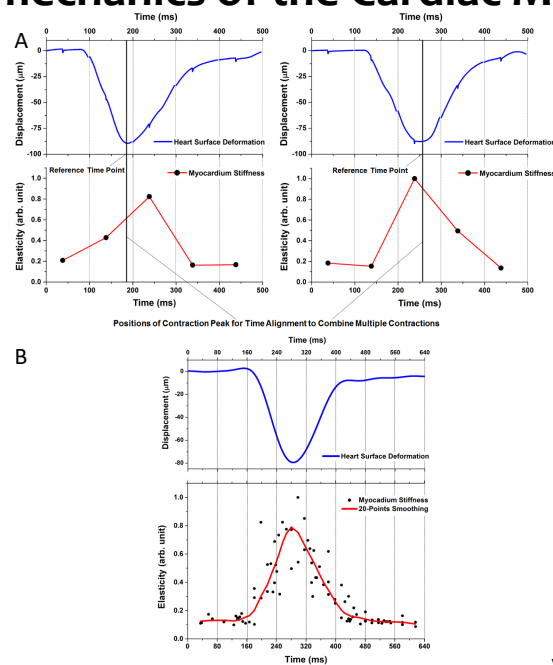
Wang, et al: Biomed Opt Express, 2018

Biomechanics of the Cardiac Muscle



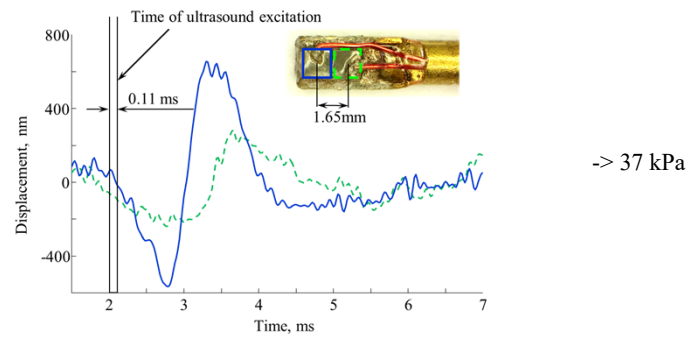
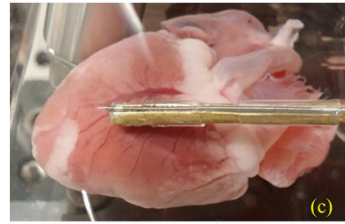
Lopez, Wang, et al, et al: unpublished

Biomechanics of the Cardiac Muscle



Wang, et al: (unpublished)

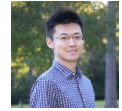
Biomechanics of the Cardiac Muscle: toward *in vivo*



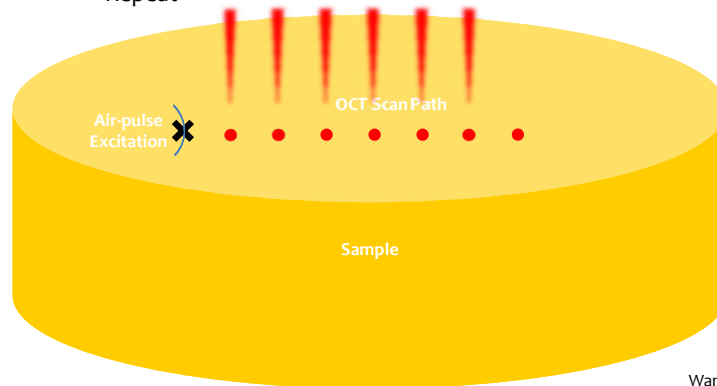
Karpiouk, et al: *J Biomed Optics*, 2018

Pushing limits of the technology

M-B mode OCE Imaging Methodology



- Synchronize successive M-mode images with focused air-pulse [1]
 - Excite, acquire M-mode image
 - Move to next position
 - Excite, acquire M-mode image
 - Repeat

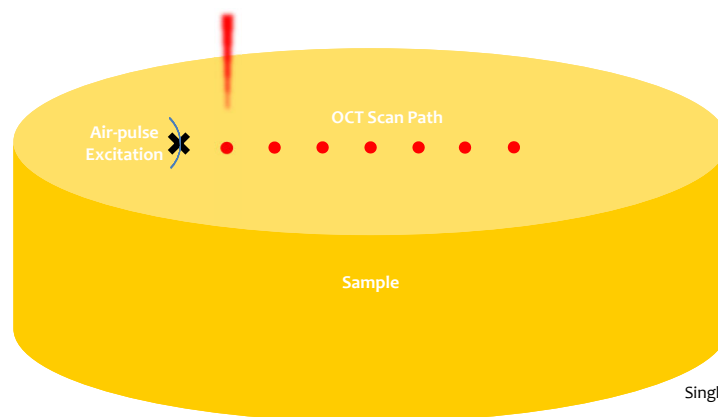


Wang et al: *Optics Lett*, 2014

B-M mode OCE Imaging Methodology

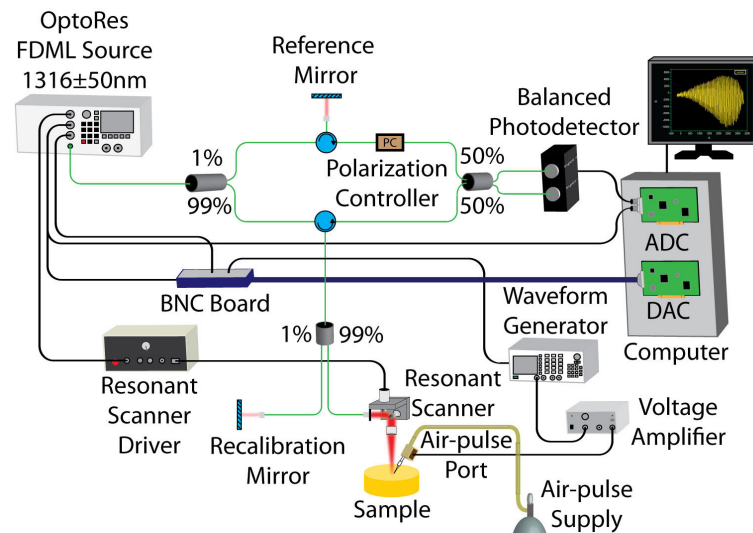


- Fast B-mode imaging **during** wave propagation



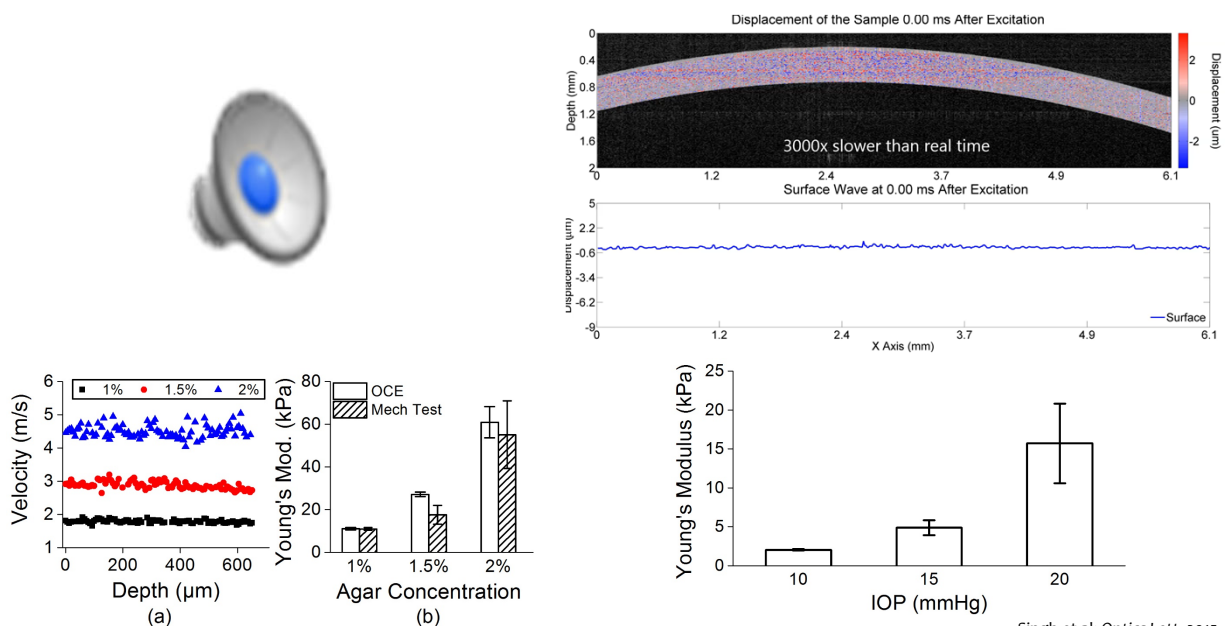
Singh et al: *Optics Lett*, 2015

Real-Time OCE: 1.5 MHz A-scan rate SSOCT



Singh et al: Optics Lett, 2015

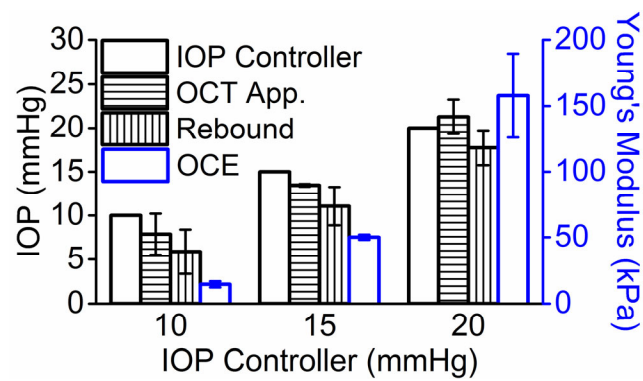
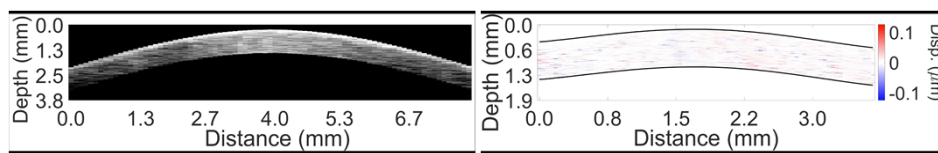
Real-Time OCE: 1.5 MHz A-scan rate SSOCT



Singh et al: Optics Lett, 2015

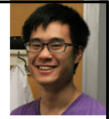
Can we measure **IOP** and **corneal stiffness** using **ONE** instrument?

Intraocular Pressure and Cornea stiffness in ONE instrument **Applanation-OCE**

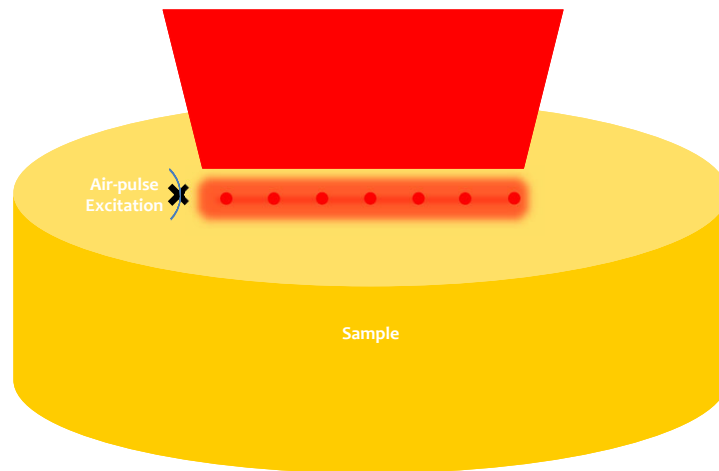


Singh et al: J Biomed Opt, 2017

Line-Field OCE Imaging Methodology

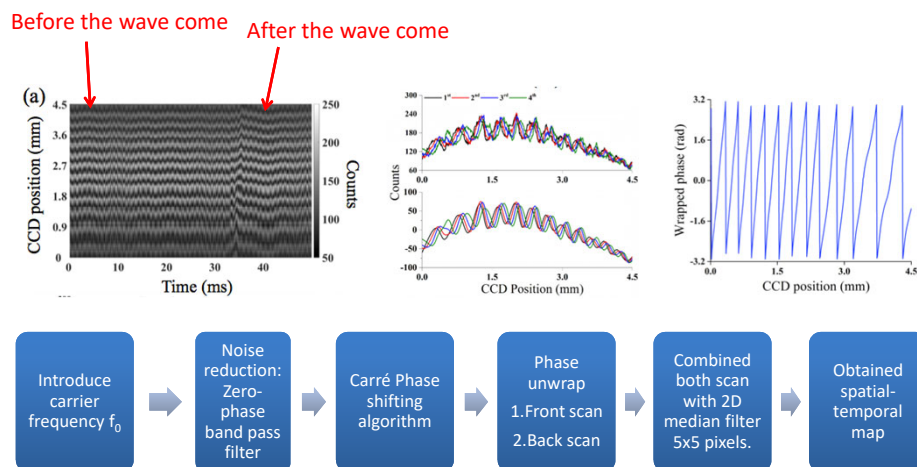
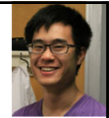


- Line imaging **during** wave propagation



Liu et al: Biomed. Opt. Express, 2017
Liu et al: Biomed. Opt. Express, 2016

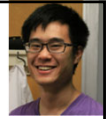
Line-field OCT methodology



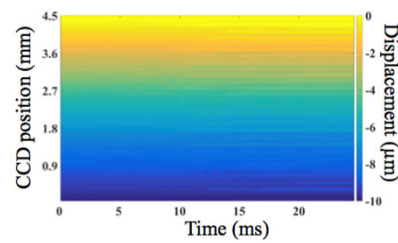
Equivalent to **72 MHz** acquisition rate – fastest demonstrated speed in elastography

Liu et al: Biomed. Opt. Express, 7(8), 2016

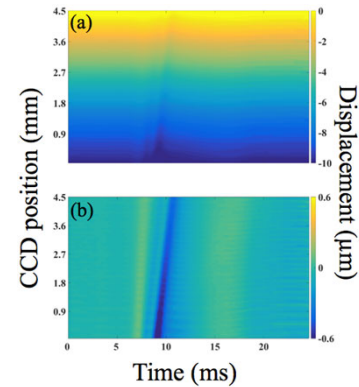
Spatial-Temporal map of an agar phantom



Before the elastic wave



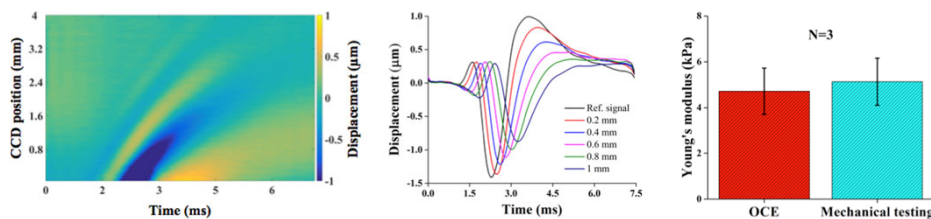
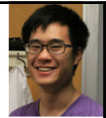
Capture of the elastic wave



Liu et al: *Biomed. Opt. Express*, 7(8), 2016

Line-Field OCE

Captured elastic wave propagation in chicken breast (N=3)

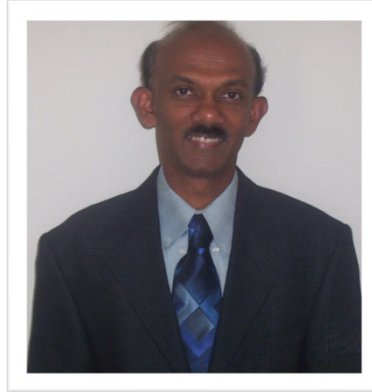


Liu et al: *Biomed. Opt. Express*, 7(8), 2016

The first clinical studies: Systemic Sclerosis



Shervin Assassi, M.D., M.S.
Associate Professor,
Rheumatology And Clinical
Immunogenetics,
UT Health



Chandra Mohan, M.D., Ph.D.
Professor, Department of
Biomedical Engineering,
University of Houston

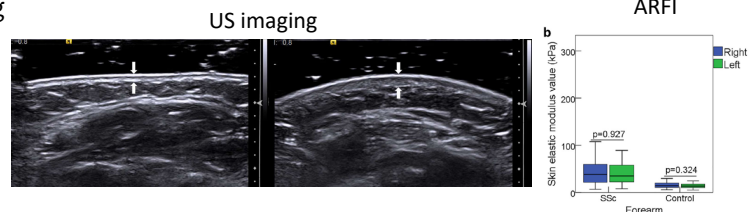
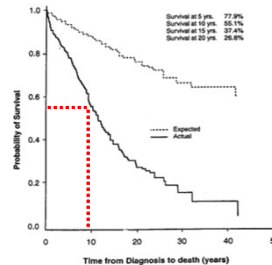
Systemic Sclerosis



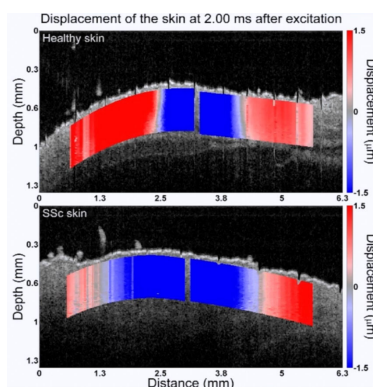
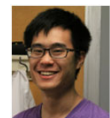
Systemic sclerosis (SSc) is a systemic connective tissue disease. Characteristics of systemic sclerosis include essential vasomotor disturbances; fibrosis; subsequent atrophy of the skin, subcutaneous tissue, muscles, and internal organs (eg, alimentary tract, lungs, heart, kidney, CNS); and immunologic disturbances accompany these findings.

Traditional systemic sclerosis examination

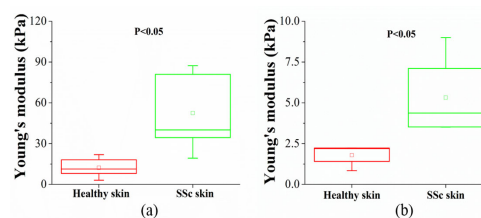
- 10-year survival rate: 58%
- Disease feature of SSc: Skin thickening
- Modified Rodnan score system (mRSS)
 - Limited sensitivity to subtle changes of the skin
 - Inter- and intra- observer variability (ICC:~0.76)
- Ultrasound imaging (US)
 - Dermal thickness detection
 - Acoustic radiation force impulse (ARFI)
 - Elastography assessment (Shear wave)
 - But, insufficient resolution for Imaging
 - Epidermal and dermal structures
 - especially dermal-epidermal junction.



Systemic Sclerosis – mice study



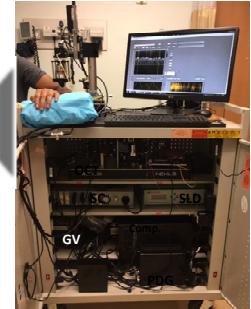
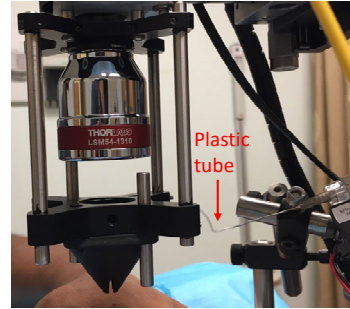
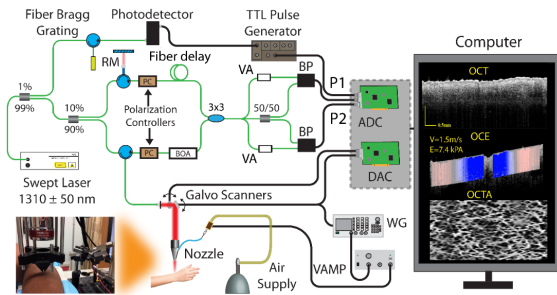
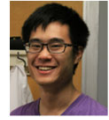
Elastic wave propagation in skin, as assessed by OCE in a typical healthy (top) and SSc (bottom) skin sample, measured at 2 ms after excitation in vivo



In vivo and in vitro OCE-assessment of skin involvement in the peripheral regions of SSc-affected skin. Plotted are the Young's modulus recorded in healthy skin and BLM-SSc afflicted skin at the periphery of the diseased region, from (a) in vitro and (b) in vivo OCE measurements (N = 4-9 mice per study group).

Du et al: JBO, 2016

Clinical OCE detection for SSc patients

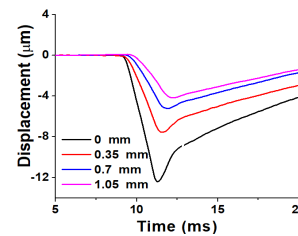
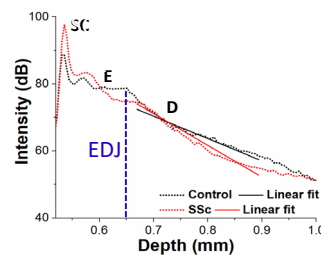
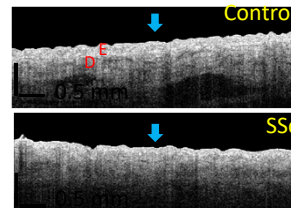
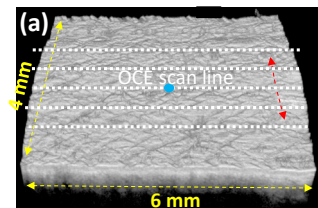
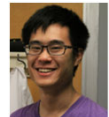


- Spectral domain OCT: 1310 ± 45 nm
- Line rate: 70k Hz
- Pixel resolution: 8 μ m in air
- Acquisition time: 3D OCT scan + 5 OCE scan
 - 15 min with 5 min break
- Site specific mRSS
- Skin biopsy



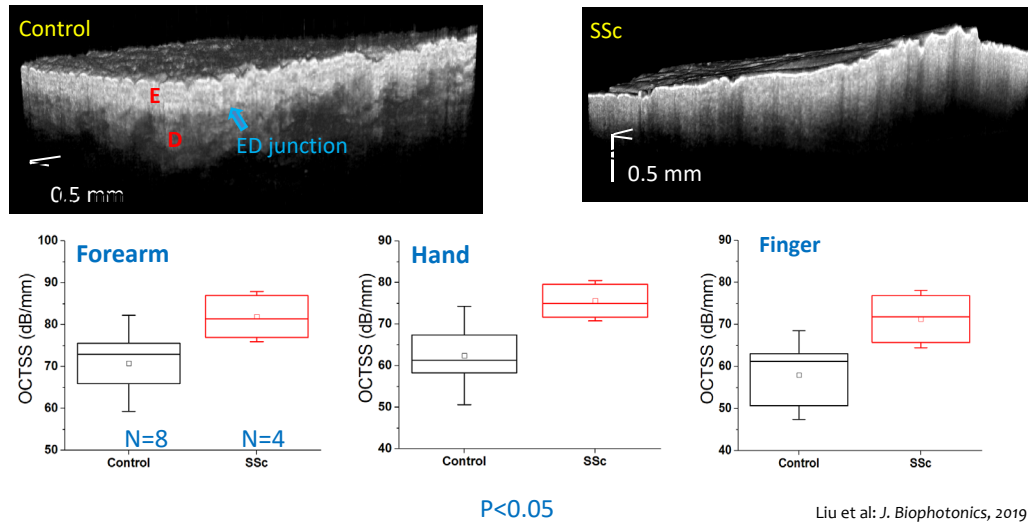
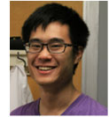
Liu et al: *J. Biophotonics*, (2019)

Experimental procedure

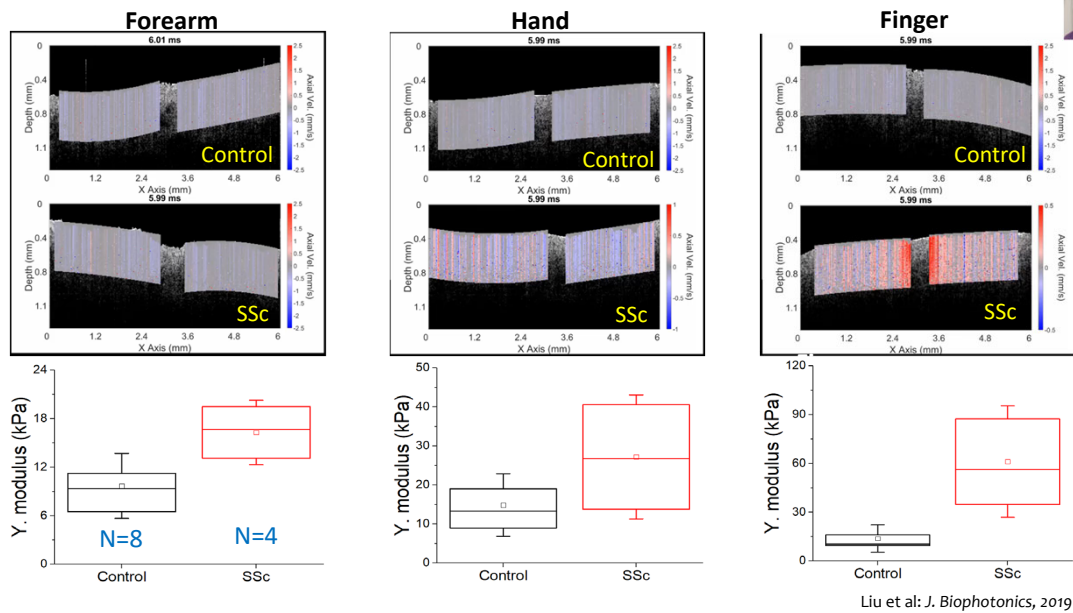
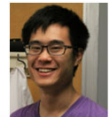


Liu et al: *J. Biophotonics*, 2019

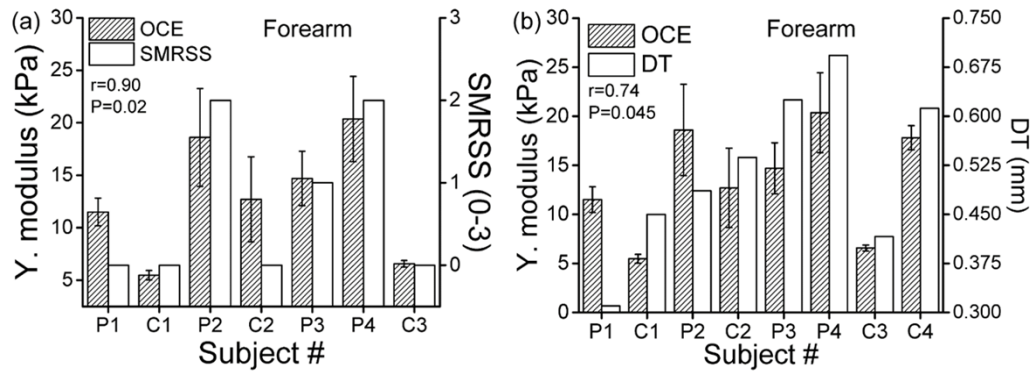
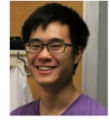
OCT analysis



OCE assessment



Clinical Summary



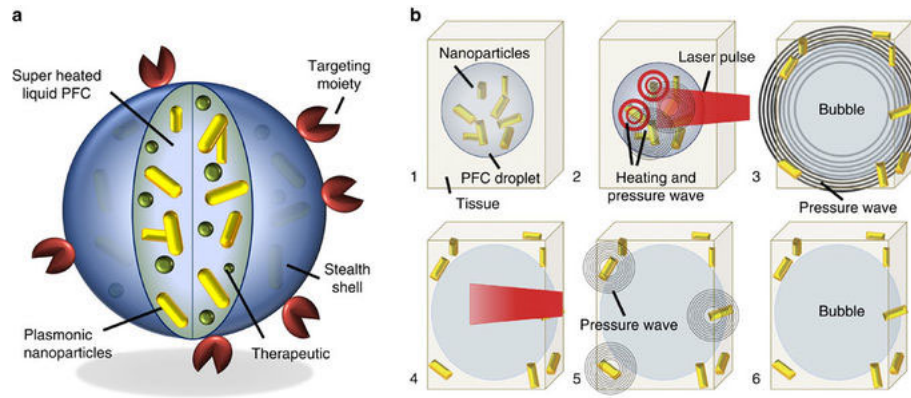
Liu et al: J. Biophotonics, 2019

“Nanobomb” Optical Coherence Elastography



Konstantin Sokolov, Ph.D.
Professor, Department of
Imaging Physics,
M.D. Anderson Cancer Center

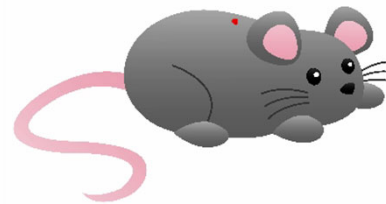
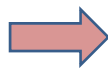
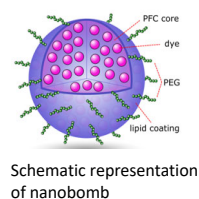
“Nanobombs”



Emelianov, S., et al, *Nature Comm*, 2012

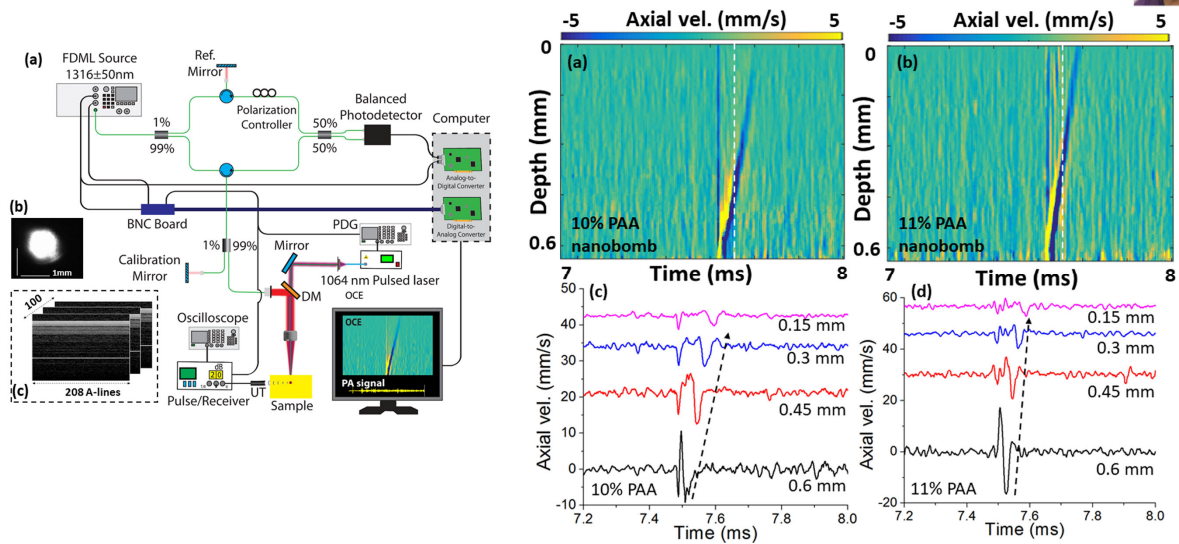
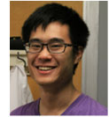
“Nanobomb” OCE

- Highly localized Targeting + High frequency response = nanobomb OCE
 - Dye-loaded perfluorocarbon (PFC) nanodroplets (“nanobombs”)
 - Excited by 532 nm pulsed laser
 - Produce highly localized stress by the nanobomb explosion.
- Detected by line-field low coherent holography system



C.-H. Liu et al., *Biomed Opt Express* 7 (2016)
C. H. Liu, et al., *Opt Lett* 43, 2006 (2018)

“Nanobomb” OCE



C. H. Liu, et al., Opt Lett (2019)

OCE: Summary and Conclusions

- OCE is a new emerging technique for tissue assessment: gives **biomechanical** contrast
- OCT structural images comes “for free”
- Allows 3D imaging and biomechanical assessment of various tissues
- Provides highly localized mechanical information of tissues with micrometer spatial resolution
- Holds a great promise for *in vivo* detection of various tissue diseases and quantitative evaluation of therapeutic interventions

Collaborators:

- Irina Larina – BCM
- Michael Twa – UH Optometry
- Salavat Aglyamov – UH
- Stas Emelianov – G. Tech
- Fabrice Manns – U. Miami
- Konstantin Sokolov - MDACC
- Shervin Assassi– UTH
- Chandra Mohan– UH
- Valery Tuchin – SSU
- Mary Dickinson – BCM
- Bruce Butler – UTHSC
- Michael Allon – HFC
- Xingde Li, JHU
- James Martin– BCM
- Rajesh Miranda– TAMU
- Mohamad Ghosn – Methodist
- Pradeep Sharma – UH
- Matthew Franchek – UH
- Richard Willson – UH
- Paul Ruchhoeft – UH
- Alexei Sobakin – UW, Madison
- Marlowe Eldridge – UW, Madison
- Raph Pollock – MDACC
- Giuliano Scarcelli, UMD
- Richard Finnell, BCM

Current Funding:

NIH	2R01EY022362 (Larin, Twa)	(2010-2020)
	1R01HD086765 (Larin, Miranda)	(2016-2020)
	1R01HL130804 (Larin, Martin)	(2016-2020)
	1R01HD095520 (Larin, Scarcelli, Finnell)	(2018-2023)
	9R01HD096335 (Larina)	(2013-2023)
	1R01EB027099 (Larina)	(2019-2022)
	1R21CA231561 (Sokolov)	(2019-2021)
	1R01HL146745 (Mayerich)	(2019-2024)

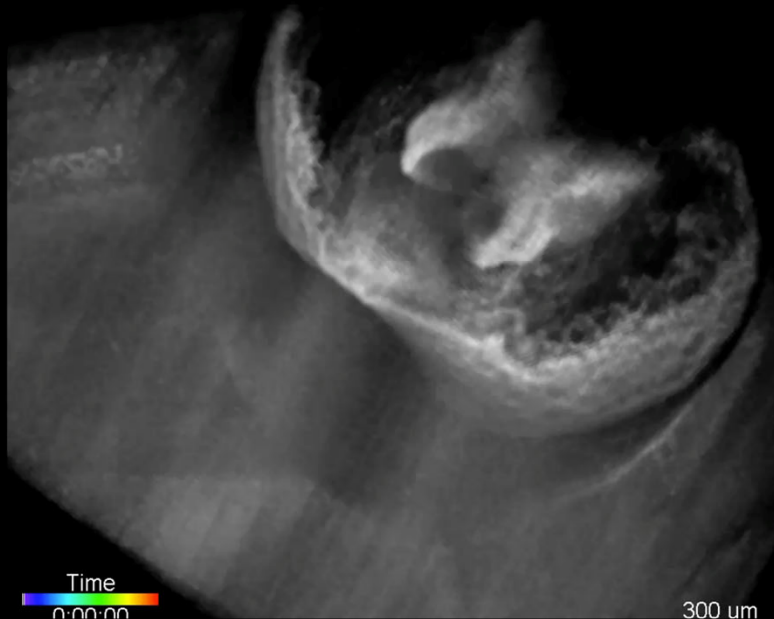
Graduate students and Postdoctoral positions are available:

Please inquire klarin@uh.edu

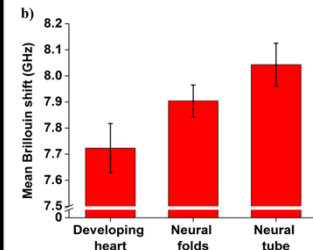
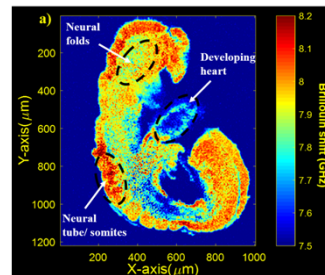
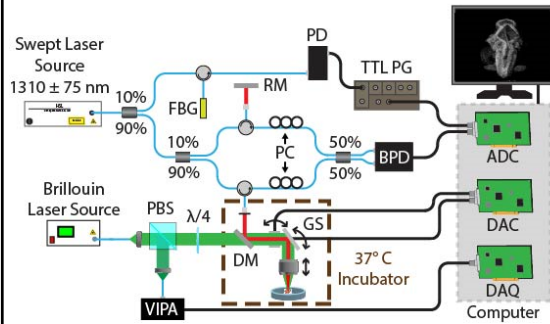
<http://bol.egr.uh.edu/>



Can we map mechanical forces during development?



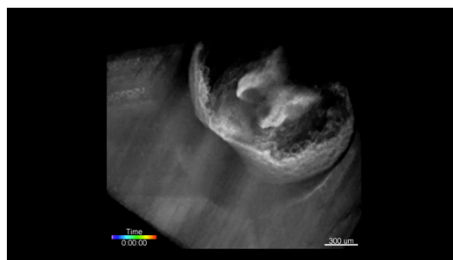
OCT + Brillouin Spectroscopy



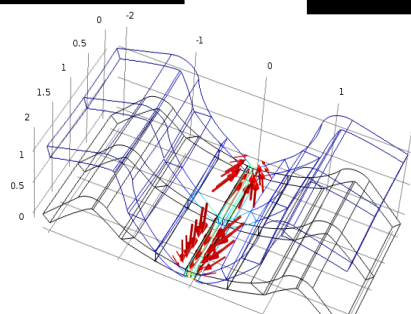
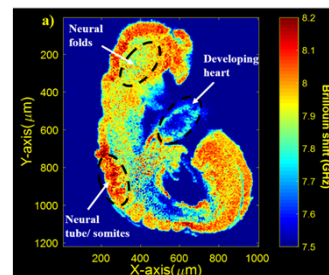
Raghunathan et al. J Biomed Opt. 2017
Zhang et al, Birth Defects Research, 2018

Future plans

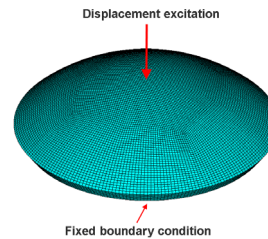
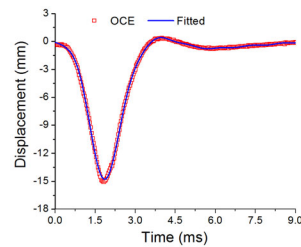
In collaboration with Scarcelli and Fennell group



+



Effect of Corneal Curvature/Geometry

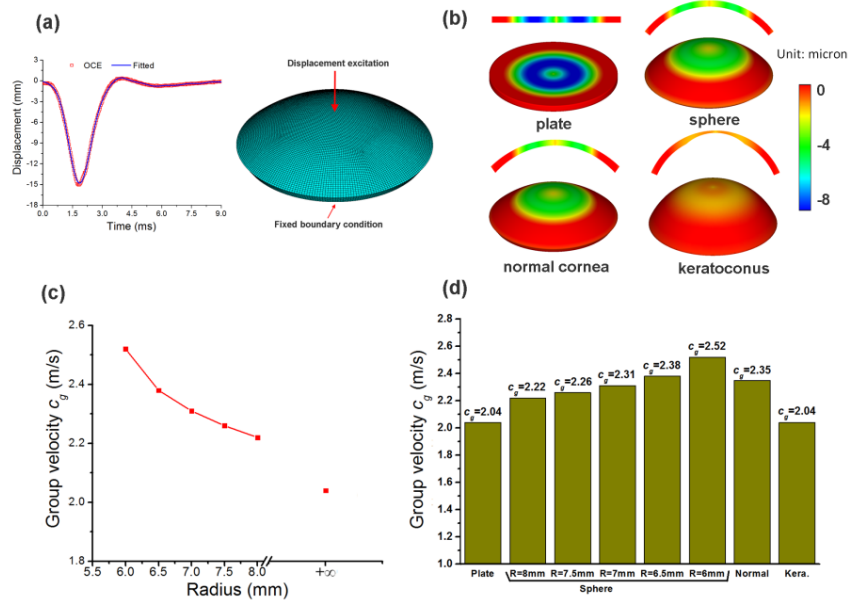


3D FEM modeling:

Young's modulus $E=60$ kPa, no viscosity
 No aqueous humor effect (will be considered later)
 Poisson's ratio $\nu=0.49$
 Density $\rho=1.06$ kg/m³

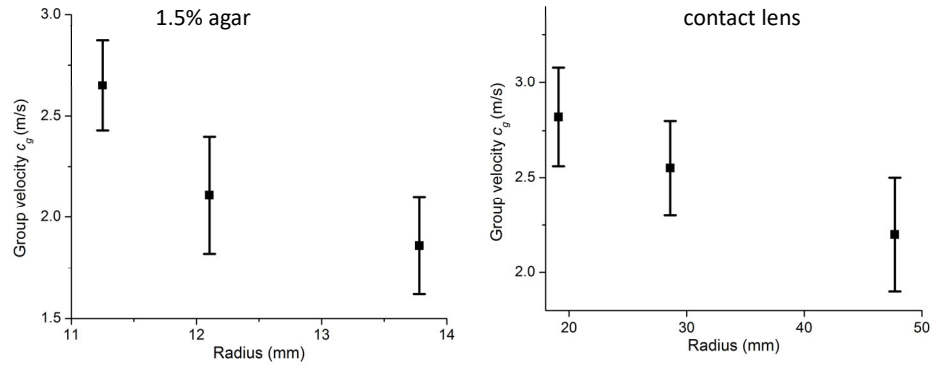
Han et al: Applied Phys Let , 2015

Effect of Corneal Curvature/Geometry



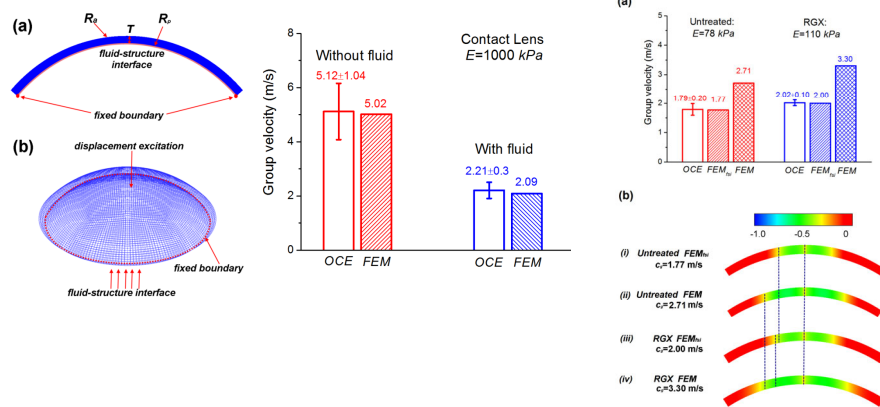
Han et al: Applied Phys Let , 2015

Effect of Corneal Curvature/Geometry: Experimental Data



Han et al: Applied Phys Let , 2015

Fluid Effect



Han et al: Applied Phys Let , 2016

Quantifying Viscoelasticity



Lamb waves: modified Rayleigh-Lamb frequency equation

Modified RLFE: $\det(M)=0$

$$M = \begin{bmatrix} (k^2 + \beta^2) \sinh(\alpha d) & 2k\beta \sinh(\beta d) & (k^2 + \beta^2) \cosh(\alpha d) & 2k\beta \cosh(\beta d) & 0 \\ 2k\alpha \cosh(\alpha d) & (k^2 + \beta^2) \cosh(\beta d) & 2k\alpha \sinh(\alpha d) & (k^2 + \beta^2) \sinh(\beta d) & 0 \\ -(k^2 + \beta^2) \sinh(\alpha d) & -2k\beta \sinh(\beta d) & (k^2 + \beta^2) \cosh(\alpha d) & 2k\beta \cosh(\beta d) & \frac{\rho_F \omega^2}{\mu_D} \\ 2k\alpha \cosh(\alpha d) & (k^2 + \beta^2) \cosh(\beta d) & -2k\alpha \sinh(\alpha d) & -(k^2 + \beta^2) \sinh(\beta d) & 0 \\ \alpha \cosh(\alpha d) & k \cosh(\beta d) & -\alpha \sinh(\alpha d) & -k \sinh(\beta d) & -\alpha_F \end{bmatrix}$$

$$k^2 = \frac{\omega^2}{c_p^2} \quad \alpha^2 = k^2 - \frac{\omega^2}{c_1^2} \quad \beta^2 = k^2 - \frac{\omega^2}{c_2^2}$$

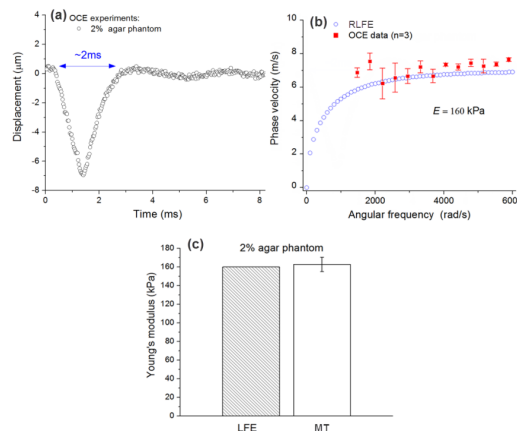
$$\mu_D = \mu + i\omega\eta \quad \alpha_F^2 = k^2 - \frac{\omega^2}{c_F^2} \quad c_2 = \sqrt{\frac{\mu_D}{\rho}}$$

ω - Circular frequency
 c_p - Phase velocity
 ρ - Density
 μ - Shear elastic modulus
 η - Shear viscous modulus

c_1 - Speed of sound
 $2d$ - Thickness
 c_F - Speed of sound in fluid
 ρ_F - Density of fluid

Han et al: PMB, v. 60, 2015

Quantifying Viscoelasticity (phantoms)

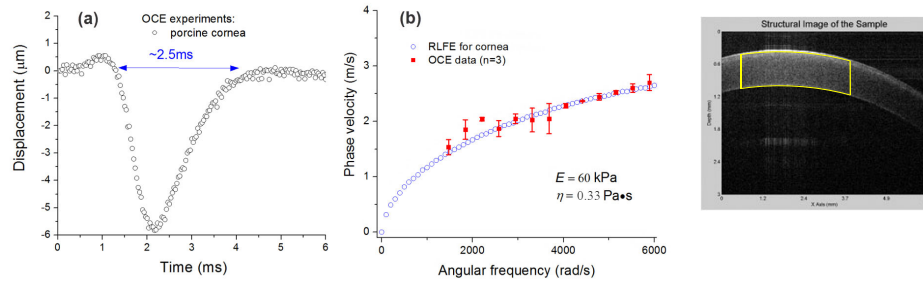


- (a) Temporal displacement profile from 2% agar phantom
 (b) OCE vs RLFE for 2% agar phantom
 (c) Young's modulus of 2% agar phantom assessed by RLFE and mechanical testing (MT).

The Young's modulus of 2% phantom measured by OCE and fitted by RLFE is 160 kPa, which was validated by uniaxial mechanical testing.

Han et al: PMB, v. 60, 2015

Quantifying Viscoelasticity (cornea)



(a) Temporal displacement profile from a porcine cornea

(b) OCE vs modified RLFE for porcine cornea ($n=3$)

For porcine cornea at IOP=20 mmHg, the Young's modulus was estimated as 60 kPa and shear viscosity as $0.33\text{ Pa}\cdot\text{s}$.

Han et al: *PMB*, v. 60, 2015
Han et al: *JMBBM*, v. 66, 2017